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## Biology and foraging behaviour of *Oenopiakirbyi* Mulsant (Coleoptera: Coccinellidae) on melon aphid, *Aphis gossypii* Glover

### D Kumari, SC Verma, PL Sharma, T Banshtu and Nidhi

#### Abstract

*Oenopia kirbyi* Mulsant is an important naturally occurring predator of *Aphis gossypii* Glover in cucumber. Before recommending the natural enemy against any pest it is important to investigate the feeding efficiency and density responsiveness of the given natural enemy. We, therefore, studied the biology and predatory potential of *O. kirbyi* against this aphid in the laboratory at  $25 \pm 0.5$  °C,  $70 \pm 5\%$  RH and 12L: 12D photoperiod. Incubation, total larval, pre pupal and pupal period of *O. kirbyi* was 3.40, 15.26, 1.30 and 4.80 days, respectively. Females on an average laid 177.60  $\pm$  6.38 eggs. Pre oviposition, oviposition and post oviposition period of *O. kirbyi* was 6.92, 23.22 and 5.48 days, respectively. Adult female and adult male consumed an average of 2634.78 and 2012.52 aphids in their life span of 38.50 and 31.38 days, respectively, whereas, total prey consumption during the larval stages was 535.90 aphids/ predator. All the predator stages exhibited a type II functional response to the aphid. Based on Rogers's random predator equation, the attack rate was lowest (0.065) in the first-instar and highest (0.171) in the adult stage. Handling times of the adult and the fourth-instar stages (0.237 and 0.263 h, respectively) were much shorter than other stages. Study indicates that the adults and the fourth-instar grubs were the most voracious stages and could be considered for the biological control of the aphid after field evaluation.

Keywords: biocontrol, attack rate, handling time, predator, functional response

#### Introduction

The melon aphid, Aphis gossypii Glover (Homoptera: Aphididae) is a cosmopolitan, polyphagous pest infesting 569 host plants in tropical, subtropical and temperate regions [32, 38, <sup>40]</sup>. Heavy infestation reduces the yield and fruit quality of cucumber. The aphids also secrete honeydew which encourages sooty mould that disturbs the normal physiology of the leaves and transfer 76 plant viruses <sup>[5, 28, 39, 2]</sup>. For the control of thisaphid, many insecticides have been recommended <sup>[22, 23]</sup> but, the indiscriminate use of synthetic pesticides leads to the development of insecticide resistance <sup>[41, 4, 34]</sup>, pest resurgence, insecticide residue, environmental pollution and killing of natural enemies. Melon aphid, A. gossypii developed resistance against many insecticides [8, 35, 25]. Insecticide resistance forces the farmers to increase the dose and frequency of pesticide application which further aggravates the problem <sup>[4]</sup>. Therefore, there is a need to find out some alternatives of chemical pesticides to protect cucumber from infestation of melon aphid <sup>[16]</sup>. Under such circumstances, biological control can be alternative, ecofriendly and sustainable approach to control this pest <sup>[14]</sup>. Coccinellid beetles are important predators of aphids and other hemipterans [27, 33, 1, 24, 17, 36]. But before the use of a predator in a biological control programme, the study of biology and foraging efficiency against the target species is necessary. Foraging efficiency of a given natural enemy can be assessed by studying its functional response to the target pest <sup>[7, 29]</sup>. Functional response is the number of prey eaten per predator as a function of prey density <sup>[13]</sup>. According to Holling (1959) functional response have been categorized into a linear relationship between prev density and the number of prey consumed (type I), a curvy linear relationship (type II), and a sigmoidal relationship (type III). In addition, dome shaped response (type IV) and negative exponential response (type V)  $^{[20, 31]}$  have also been reported. Functional response studies are used to estimate the attack rate (a) and handling time (Th) of the predator. The natural enemies with high attack rate(a) and low handling time (Th) are known as effective biocontrol agents [2, ]<sup>40]</sup>. The present study was, therefore, conducted with a aim to study the biology and foraging behaviour of O. kirbyi against A. gossypii so that it can be utilized in pest management

programme of the target pest.

