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## Biochemical & histological study of glycogen profile in *Moniezia (b.) Bombayensis* sp. nov. Infecting *Capra hircus*.

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### Abstract

The present communication deals with the study of glycogen content of the cestode *Moniezia* (B.) *bombayensis* sp.nov.infecting *Capra hircus* from Kaij, Dist. Beed, MS, and India. The glycogen content of the worm is estimated from whole body and mature segment of taxonomically identified worm *Moniezia* (B.) *bombayensis* sp.nov.by Kemp *et al.*, (1954) method.

**Keywords:** Biology, brinjal, leucinodes orbonalis, morphometry

### Introduction

Cestodes are the endoparasites of alimentary canal of vertebrates which utilize the readymade nourishment available in the gastrointestinal tract for their growth and development. The literature study reveals the quantitative distribution of carbohydrates into the body of parasite. Winland (1901); Schulte, 1971, Von Brand, 1934; Salisbury and Anderson, 1939; Dougherty and Taylor, 1956; Goodchild and Vilar Alrarez, 1962 and others they have been obtained rather by unspecific chemical methods, which gives higher values than those obtained by an enzymatic procedure. Widely differing glucose values reported by Fairbrin (1958) and L'Opez-Gorge and Monteolive (1965) for *Moniezia expansa* by means of paper chromatography. The glucose contents of various helminth fluctuate considerably. The carbohydrate content of host also has an effect on growth of worms. The worms grow better in a host, feed on protein free diet containing carbohydrates.

Carbohydrate metabolism of cestode have been estimated by some workers in *Oochoristica*, *Moniezia expansa*, *M.benedini*, *Taenia saginata*, *T. pisiformis*, *T. crossiceps*, *Hymenolepis nana*, *H.utelii*, *H.diminuta*, *Phyllobothrium folliatum*, *Echinococcus*, *Dipylidium caninum* and *Bothriocephalus gowkongensis*. Cheng and Dyckman (1964) <sup>[1]</sup> described glycogen dposition in *Hymenolepis diminuta*, Chopra (1981) <sup>[2]</sup> studied glycogen contents and its distribution in cyclophyllidean cestode of sheep. Singh *et al.*, (1987) <sup>[15]</sup> described total carbohydrates and glycogen in cestode, Hiware and Jadhav (1994) <sup>[8]</sup> studied quantitative studies of glycogen in some cestodes. Pappas Barly and Werdropsm (1999) <sup>[10]</sup> studied glucose and glycogen gradient in *H.diminuta* and Ramlingam *et al.*, (2004) studied carbohydrate profile in relation to growth and differentiation of proglottids in *Avitellina lahora*, Nanware and Bhure, (2011) <sup>[8]</sup> have studied glycogen profile of cestodes of *Capra hircus*.

### Material and Methods

Ten intestines of *Capra hircus* were brought to the laboratory and dissected. Five of them were formed to be infected with cestode parasites. Identical worms were stored out with the help of microscope and few of them fixed in 4% formalin for identification. Later these were identified as *Moniezia (B.) bombayensis* sp. nov.

The collected worms were dried on blotting paper to remove excess of water. The material transferred in previously weighed watch glass and weighed on a electronic balance. The wet weight of tissue taken and kept in oven at 60<sup>o</sup>c for twenty four hours to make the material dry. The dry weight of material was taken and powder is prepared. The 100 mg powder is taken and homogenized in mortar and pestle; 5 ml of 5 % T.C.A. solution is added into it and transferred in centrifuge tube. Then this material is digested in boiling water bath for 5 minutes, then cooled and centrifuged for 15 minutes at 2000 RPM. One ml of supernatant liquid taken in a test tube added 3 ml of sulphuric acid and cooled for 5 minutes.

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The mixture shaken well, then immediately cooled and readings were taken in a Ermas colorimeter at 530 mu filter. The glycogen content of these worms was determined by Kemp *et al.*, (1954)<sup>[11]</sup> method.

For the histochemical study of *Moniezia (B.) bombayensis* sp.nov. The worms washed with the tap water, few of them were fixed in Carnoy's fluid for determination of glycogen content of the worm. The worm were removed from the fixative, washed with water, dehydrated through alcoholic grades and embedded in Paraffin wax (60 % C.M.P.) and blocks were prepared. The blocks were cut at 7 micron thickness with the help of microtome machine, stained with haematoxyline and Best's caramine high concentration of the glycogen integument, up to muscle layer. The traces of glycogen are observed.

### Discussion

1. Estimation of Glycogen by Kemp *et al.*, (1954)<sup>[11]</sup> method

The amount of glycogen in the worm is calculated by-

$$\text{Percentage of Glycogen} = \frac{100 \times U}{1.11 \times S}$$

Where

U = O.D. of the unknown test solution.

S = O.D. of 100 mg of glucose to glycogen.

1.11 = Conservation factor of glucose to glycogen.

U = 0.52, S = 2

$$\text{Percentage of Glycogen} = \frac{100 \times 0.54}{1.11 \times 2}$$

= 23.42 mgs/100 ml test solution.

