

E-ISSN: 2320-7078 P-ISSN: 2349-6800 www.entomoljournal.com

JEZS 2020; 8(5): 1301-1305 © 2020 JEZS Received: 26-07-2020 Accepted: 30-08-2020

M Das

Senior Scientist, Division of Animal Health, ICAR Research Complex for NEH Region, Umiam, Meghalaya, India

R Laha

Principal Scientist, Division of Animal Health, ICAR Research Complex for NEH Region, Umiam, Meghalaya, India

S Doley

Principal Scientist, Division of Livestock Production, ICAR Research Complex for NEH Region, Umiam, Meghalaya, India

Corresponding Author: M Das Senior Scientist, Division of Animal Health, ICAR Research Complex for NEH Region, Umiam, Meghalaya, India

Journal of Entomology and Zoology Studies

Available online at www.entomoljournal.com



Gastrointestinal parasites in backyard poultry of subtropical hilly region of Meghalaya

M Das, R Laha and S Doley

Abstract

The aim of the present study was to determine the prevalence, species diversity and intensity of gastrointestinal (G.I.) parasitic infections in the backyard poultry of Meghalaya. A total of 1775 numbers of fecal samples were collected from different age groups *viz.* < 8 weeks (523 nos.), 8-28 weeks (602 nos.) and > 28 weeks (650 nos.) and examined by flotation, sedimentation and modified McMaster techniques. Overall prevalence of G.I. parasitic infections was 37.97%. Eight species *viz. Eimeria* sp. (30.12%), *Heterakis gallinarum* (14.09%), *Ascaridia galli* (21.22%), *Strongyloides avium* (12.46%), *Capillaria* sp. (7.57%), *Raillietina* sp. (8.61%), *Syngamus trachea* (3.56%) and *Choanotaenia infundibulum* (2.37%) were recorded. Highest and lowest infections were recorded in October (52.88%) and February (26.34%), respectively. Season wise highest infection recorded during monsoon (44.71%) followed by autumn (44.34%), winter (27.22%) and spring (36.62%). *Eimeria* sp. was recorded highest in monsoon (33.87%), winter (27.78%) and spring (12%) seasons while A. galli (31.63%) in autumn season. Age wise variations in infections were observed in < 8 (25.24%), 8-28 (48.17%) and > 28 (38.77%) weeks old birds. *Eimeria* sp. was observed highest in both < 8 (68.18%) and 8-28 (25.86%) weeks. *A. galli* (27.38%) was recorded highest in > 28 weeks old birds.

Keywords: Backyard poultry, gastrointestinal, parasites, Meghalaya

Introduction

Animal husbandry is an important subsector of Indian agriculture and backyard poultry farming is one of the important components of animal husbandry among the tribal farmers of Meghalaya. Backyard poultry farming is increasing rapidly due to low establishment cost, cheap source of proteins and employment ^[1, 2]. As per the 20th Livestock census, the total poultry population of India is 851.81 million, increased by 16.8% from the previous census. Total backyard and commercial poultry in the country is 317.07 million and 534.74 million, respectively ^[3]. Parasitism is an association in which the parasite is metabolically dependent to a greater or lesser extent to the host. Poultry birds gets infection by ingestion of contaminated feed, water, litter, intermediate host etc ^[4]. Gastrointestinal (G.I.) parasites are most prevalent and devastating parasites affecting its productivity ^[5]. Severe infections with G.I. parasites may decrease production performance as well as cause high morbidity and mortality ^[6, 7]. Bhowmik et al.^[8] observed that in growing chicks parasitism causes 17% reduction in weight gain, high morbidity and mortality while in egg laying hens it causes 12.5% reduction in egg production. They are also associated with catarrh, diarrhea, intestinal obstruction, loss of appetite, anemia, weakness, paralysis and poor feathering in birds ^[9, 10]. The climatic condition of the North Eastern region is highly congenial for the development and propagation of different parasites. Though there are reports on the prevalence of G.I. parasitic infections in the poultry from different states of India like Chhattisgarh ^[11], Assam ^[12], Uttar Pradesh and Uttarakhand ^[13], Maharashtra ^[14], Madhya Pradesh ^[15], Jammu ^[16], Karnataka ^[17] but no information is available from Meghalaya. Thus, the present study has been undertaken to explore the prevalence of G.I. parasites in the backyard poultry of hilly region of Meghalaya.

Materials and Methods

Study area

The present study was conducted in the Ri Bhoi district of Meghalaya which lies between $25^{\circ}15'$ and $26^{\circ}15'$ North latitudes and $91^{\circ}45'$ and $92^{\circ}15'$ East longitudes. The district is characterized by rugged and irregular land surface and includes a series of hill ranges. (https://en.wikipedia.org/wiki/Ri-Bhoi_district).

Study period

The study was conducted for two years (2018, 2019) and divided into four seasons, *viz.* Spring (March, April), Monsoon (May, June, July, August, September), Autumn (October, November) and Winter (December, January, February).

Sample collection

Freshly voided fecal samples were collected from the poultry shed of different locations i.e. Umiam, Umsawkhwan, Mawphrew, Nalapara, Borkhatsari, Sarikuchi, Umthan and Lalumpam in marked plastic pouch/vials. All the birds were categorized according to age *viz.* < 8 weeks, 8-28 weeks and >28 weeks. A total of 1775 numbers of fecal samples of poultry were collected from different age groups i.e. < 8 weeks (523nos.), 8-28 weeks (602nos.) and >28 weeks (650nos.).