#### Materials and Methods

The stock culture of *A. gossypii* was maintained on cucumber plants raised in the greenhouse. For the experimental studies aphid colonies were also maintained in the laboratory on cucumber seedlings at  $25 \pm 0.5$  °C,  $70 \pm 5\%$  RH and 12 L: 12D h photoperiod. The culture of *O. kirbyi* was maintained from field collected beetles on *A. gossypii* in the biological control laboratory at previously described environmental conditions. The beetle was reared for one generation before using for the experiment.

#### Developmental biology of O. kirbyi

Developmental biology of the O. kirbyi was studied on the melon aphid, A. gossypii at  $25 \pm 0.5$  °C,  $70\pm 5\%$  RH and 12 L: 12D h photoperiod. For this purpose, one pair of newly emerged adults was released in the Petriplate containing wet blotting paper on the base. Cucumber leaves (6×6cm) having mixed population of second and third instar of A. gossypii were placed on the wet blotting paper in the Petriplate. Each set was replicated 10 times. The food was changed daily and the Petriplates under observation were examined at 24 h interval for egg laying and moulting of the grubs until pupation. Pupae obtained were collected and kept for adult emergence. The duration of egg, each larval instar, total larval duration, pre pupal, pupal and egg to adult emergence was recorded. After emergence, the adults were sexed under a stereozoom microscope and kept in pairs in Petriplates with ample supply of food. Observations on pre-oviposition period, oviposition period, post-oviposition period, male longevity and female longevity were recorded.

#### Feeding potential of O. kirbyi

Feeding potential of different developmental stages of *O. kirbyi* was studied against *A. gossypii*. Newly emerged larvae of the desired stage were shifted individually to a petri plate with counted number of aphids of same instra. In order to select the aphids of the desired age, the cucumber leaves were carefully examined under the stereo zoom binocular microscope and the unwanted stages were removed carefully with the help of camel hair brush. The experiment was carried out in the insect culture room fixed at 12:12 h photoperiod, 25  $\pm$  0.5°C and 70 $\pm$  5 per cent relative humidity. The data on aphid consumption by the predator was recorded after every 24 h untilthey enter the next stage and the food was changed daily. The experiment was replicated ten times. The number of aphids consumed per day by each larval instar was calculated.

#### Foraging behaviour of O. kirbyi to A. gossypii

Functional response of first, second, third and fourth instar, and adult female of *O. kirbyi* was studied by offering varying densities of the second/third instar nymphs of the aphid on a cucumber leaf disc (6 cm diameter) placed in a Petri plate with moist filter paper. The prey densities offered were 10, 15, 20, 25 and from 10, 15, 30, 45, 60 and 75; 20, 40, 60, 80 and 100 and 25, 50, 75, 100 and 125 for the first, second, third and fourth instar as well the adult female of *O. kirbyi*, respectively. The prey densities for each stage were decided based on a preliminary experiment carried to study their feeding over a 24 h period. After allocating the prey at different densities, one predator was confined into each arena. The Petri plates were kept in a culture room at  $25 \pm 0.5$  °C, 70

 $\pm$  5% RH and 12 L: 12D h photoperiod. Each prey density was replicated 10 times simultaneously for each stage. The data on the number of prey eaten by each predator was recorded after 24 h.

#### Data analysis

Analysis of variance was performed to test differences in daily prey consumption at different prey densities and means were separated using the least significant difference. The functional response was analyzed from prey consumption data over a period of 24 h. The logistic regression was fit to the prey density offered (N) and the proportion of prey consumed (Na/N) to distinguish between Type II and Type III functional response was:

 $N_{a'} N = \exp(p_0 + p_1 N + p_2 N^2 + p_3 N^3) / (1 + \exp(p_0 + p_1 N + p_2 N^2 + p_3 N^3))$ 

Where, Na is the prey density eaten, N is the prey density offered, and  $p_0$ ,  $p_1$ ,  $p_2$  and  $p_3$  are constant, linear, quadratic and cubic coefficients. Significant negative  $p_1$  indicates Type II, whereas, a significant positive  $p_1$  and negative  $p_2$  indicates Type III response <sup>[15]</sup>. Because the cubic polynomial fit indicated the Type II response, further analysis was applied for Type II response. Rogers's random predator equation,  $N_a = N[1 - exp \{a(T_hN - T)\}]$ <sup>[30]</sup> was used to estimate the predator's attack rate (a) and predator's handling time (T<sub>h</sub>). Predator's attack rate per handling time (a/T<sub>h</sub>) and maximum theoretical predation rate per day (K = T/T<sub>h</sub>) was also calculated <sup>[6]</sup>. Analysis of variance was applied to test differences in a and T<sub>h</sub> values, and means were separated by least significant difference.