The amount of glycogen calculated in the host intestine of *Capra hircus* is 23.42 mgs/glycogen/gm of intestine homogenate.

### 2. Histological study of Glycogen in *Moniezia (B.) bombayensis* sp. nov.

Observations: The traces of glycogen are observed at the centers of segment and peripheral region and ovary. In the testes moderate quantity of glycogen is seen. The glycogen concentration is also seen in the lateral parenchyma. The mature segment shows relatively moderate amount of glycogen in the longitudinal muscles and peripheral region and more concentrated in the reproductive organs. The glycogen stained pink red to red in color. The prepared slide when observed under microscope shows high concentration of glycogen in its body.



**Fig 1:** Glycogen content in *Moniezia (B.) bombayensis* sp.nov. Longitudinal section of mature segment showing distribution of glycogen.

### Result

The glycogen content of the host and the worm *Moniezia (B.) bombayensis* sp.nov. Shows that, the worm is quite successful in obtaining the sufficient amount of glycogen from environment. Thus the worm could maintain good balance of glycogen content.

### Conclusion

Histologically worm *Moniezia (B.) bombayensis* sp.nov. Shows high concentration of glycogen in the body which is absorbed from the host tissue.

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### References

1. Cheng TC, Dyckman E. Sites of glycogen deposition in *H. diminsuta* during the growth phase in the Rodents Z.

Parasitkde 1964;24:27-48.

2. Chopra AK. Glycogen content and its distribution in three *Cylophylidean* cestodes of sheep. *Comp. Physiol. Ecol* 1981;6:173-176.

3. Daugherty JW, Taylor D. Regional distribution of glycogen in the rat cestode, *Hymenolepis diminuta*. *Expt. Parasitol* 1956;5:376-390.

4. Goodchild DG. Carbohydrate content of the tapeworm *H. diminuta* from normal bile less and starved rats. *J Parasit* 1961;47:401-405.

5. Graff D, Allen K. Glycogen content in *Moniliformis dubius* (Acanthocephala). *J Parasitol* 1963;49(2):204-208.

6. Hiware CJ, Jadhav BV. Quantitative studies on Glycogen in some cestodes collected from different hosts and localities of Western Maharashtra. *Ind. Jr. Helminthology*, Dr. C.B. Srivastava Comm 1994, 219-222.

7. Hopkins CA. Studies on cestode metabolism I. Glycogen metabolism in *Schistocephalus solidus* *in vivo*. *J Parasit* 1950;36:384-390.

8. Jadhav BV, Shivesh Singh P, Bhure DB, Padwal ND.

- Biosystematic studies of *Davainea shindei* n.sp. (Cestoda- Davainidae) Fuhrmann, 1907 from *Gallus gallus domesticus*. National Academy of Science Letter 2008;31(7-8):245-250.
9. Kemp Vankites A, Haijnin Gen AJM. A Colourimetric method for the determination of glycogen in the tissue Biochem J 1954;56:646-648.
  10. Pappas PW, Barley AJ, Werdrop SM. Hymenolepis diminuta: Glucose and glycogen gradients in adult tapeworm. Exptl. Parasitol 1999;46:315-326.
  11. Ramalingam K, Vijayalakshmi V, Satyaprema VA. Carbohydrate profile in relation to growth and differentiaationn of proglottids in *Avitellina lahorea* (Woodland, 1927), an aoplocephalid cestode Uttar Pradesh J. Zool 2004,24(3).
  12. Read CP. Carbohydrate metabolism of Hymenolepis *diminuta*, Expt. Parasitol 1956;5:325-344.
  13. Read CP, Rothman AH. The role of carbohydrates in the biology of cestodes VI. The carbohydrates metabolised *in vitro* by some Cyclophyllidean species. Expt. Parasitol 1958;7:217-223.
  14. Read CP, Simmons JE Jr. Carbohydrate metabolism in Hymenolepis (cestoda). J. Parasit 1967;53:1023-1029.
  15. Singh SP, Capoor VN, Misra DS. Quantitative estimation of total carbohydrate and glycogen contents with carbohydrate metabolism in four species of cestode parasites Indian J. Helminth 1987;19:101-106.
  16. Smyth JD, Hopkins CA. The physiology of parasites, Cambridge University Press, 1983.
  17. Theron Odlaug O. The quantitative determination of glycogen in some parasites of Amphibia, The Journal of Parasitology 1955;41(3):258-268.
  18. Von Brand. The carbohydrate metabolism of parasites J Parasit 1950;36:174-192.
  19. Von Brand T. Biochemistry of parasites, Academic Press, New York and London, 1966.
  20. Von Brand T. The carbohydrate metabolism of parasites J Parasit 1950;36:174-192.
  21. Woodland WNF. On some remarkable new forms of Carbohydrates form the Angolo Egyptian, Sudan and revision of the families of the cestodaria Quardr J Micr. Sci 1923;67:435-472.