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# Diversity of *Eimeria* species in backyard poultry of subtropical hilly region of Meghalaya, India

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#### Abstract

The aim of the present study was to determine the prevalence and diversity of *Eimeria* species in the backyard poultry of hilly region of Meghalaya. A total of 674 nos. of fecal samples of poultry were collected from different age groups *viz.* < 8 weeks (132 nos.), 8-28 weeks (290 nos.) and > 28 weeks (252 nos.) and examined for detection of *Eimeria* oocysts by flotation techniques. Sporulation of the oocyst was done in 2.5% potassium dichromate solution for identification of the *Eimeria* species. The overall prevalence of *Eimeria* sp. in the backyard poultry was 30.12%. Eight species of *Eimeria viz. E. tenella* (24.63%), *E. necatrix* (10.84%), *E. maxima* (0.98%), *E. mitis* (1.48%), *E. brunetti* (1.97%), *E. praecox* (1.48%), *E. mivati* (0.98%) and *E. acervulina* (2.96%) were recorded. Mixed infections were recorded in 54.68% birds. Highest and lowest infections was recorded in the month of May (40.29%) and December (15.15%), respectively. Season wise, highest infection recorded during monsoon (33.87%) followed by spring (32.77%), winter (27.78%) and autumn (18.37%). Age wise, 68.18%, 25.86% and 15.08% infections were recorded in < 8, 8-28 and > 28 weeks old birds, respectively.

Keywords: diversity, Eimeria sp., backyard poultry, Meghalaya

## 1. Introduction

Animal husbandry is an important subsector of Indian agriculture and backyard poultry farming is one of the important components among the tribal farmers of Meghalaya. Backyard poultry farming is increasing rapidly due to low establishment cost, cheap source of proteins and employment <sup>[1, 2]</sup>. The total poultry population of the country is 851.81 million (backyard poultry: 317.07 million; commercial poultry: 534.74 million), in which Meghalaya's contribution is 0.63% (5.38 million) <sup>[3]</sup>.

Poultry coccidiosis is an intracellular intestinal parasitic disease caused by the different species of Eimeria. Morbidity of coccidiosis is estimated to be 50-70% and the disease is a major threat to 15-50 day old birds <sup>[4]</sup>. It seriously impairs the growth and feed utilization of the infected birds resulting in the loss of productivity and inflicts tremendous economic losses to the poultry farmers. It causes intestinal tissue damage and interferes with the food digestion and absorption resulting in the weight loss and bloody droppings. Sometime high rate of mortality may be observed in a farm. Secondary bacterial infection with Clostridium *perfringens* may occur which may predispose them to other gut infections such as necrotic enteritis. About nine species of *Eimeria* have been recognized in poultry birds, of which E. brunetti, E. maxima, E. necatrix and E. tenella are the most pathogenic; E. acervulina, E. mitis, E. mivati are the less pathogenic while E. praecox and E. hagani are the lesser pathogenic <sup>[5, 6]</sup>. The prevalence of coccidiosis in poultry was reported in many countries such as Iran <sup>[7]</sup>, Egypt <sup>[8]</sup>, Ethiopia <sup>[9]</sup>, South Africa <sup>[10]</sup>, Nepal <sup>[11]</sup>, Korea <sup>[12]</sup> and Nigeria <sup>[13]</sup>. Both clinical and sub-clinical coccidiosis retards the growth of flocks and cause huge economic loss to the farmers <sup>[14]</sup>, thus it is important to know the prevalence of different species of *Eimeria* in a particular geographical region. Though there are reports on the prevalence of different species of *Eimeria* in the poultry from different states of India like Assam<sup>[15]</sup>, Uttar Pradesh and Uttarakhand <sup>[16]</sup>, Jammu <sup>[17]</sup>, Karnataka <sup>[18]</sup>, Tamil Nadu <sup>[19]</sup> but there is paucity of information on different species of *Eimeria* in backyard poultry from Meghalaya. Thus, the present study has been undertaken to find out the diversity of *Eimeria* sp. in the backyard poultry of subtropical hilly region of Meghalaya.

# 2. Materials and Methods

2.1 Study area: The present study was conducted in the Ri-Bhoi district of Meghalaya which

Corresponding Author: M Das Senior Scientist, Division of Animal Health, ICAR Research Complex for NEH Region, Umiam, Meghalaya, India lies between  $25^{\circ}15'$  and  $26^{\circ}15'$  North latitudes and  $91^{\circ}45'$  and  $92^{\circ}15'$  East longitudes. The district is characterized by rugged and irregular land surface and includes a series of hill ranges.