#### Results

#### **Developmental biology**

The mean duration of the egg, larval, prepupal and pupal stage was 3.4, 15.26, 1.4 and 1.3 days, respectively. The time taken from egg to adult emergence was 24.82 days. Female and male longevity of *O. kirbyi* reared on *A. gossypii* was 38.50 and 31.38 days, respectively. Pre-oviposition, oviposition and post-oviposition period was 6.92, 23.22 and 5.48 days, respectively. The female of *O. kirbyi* laid on an average 177.60 eggs/female.

#### **Feeding potential**

The number of aphids consumed increased with the advancement of developmental stages of the predator (Table 2). The average number of aphids consumed by the first, second, third and fourth instar of the predator was 32.40, 73.52, 171.06 and 252.80, respectively. Whereas, the average number of aphids consumed per day by these stages was 11.58, 22.28, 43.86 and 56.18 aphids, respectively. To complete the larval development, *O. kirbyi* consumed 535.90 aphids at the rate of 35.12 aphids/ day (Table 2). The adult female and male consumed on an average 2634.78 and 2012.52 aphids, respectively, during their life span.

#### **Functional response**

Mean number of aphids consumed by different developmental stages of *O. kirbyi* differed significantly among different prey densities. Logistic regression fit between prey density offered (N) and proportion of prey eaten (Na/N) by all the stages of the predator resulted in significant negative linear coefficients confirming a type II functional response for all the developmental stages of the predator (Table 3). The parameters of functional response i.e. predator's attack rate

(a) and handling time ( $T_h$ ) obtained after fitting Rogers's random predator equation are presented in Table 4. The predation efficiency of *O. kirbyi* against *A. gossypii* increased with the advancement of the developmental stage of the predator which is numerically illustrated by parameters of attackrate (a) and handling time ( $T_h$ ). Predator's attack rate was lowest (0.065) in the first-instar and highest (0.171) in the adult stage. Handling time of the adult beetle (0.237 h) and the fourth-instar larva (0.263 h) was much shorter than that of other stages (Table 4). The maximum number of aphids that could be consumed over a period of 24 h by the first, second, third and fourth instargrubs and adult of *O. kirbyi* was estimated to be 20.70, 28.60, 60.00, 92.30 and 100.00 aphids/predator, respectively.

#### Discussion

In the present study, O. kirbyi exhibited a Type II functional response in all the developmental stages to A. gossypii. The proportion of prey consumed at higher density decreased which may be due to satiation of the predator at the increased prey density. Similar to the present study, Kumari et al. (2020) <sup>[18]</sup> reported with same prey but with different predator (C. septempunctata). The present findings corroborate with the findings of Pervez and Omkar, (2005) [27] but with different predator and aphids which reported that the number of the aphids consumed increased with the increase in prey densities for all the developmental stages of the predator and when satiation was attained by the predator there was no significant increase in the number of aphids consumed and there was no effect of the prey density. The proportion of aphids consumed was maximum at the lowest density and minimum at the highest prey density.

In the present findings, a type-II functional response was observed. Similar to present findings, Harmonia axyridis exhibited a Type II functional response at all growth stages against A. gossypii, though type I (Lou, 1987)<sup>[21]</sup> or Type III (Hu et al. 1989)<sup>[11]</sup> response has also been reported for adult H. axyridis against different prey species (Rhopalosiphum prunifoliae and Cinara sp., respectively). Similar to present findings, a type II functional response is reported in many coccinellids by previous workers <sup>[19, 27, 26, 43, 12, 1, 36, 17, 24, 18]</sup>. A predator with a type II functional response has an inverse density-dependent action against its prey, whereas, a predator with a type III functional response acts as a positive density dependent mortality factor <sup>[27, 3, 42]</sup>. Sharma et al. (2017) <sup>[37]</sup> and Kumari et al. (2020)<sup>[18]</sup> also observed that the attack rate of Harmonia dimidiata and Coccinella septempuncta increased with the advancement of stages against A.gossypii and handling time shortened with the increase in stages of the predator. The predation efficiency of O. kirbvi against A. gossypii increased with the advancement of the developmental stages of the predator which is numerically illustrated by parameters of attack rate (a) and handling time (T<sub>h</sub>). This is because the fourth instar and the adult beetles arebigger in size and capable of moving faster than the other stages. In coccinellids, the fourth instar is generally responsible for approximately 60-80% of total prey consumption by larvae (Hodek and Honek, 1996) [10]. Asimilar trend was observed in our study. These proportions are similar to those of H. axyridis preying on Myzus persicae (Sulzer) (Hodek and Honek, 1996) [10]. The low predation rates observed in the first and second instars of O. kirbyi were due to their low attack rates and longer prey handling times (Table 4). This may be due to the reason that the first and second instar larvae are small in size and their movements are slow. In the present study, third instar had the highest attack rate, but the maximum mean aphid consumption was lower than the fourth instars and female adults, which may be due to a longer handling time of the third instar (Table 4). Low variation in prey consumption, high attack rate, and short handling time in fourth instars indicated that thefourth instar is the most voracious stage. Similar to present findings, Lee and Kang (2004) <sup>[19]</sup> reported that all stages of *Harmonia axyridis* showed a type II functional response against *A.gossypii* on cucumber leaves and attack rates of *H. axyridis* at 24 h were 0.0037, 0.0442, 0.3590, 0.3228, and 0.1456, and estimated handling times were 4.1001, 2.4575, 0.7500, 0.2132, and 0.1708 h for the first, second, third, and fourth instars, and female adult, respectively.

#### Conclusion

The adults and the 4<sup>th</sup> instar grubs were the voracious feeders of the melon aphid and could be important for the biological control of the aphid. Hence it can be utilized as one of the components of the integrated pest management programme of the aphid

#### References

- 1. Atlihan R, Kaydan MB, Yarimbatman A, Okut H. Functional response of the coccinellid predator *Adalia fasciatopunctata* Revelierei to walnut aphid *Callaphis juglandis*. Phytoparasitica. 2010;38:23-29.
- 2. Bayoumy MH. Foraging behavior of the coccinellid *Nephus includes* (Coleoptera: Coccinellidae) in response to *Aphis gossypii* (Hemiptera: Aphididae) with particular emphasis on larval parasitism. Environmental Entomology 2010;40:835-843.
- Costa JF, Matos CHC, de Oliveira CRF, da Silva TGF, Lima Neto IFA. Functional and numerical responses of *Stethorus tridens* Gordon (Coleoptera: Coccinellidae) preying on *Tetranychus bastosi* Tuttle, Baker & Sales (Acari: Tetranychidae) on physic nut (*Jatropha curcas*). Biological Control 2017;111:1-5.
- Denholm I, Devine G. Insecticide resistance. Encyclopedia of biodiversity (second edition). 2013, 298-307.
- 5. Escriu F, Perry KL, Garcia-Arenal F. Transmissibility of cucumber mosaic virus by *Aphis gossypii* correlates with viral accumulation. Virology 2000;90:1069-1072.
- 6. Hassell MP. The spatial and temporal dynamics of host parasitoid interactions. Oxford: Oxford University Press 2000.
- Heidrian M, Fathipour Y, Kamali K. Functional response, switching, and prey-stage preference of *Scolothrips longicornis* (Thysanoptera: Thripidae) on *Schizotetranychus smirnovi* (Acari: Tetranychidae). Journal of Asia-Pacific Entomology. 2012;15: 89-93.
- 8. Herron GA, Wilson LJ. Neonicotinoid resistance in *Aphis gossypii* Glover (Aphididae: Hemiptera) from Australian cotton. Australian Journal of Entomology 2010;50:93-98.
- Holling CS. Some characteristics of simple types of predation and parasitism. Canadian Entomology 1959;91: 385-398.
- 10. Hodek I, Honek A. Ecology of Coccinellidae. Kluwer Academic Publishers, Dordrecht 1996, 46.
- 11. Hu YS, Wang ZM, Ning CL, Pi ZQ, Gao GQ. The functional response of *Harmonia* (Leis) *axyridis* to their prey of *Cinara* sp. Natural Enemies of Insects

1989;11:164-168.