**2.2 Study period:** The study was conducted for two years (2018-19, 2019-20) and divided into four seasons, *viz.* Spring (March, April), Monsoon (May, June, July, August, September), Autumn (October, November) and Winter (December, January, February).

**2.3 Sample collection:** Freshly voided fecal samples were collected from different locations *viz*. Umiam, Sarikuchi, Umthan, Umsawkhwan, Mawphrew, Nalapara, Borkhatsari, Purangang, Borgang and Lalumpam in the marked plastic pouch/vials. All the birds were categorized according to the age *viz*. < 8 weeks, 8-28 weeks and >28 weeks. A total of 674 nos. of fecal samples of poultry were collected from different age groups *viz*. < 8 weeks (132 nos.), 8-28 weeks (290 nos.) and > 28 weeks (252 nos.).

**2.4 Parasitological techniques:** To detect *Eimeria* oocysts in the fecal samples of backyard poultry, samples were examined by direct flotation technique using saturated salt (sp.gr. 1.20) and sucrose (sp.gr. 1.27) solution <sup>[20]</sup>. Positive samples were then quantified to estimate the oocyst per gram (OPG) of feces by using modified McMaster technique <sup>[21]</sup>. Samples not being examined on the same day were preserved at refrigerated temperature (4°C) for next day examination.

Sporulation of the oocyst was done by mixing positive samples having *Eimeria* oocysts with 2.5% potassium dichromate solution in a ratio of 1:5 volumes as per the standard procedure <sup>[21, 22]</sup>. Morphological characterization of the oocysts <sup>[20]</sup> was done by using an Olympus BX51 light microscope at 100x and 400x magnifications.

# 3. Results and Discussion

The overall prevalence of coccidiosis in backyard poultry of Meghalaya was 30.12% (Table 1). Eight species of Eimeria viz. E. tenella (24.63%), E. necatrix (10.84%), E. maxima (0.98%), E. mitis (1.48%), E. brunetti (1.97%), E. praecox (1.48%), E. mivati (0.98%) and E. acervulina (2.96%) were identified by morphological characterization (Fig. 1). Mixed infections were recorded in 54.68% birds. Month wise, highest and lowest infection was recorded in the month of May (40.29%) and December (15.15%), respectively (Fig. 2). Oocyst per gram (OPG) was recorded highest and lowest in the month of August (30000) and February (9500), respectively. Season wise, highest infection recorded during monsoon (33.87%) followed by spring (32.77%), winter (27.78%) and autumn (18.37%) (Table 1, Fig.3). E. tenella, E. necatrix and mixed infections were observed throughout the year (Fig. 2). However, in monsoon season E. praecox, E. maxima, E. mitis, E. brunetti and E. acervulina are also observed; E. brunetti in spring; E. maxima, E. brunetti, E. praecox, E. mivati and E. acervulina in winter; E. mitis, E. praecox and E. acervulina in autumn.

Table 1: Seasonal prevalence of Eimeria sp. in backyard poultry of hilly region of Meghalaya

Season	Sample examined	Sample positive	E. tenella	E. necatrix	E. maxima	E. mitis	E. brunetti	E. praecox	E. mivati	E. acervulina	Mixed Infection	OPG
Spring	119	39 (32.77)	10(25.64)	5 (12.82)	-	-	1 (2.56)	-	-	-	23 (58.97)	1162.50
Monsoon	313	106(33.87)	25(23.58)	12(11.32)	1(0.94)	2(1.89)	1 (0.94)	1 (0.94)	-	2 (1.89)	62 (58.49)	23750
Autumn	98	18 (18.37)	6 (33.33)	1 (5.56)	-	1(5.56)	-	1(5.56)	-	1(5.55)	8(44.44)	23250
Winter	144	40 (27.78)	9 (22.5)	4 (10)	1 (2.5)	-	2 (5)	1 (2.5)	2 (5)	3 (7.5)	18 (45)	11866.66
Total	674	203 (30.12)	50(24.63)	22(10.84)	2(0.98)	3(1.48)	4(1.97)	3(1.48)	2(0.98)	6(2.96)	111(54.68)	18550

Figures in parentheses indicates percent positivity



Fig 1: Different species of Eimeria in backyard poultry of Meghalaya



Fig 2: Month wise Intensity of Eimeria infection in poultry of Meghalaya



Fig 3: Season wise prevalence of different species of Eimeria infection in poultry

In the present study, different species of Eimeria in poultry has been observed which was in agreement with the findings of Adhikari et al. [11], Sharma et al. [17] and Grabi et al. [23]. Mixed infections of Eimeria sp. was observed in 54.68% birds. Kaboudi et al. [24] and Molla and Ali [25] reported 26.55% and 45.65% mixed infections in the poultry birds from Tunisia and Ethiopia, respectively. The difference in the percent prevalence from the present study may be due to different geographical locations.

In the present study, highest and lowest infections were recorded in the month of May and December, respectively. Similarly, Naveed and Faryal <sup>[14]</sup> reported highest numbers of cases in August (90.6%) and least in January (63%). They also observed that the type of litter materials, time duration, season, humidity, temperature and disposal of dead birds are other critical factors for poultry coccidiosis. High rate of infection in the monsoon season may be due to wet floor and litter of poultry farm which is very conducive for the growth and development of the Eimeria oocysts. E. tenella, E. brunetti and E. necatrix are associated with haemorrhagic coccidiosis and can be highly pathogenic, with high mortality and morbidity <sup>[26]</sup>. However, E. acervulina, E. maxima, E. mitis and E. praecox are less pathogenic, incurring malabsorptive pathologies, although morbidity and mortality

can occur depending on dose ingested, parasite strain-specific variation in virulence and host factors such as age, breed and immune status [27]. According to the Williams et al.[28] coinfection with multiple species is common and can complicate diagnosis. E. necatrix has been recognized as the most pathogenic Eimeria sp. but E. tenella is more common and exerts a greater impact on poultry production <sup>[29]</sup>. The occurrence of coccidiosis can also vary due to climatic conditions, with evidence of elevated parasite levels and disease during wetter and warmer seasons [30,31,32].

Age wise 68.18%, 25.86% and 15.08% infections were recorded in < 8, 8-28 and > 28 weeks old birds, respectively (Table 2). In poultry birds of 8-28 weeks, all eight species of Eimeria were prevalent viz. E. tenella (21.33%), E. necatrix (14.67%), E. maxima (1.33%), E. mitis (4%), E. brunetti (5.33%), E. praecox (2.67%), E. mivati (2.67%) and E. acervulina (5.33%). In birds of < 8 weeks, E. tenella (31.11%), E. necatrix (8.89%), E. maxima (1.11%), E. praecox (1.11%) and E. acervulina (2.22%) were recorded. However, in birds of > 28 weeks only E. tenella (15.79%) and E. necatrix (7.89%) were recorded. Percent prevalence of mixed infections was 55.56%, 42.67% and 76.32% in < 8, 8-28 and > 28 weeks old birds, respectively.

Age group	Sample examined	Sample positive	E. tenella	E. necatrix	E. acervulina	E. praecox	E. mivati	E. mitis	E. maxima	E. brunetti	Mixed infection
< 8	132	90 (68.18)	28 (31.11)	8 (8.89)	2 (2.22)	1 (1.11)	-	-	1 (1.11)	-	50 (55.56)
8-28	290	75 (25.86)	16(21.33)	11(14.67)	4(5.33)	2(2.67)	2(2.67)	3(4)	1(1.33)	4(5.33)	32(42.67)
> 28	252	38 (15.08)	6 (15.79)	3 (7.89)	-	-	-	-	-	-	29 (76.32)
Total	674	203(30.12)	50(24.63)	22(10.84)	6(2.96)	3(1.48)	2(0.98)	3(1.48)	2(0.98)	4(1.97)	111(54.68)

Table 2: Age wise prevalence of Eimeria sp. in backyard poultry of hilly region of Meghalaya

Figures in parentheses indicates percent positivity

In the present study, age wise variation in the prevalence of *Eimeria* infection in backyard poultry was observed and younger groups are more susceptible to infection. This is in congruence with the findings of Jemimah *et al.* <sup>[13]</sup>, Wondimu *et al.* <sup>[33]</sup>, Prakashbabu *et al.* <sup>[34]</sup> and Lawal *et al.* <sup>[35]</sup>. Williams <sup>[36]</sup> also reported that young birds are more susceptible to *Eimeria* sp. According to Long *et al.* <sup>[37]</sup> and Williams <sup>[38]</sup>, excretion of oocysts has also been reported to be higher in birds between 4 and 8 weeks of age.