- Jafari R, Goldasteh S. Functional response of *Hippodamia variegata* (Goeze) (Coleoptera: Coccinellidae) on *Aphis fabae* (Scopoli) (Homoptera: Aphididae) in laboratory conditions. Acta Entomologica Serbica 2009;14:93-100.
- 13. Jervis MA, Kidd NAC. Insect natural enemies: Practical Approaches to Their Study and Evaluation. Chapmanand Hall, London 1996, 491.
- 14. Joshi S, Rabindra RJ, Rajendran TP. Biological control of aphids. Journal of Biological Control 2010;24:185-202.
- 15. Juliano SA. Non-linear curve fitting: Predation and functional response curves. In SM Scheiner& J. Gurevitch (Eds.), Design and analysis of ecological experiments, 2nd edn. New York: Chapman and Hall 2001.
- 16. Kianpour R, Fathipour Y, Kamali K, Naseri B. Bionomics of *Aphis gossypii* (Homoptera: Aphididae) and its predators *Coccinella septempunctata* and *Hippodamia variegata* (Coleoptera: Coccinellidae) in natural conditions. Journal of Agriculture Science and Technology 2010;12:1-11.
- 17. Kumar B, Mishra G, Omkar. Functional response and predatory interactions in conspecific and heterospecific combinations of two congeneric species (Coleoptera: Coccinellidae). European Journal of Entomology 2014;111: 257-265.
- Kumari D, Verma SC, Sharma PL. Biology, feeding potential and functional response of *Coccinella septempunctata* L. against *Aphis gossypii* Glover infesting cucumber. Journal of Entomology and Zoological Studies 2020;8:631-636.
- 19. Lee JH, Kang TJ. Functional response of *Harmonia axyridis* (Pallas) (Coleoptera: Coccinellidae) to *Aphis gossypii* Glover (Homoptera: Aphididae) in the laboratory. Biological Control 2004;31:306-310.
- Luck. Principles of arthropod predation. In Huffaker CB & Rabb RL (eds): Ecological Entomology. Wiley, New York 1985, 497-530.
- 21. Lou HH. Functional response of *Harmonia axyridis* to the density of *Rhopalosiphum prunifoliae*. Natural Enemies of Insects 1987;9:84-87.
- 22. Mhaske BM, Pardesh SR, Bhoite KD, Rasal PN. Biosafety of coccinellid predators and chemical control of wheat aphids. Agriculture Science Digest 2007;27:264-266.
- Misra HP. Newer insecticides for the management of aphid, Aphis gossypii Glov in gherkins (*Cucumis anguria* L.) and their effect on the predator, *Coccinella septempunctata* L. Pest Management in Horticulture Ecosystem 2013;19:123-127.
- 24. Mrosso F, Mwatawala M, Rwegasira G. Functional response of *Cheilomenes propinqua*, *C. lunata* and *C. sulphurea* (Coleoptera: Coccinellidae) to predation on *Aphis gossypii* (Homoptera: Aphididae) in eastern Tanzania. Journal of Entomology 2013;doi:10.2923/je.2013.
- 25. Pan Y, Tian F, Wei X, Wu Y, Gao X, Xi J *et al.* Thiamethoxam resistance in *Aphis gossypii* Glover Relies on Multiple UDP-Glucuronosyltransferases. Frontier in Physiology 2018;9:322-327.
- 26. Parajulee MN, Shrestha RB, Leser JF, Wester DB, Blanco CA. Evaluation of the functional response of selected arthropod predators on bollworm eggs in the

laboratory and effect of temperature on their predation efficiency. Environmental Entomology 2006;35:379-386.