According to Omer *et al.* <sup>[39]</sup>, all ages of birds are susceptible to coccidiosis, but younger birds are more susceptible to infection than older birds which might be associated with the immature immune system in young birds leaving them susceptible to infection even with the lower or less pathogenic strain of *Eimeria* species. Das *et al.* <sup>[40]</sup> also observed that high rate of infection in young birds may be due to decreased immunity as well as continuous exposure to infections from the contaminated litter. According to Morris and Gasser <sup>[41]</sup>, *Eimeria* sp. multiply in the intestinal tract, causing tissue damage, interruption in feeding and digestive processes as well as nutrient absorption, blood loss and increased susceptibility to other disease agents.

The combinations of different Eimeria species and the intensity of infection vary considerably, both locally and globally <sup>[42, 43]</sup>. Prakashbabu *et al.* <sup>[34]</sup> also observed that occurrence of Eimeria species varies between geographic regions and poultry production systems and may also influence its genetic diversity. High incidence of coccidiosis is usually observed in poultry managed under intensive management system like deep litter due to increased likelihood of high oocysts accumulation in the litters <sup>[44, 45]</sup>. Factors which contribute to the outbreaks of clinical coccidiosis include litter moisture exceeding 30%, immune suppression, suboptimal inclusion of anticoccidials in feed and environmental and managemental stress <sup>[46]</sup>. Thus, the present study has significance because eight species of Eimeria were recorded for the first time in the different age groups of backyard poultry in the subtropical hilly region of Meghalaya. So, to make poultry farming profitable in this region it is necessary to implement preventive and control measures strictly against poultry coccidiosis.

## 4. Conclusion

The present study revealed that eight species of *Eimeria* are prevalent in the subtropical hilly region of Meghalaya and mostly prevalent during the monsoon season. Younger age groups are highly susceptible to the infections. Regular screening and use of anticoccidial drug is necessary for profitable backyard poultry farming.

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## 6. References

- 1. Frantovo D. Some parasitic nematodes (Nematoda) of birds (Aves) in the Czech Republic. Acta Societatis Zoologicae Bohemicae 2000;66(1):13-28.
- Bachaya HA, Raza MA, Khan MN, Iqbal Z, Abbas RZ, Murtaza S. Predominance and detection of different *Eimeria* species causing coccidiosis in layer chickens. Journal of Animal and Plant Sciences 2012;22:597-600.
- 3. Livestock Census. 20<sup>th</sup> Livestock Census. Department of Animal Husbandry, Dairying and Fisheries, Ministry of Agriculture, Government of India 2019.
- 4. Wang M. Veterinary Parasitology. Beijing: China Agricultural Press 2003.
- 5. Jadhav BN, Nikam SV, Bhamre SN, Jaid EL. Study of *Eimeria necatrix* in broiler chicken from Aurangabad District of Maharashtra State India. International Multidisciplinary Research Journal 2011;1(11):11-12.
- 6. Nematollahi A, Moghaddam GH, Niyazpour F. Prevalence of *Eimeria* spp. among broiler chicks in Tabriz (Northwest of Iran). Research Journal of Poultry Science 2008;2:72-74.
- 7. Hadipour MM, Olyaie A, Naderi M, Azad F, Nekouie O. Prevalence of Eimeria species in scavenging native chickens of Shiraz, Iran. African Journal of Microbiology Research 2011;5:3296-3299.
- Ahmed AA, Olfat AM, Aida ANE, Mohamed SA. Studies on coccidia of Egyptian Balady breed chickens. Life Science Journal 2012;9(3):568-576.
- 9. Belaynew A, Wudu T, Mengestie A, Ayalew N, Kassa D, Mebrie Z. Study of the prevalence, species identification and risk factors associated with poultry coccidiosis in Gondar Town, North Ethiopia. Nature and Science 2016;14(7):119-124.
- Mwale M, Masika P. Point prevalence study of gastrointestinal parasites in village chickens of Centane district, South Africa. African Journal of Agricultural Research 2011;6:2033-2038.
- 11. Adhikari A, Gupta R, Pant GR. Prevalence and identification of coccidian parasite (*Eimeria* spp.) in layer chicken of Ratnanagar municipality, Chitwan district, Nepal. Journal of Natural History Museum 2008;23:45-50.
- 12. You MJ. The comparative analysis of infection pattern and oocyst output in *Eimeria tenella, E. maxima* and *E. acervulina* in young broiler chicken. Veterinary World 2014;7(7):542-547.
- Jemimah A, James TI, Abba E, Rejoice A, Lamogo Y. Prevalence and Associated Risk Factors of Coccidia Infection among Desi and Broiler Chickens in Gombe Metropolis, Gombe State, Nigeria. South Asian Journal of Parasitology 2020;4(1):40-46.
- Naveed Q, Faryal R. Risk factors and prevalence of coccidiosis in chicken in district Gujarat, Punjab, Pakistan. International Journal of Biosciences 2019;15(3):66-79.

- 15. Kalita A, Sarmah PC, Borah MK, Hussain L, Bhattacharjee K. Magnitude of Coccidia Infection in Small Scale Broiler Chicken Farms of Rural Assam (India). International Journal of Current Microbiology and Applied Science 2018;7(10):3399-3403.
- Kumar S, Garg R, Ram H, Maurya PS, Banerjee PS. Gastrointestinal parasitic infections in chickens of upper Gangetic plains of India with special reference to poultry coccidiosis. Journal of Parasitic Disease 2015;39(1):22-26.
- 17. Sharma S, Iqbal A, Azmi S, Mushtag I, Wani AZ, Ahmad S. Prevalence of poultry coccidiosis in Jammu region of Jammu and Kashmir State. Journal of Parasitic Disease 2015;39(1):85-89.
- 18. Puttalakshmamma GC, Ananda KJ, Prathiush PR, Mamatha GS, Rao S. Prevalence of Gastrointestinal parasites of Poultry in and around Banglore. Veterinary World 2008;1(7):201-202.
- Aarthi S, Raj GD, Raman M, Gomathinayagam S, Kumanan K. Molecular prevalence and preponderance of *Eimeria* spp. among chickens in Tamil Nadu, India. Parasitology Research 2010;107:1013-1017.
- Soulsby EJ. Helminths, Arthropods and Protozoan's of Domesticated Animals. 7<sup>th</sup> ed. Bailliere Tindall, London. 1982.
- 21. MAFF. Ministry of Agriculture, Fisheries and Food. Manual of veterinary parasitological techniques, Her Majesty's Stationery Office, London 1986.
- 22. Sloss MW, Kemp RL, Zajac AM. Veterinary Clinical Parasitology. 6<sup>th</sup> Edition. Iowa State University Press, Ames, Iowa. 1994.
- 23. Grabi F, Tesfaye A, Mezene W. Study on prevalence of poultry coccidiosis in Nekemte town, East Wollega, Ethiopia. African Journal of Agricultural Research 2015;10(5):328-333.
- 24. Kaboudi K, Umar S, Munir T. Prevalence of Coccidiosis in Free-Range Chicken in Sidi Thabet, Tunisia. Scientifica. 2016.
  - http://dx.doi.org/10.1155/2016/7075195.
- 25. Molla B, Ali A. Epidemiological study on poultry coccidiosis: Prevalence, species identification and post mortem lesions in grower chicken in Kombolcha, North-Eastern Ethiopia. Journal of Veterinary Medicine and Animal Health 2015;7(1):1-8.
- 26. Long P, Joyner L, Millard B, Norton C. A guide to laboratory techniques used in the study and diagnosis of avian coccidiosis. Folia Veterinaria Latina 1976;6:201-217.
- Williams RB, Marshall RN, Pages M, Dardi M, Cacho E. Pathogenesis of *Eimeria praecox* in chickens: virulence of field strains compared with laboratory strains of *E. praecox* and *E. acervulina*. Avian Pathology. 2009;38:359-366.
- 28. Williams RB, Bushell AC, Reperant JM, Doy TG, Morgan JH, Shirley MW. A survey of *Eimeria* species in commercially-reared chickens in France during 1994. Avian Pathology 1996;25:113-130.
- Blake DP, Clark EL, Macdonald SE, Thenmozhi V, Kundu K, Garg R *et al.* Population, genetic, and antigenic diversity of the apicomplexan *Eimeria tenella* and their relevance to vaccine development. 2015. https://doi.org/10.1073/pnas.1506468112.
- 30. Awais MM, Akhtar M, Iqbal Z, Muhammad F, Anwar MI. Seasonal prevalence of coccidiosis in industrial

broiler chickens in Faisalabad, Punjab, Pakistan. Tropical Animal Health and Production 2012;44:323-328.