- 27. Pervez A, Omkar. Functional responses of coccinellid predators: An illustration of a logistic approach. Journal of Insect Science 2005;5:1-6.
- 28. Pinto ZV, Rezende JAM, Valdir A, Yuki VA, Piedade SMS. Ability of *Aphis gossypii* and *Myzus persicae* to transmit cucumber mosaic virus in single and mixed infection with two Poty viruses to zucchini squash summa. Phytopathological Botucatu 2008;34:183-185.
- 29. Rezaei M, Talebi AA, Fathipour Y, Karimzadeh J, Mehrabadi M. Foraging behavior of *Aphidius matricariae* (Hymenoptera: Braconidae ) on tobacco aphid, *Myzus persicae nicotianae* (Hemiptera: Aphididae). Bulletin of Entomological Research 2019;109:840-848.
- 30. Rogers D. Random search and insect population models. Journal of Animal Ecology 1972;41:369-383.
- Sabelis MW. Predatory arthropods. In Crawley M.J. (ed.): Natural Enemies: the Population Biology of Predators, Para- sites and Diseases. Blackwell, Oxford. 1992, 225-264.
- 32. Sakaki S, Sahragard A. A new method to study the functional response of *Scymnus syriacus* (Coleoptera: Coccinellidae) to different densities of *Aphis gossypii*. Journal of Asia-Pacific Entomology 2011;14:459-462.
- Saleh AAA, El-Sharkawy HM, El-Santel FS, El-Salam RAA. Studies on the predator *Chrysoperla carnea* (Stephens) in Egypt. International Journal of Environment 2017;6:70-77.
- 34. Sarwar MK, Azam I, Iram N, Iqbal W, Rashda A, Anwer F et al. Cotton aphid Aphis gossypii L. (Homoptera; Aphididae); a challenging pest; biology and control strategies: A review. International Journal of Applied Biology and Pharmaceutical Technology 2014;5:288-294.
- 35. Seyedebrahimi SS, Jahromi KT, Imani S, Naveh VH, Hesami S. Characterization of imidacloprid resistance in *Aphis gossypii* Glover (Hemiptera: Aphididae) in Southern Iran. Turkish Journal of Entomology 2015;39:413-423.
- 36. Shah MA, Khan AA. Qualitative and quantitative prey requirements of two aphidophagous coccinellids, *Adalia tetraspilota* and *Hippodamia variegata*. Journal of Insect Science 2014;14:1-19.
- 37. Sharma PL, Verma SC, Chandel RS, Shah MA, Gavkare O. Functional response of *Harmonia dimidiata* (fab.) to melon aphid, *Aphis gossypii* Glover under laboratory conditions. Phytoparasitica 2017;45:373-79.
- Singh G, Singh NP, Singh R. Food plants of a major agricultural pest, *Aphis gossypii* Glover (Homoptera:Aphididae) from India: An updated checklist. International Journal of Life Science Biotechnology and Pharm Research 2014;3:1-28.
- Srinivasulu M, Sarovar B, Anthony Jhonson AM, Sai Gopal DVR. Association of Poty virus with mosaic virus disease of gherkin (*Cucumisanguira* L.) in India. Indian Journal of Microbiology 2010;50:221-224.
- 40. Tazerouni Z, Talebi AA, Fathipour Y, Soufbaf M. Bottom-up effect of two host plants on life table parameters of *Aphis gossypii* (Hemiptera: Aphididae). Journal of Agriculture Science and Technology 2016;18:179-190.
- 41. Tomquelski GV, Martins GLM, Papa G. Efeitos dos indutores de resistênciaacibenzolar-

smetilesilícionabiologia de *Alabama argilacea* (Lepidoptera: Noctuidae) emalgodoeiro. Revista de Agricultura 2007;82:170-75.

- 42. Uszko W, Diehl S, Pitsch N, Lengfellner K, Müller T. When is a type III functional response stabilizing? Theory and practice of predicting plankton dynamics under enrichment. figshare. Ecology 2016;96:3243-3256.
- 43. Xue Y, Bahlai CA, Frewin A, Sears MK, Schaafsma AW, Hallett RH. Predation by *Coccinella septempunctata* and *Harmonia axyridis* (Coleoptera: Coccinellidae) on *Aphis glycines* (Homoptera: Aphididae). Environmental Entomology 2009;38:708-714
- 44. Zarghami S, Mossadegh MS, Kocheili F, Allahyari H, Rasekh A. Functional Responses of *Nephus arcuatus* Kapur (Coleoptera: Coccinellidae), the Most Important Predator of Spherical Mealybug *Nipaecoccus viridis* (Newstead). Psyche 2016, 1-9 https://doi.org/10.1155/2016/9417496