- 31. Jatau ID, Sulaiman NH, Musa IW, Lawal AI, Okubanjo Isah OO, Magaji Y. Prevalence of Coccidia infection and preponderance *Eimeria* species in free range indigenous and intensively managed exotic chickens during hot-wet season, in Zaria, Nigeria. Asian Journal of Poultry Science 2012;6:79-88.
- 32. Luu L, Bettridge J, Christley RM, Melese K, Blake D, Dessie T *et al*, Collins M, Lynch SE. Prevalence and molecular characterisation of *Eimeria* species in Ethiopian village chickens. BMC Veterinary Research 2013;9:208.
- Wondimu A, Mesfin E, Bayu Y. Prevalence of Poultry Coccidiosis and Associated Risk Factors in Intensive Farming System of Gondar Town, Ethiopia. Veterinary Medicine International 2019. Article ID 5748690, doi.org/10.1155/2019/5748690.
- 34. Prakashbabu BC, Thenmozhi V, Limon G, Kundu K, Kumar S, Garg R *et al. Eimeria* sp. occurrence varies between geographic regions and poultry production systems and may influence parasite genetic diversity. Veterinary Parasitology 2017;233:62-72.
- 35. Lawal JR, Jajere SM, Ibrahim UI, Geidam YA, Gulani IA, Musa G *et al.* Prevalence of coccidiosis among village and exotic breed of chickens in Maiduguri, Nigeria. Veterinary World 2016;9(6):653-659.
- 36. Williams RB. Epidemiological studies of coccidiosis in the domestic fowl (Gallus gallus). II. Physical condition and survival of *Eimeria acervulina* oocysts in poultry house litter. Applied Parasitology 1995;36:90-96.
- Long PL, Tompkins RV, Millard BJ. Coccodiosis in broilers: evaluation of infection by the examination of broiler house litter for oocysts. Avian Pathol 1975;4(4):287-294.
- Williams R. Epidemiological aspects of the use of live anticoccidial vaccines for chickens. International Journal of Parasitology 1998;28:1089-1098.
- 39. Omer SA, Apio A, Wronski T, Mohammad OB. A new coccidian parasite (*Eimeria farasanii* n. sp.) indicates parasite-host specificity in endemic Farasan gazelle. International Journal of Zoological Research 2011;7:85-92.
- 40. Das M, Laha R, Doley S. Gastrointestinal parasites in backyard poultry of subtropical hilly region of Meghalaya. Journal of Entomology and Zoology Studies 2020;8(5):1301-1305.
- Morris GM, Gasser RB. Biotechnological advances in the diagnosis of avian coccidiosis and the analysis of genetic variation in *Eimeria*. Biotechnology Advances 2006;24(6):590-603.
- 42. Haug A, Gjevre AG, Thebo P, Mattsson JG, Kaldhusdal M. Coccidial infections in commercial broilers: Epidemiological aspects and comparison of *Eimeria* species identification by morphometric and polymerase chain reaction techniques. Avian Pathology 2008;37:161-170.
- 43. Amer MM, Awaad MHH, El-Khateeb RM, Abu-Elezz NMT, Sherein-Said A, Ghetas MM *et al.* Isolation and identification of *Eimeria* from field coccidiosis in chickens. Journal of American Science 2010;6(10):1107-1114.
- 44. Dakpogan HB, Salifou S. Coccidiosis prevalence and intensity in litter based high stocking density layer

rearing system of Benin. Journal of Animal and Plant Sciences 2013;17(2):2522-2526.

- 45. Nnadi PA, George SO. A cross-sectional survey on parasites of chickens in selected villages in the sub humid zones of South-Eastern Nigeria. Journal of Parasitology Research 2010;141:1-6.
- 46. Singla LD, Pangasa A, Juyal PD. Caecal coccidiosis: efficacy of ayurvedic and allopathic coccidiostats in immunomodulated broiler chicks. Proceedings of the 12<sup>th</sup> International Conference of the Association of Institutions of Tropical Veterinary Medicine held from August, 2007 at Montpellier, France 2007, 19-22.