Parasitological techniques

To detect G.I. parasitic infections in the backyard poultry of hilly region of Meghalaya, faecal samples were examined by direct flotation technique using saturated salt (sp.gr. 1.20) and sucrose (sp.gr. 1.27) solution ^[4]. Positive samples were then quantified to estimate the egg per gram (EPG) of faeces by using modified McMaster technique ^[18]. Samples not being

examined on the same day were preserved at refrigerated temperature $(4^{\circ}C)$ for next day examination.

Results and Discussion

Prevalence of G.I. parasitic infections in backyard poultry of Meghalaya

The overall prevalence of G.I. parasitic infections in the backyard poultry of hilly region of Meghalaya was 37.97% (Table 1). Eight species of G.I. parasites were recorded viz. Eimeria sp. (30.12%), Heterakis gallinarum (14.09%), Ascaridia galli (21.22%), Capillaria sp. (7.57%), Syngamus trachea (3.56%), Raillietina sp. (8.61%), Choanotaenia infundibulum (2.37%) and Strongyloides avium (12.46%) (Fig.1). In congruence to the present findings, Kumari et al. ^[11], Hembram *et al.* ^[19], Katoch *et al.* ^[16] and Naphade and Chaudhari [14] reported 25%, 58.75%, 72% and 84.05% parasitic infections from Chattishgarh, Odisha, Jammu and Madhya Pradesh, respectively. Similarly, Nguyen et al. ^[20], Wamboi et al. ^[21], Islam et al. ^[22] and Berhe et al. ^[23] reported 54.2%, 86.6%, 19.4% and 90.97% infections from Vietnam, Kenya, Bangladesh and Ethiopia, respectively. Variation in the percent prevalence from the current study may be due to difference in the geographical location, environmental condition and management practices adopted by the farmers.

Table 1: Season wise prevalence of G.I. parasites in backyard poultry of Meghalaya

Season	Sample examined	Sample positive	<i>Eimeria</i> sp.	H. gallinarum	A. galli	Capillaria sp.	S. trachea	Raillietina sp.	Choanotaenia sp.	S. avium
Winter	529	144 (27.22)	40 (27.78)	25 (17.36)	31 (21.53)	12 (8.33)	3 (2.08)	13 (9.03)	-	20 (13.89)
Spring	325	119 (36.62)	39 (12)	20 (6.15)	18 (5.54)	12 (3.69)	5 (1.54)	7 (2.15)	5 (1.54)	13 (4)
Monsoon	700	313 (44.71)	106 (33.87)	33 (10.54)	63 (20.13)	24 (7.67)	9 (2.88)	27 (8.63)	6 (1.92)	45 (14.38)
Autumn	221	98 (44.34)	18 (18.37)	17 (17.35)	31 (31.63)	3 (3.06)	7 (7.14)	11 (11.22)	5 (5.10)	6 (6.12)
Total	1775	674 (37.97)	203 (30.12)	95 (14.09)	143 (21.22)	51 (7.57)	24 (3.56)	58 (8.61)	16 (2.37)	84 (12.46)

Figures in parentheses indicates percent positivity



Fig 1: Eggs/oocyst of G.I. parasites in backyard poultry of Meghalaya

In the present study, monthwise highest and lowest infections were recorded in the month of September (49.52%) and February (26.34%), respectively. The intensity of infection i.e. egg per gram (EPG) of faeces ranges from 50-550.

Maximum and minimum mean intensity (EPG) of infection was recorded in the month of July (285.43) and February (149.35), respectively (Fig. 2).



Fig 2: Month wise prevalence of G.I. parasites in backyard poultry of Meghalaya

Season wise infection was recorded highest during monsoon (44.71%) followed by autumn (44.34%), winter (27.22%) and spring (36.62%) seasons (Table 1). In agreement with the present findings, earlier Islam et al. [22], Sreedevi et al. [24], Hembram et al.^[19] and Salam et al.^[25] from Bangladesh, Andhra Pradesh, Odisha and Kashmir reported 26.5%, 43.41%, 68.88% and 33.62% infections during rainy/monsoon seasons, respectively. Environment, management practices, level of bio-security, availability of intermediate hosts and reservoirs are also key factors for the high prevalence of parasitic infections in poultry ^[26, 27]. Taylor *et al.* ^[28] observed that optimum temperature and relative humidity for development and hatching of eggs or oocyst are 26-29°C and >80%, respectively. The development is decreased below 10°C and low relative humidity. This shows that monsoon season is very conducive for the development and propagation of parasites in the backyard poultry of Meghalaya.

During monsoon season, Eimeria sp. (33.87%) was recorded highest followed by A. galli (20.13%), S. avium (14.38%), H. gallinarum (10.54%), Raillietina sp. (8.63%), Capillaria sp. (7.67%), S. trachea (2.88%) and C. infundibulum (1.92%) (Table 1). However, in autumn season A. galli (31.63%) was recorded highest followed by Eimeria sp. (18.37%), H. gallinarum (17.35%), Raillietina sp. (11.22%), S. trachea (7.14%), S. avium (6.12%), C. infundibulum (5.10%) and Capillaria sp. (3.06%). In spring season, Eimeria sp. (12%) was recorded highest followed by H. gallinarum (6.15%), A. galli (5.54%), S. avium (4%), Capillaria sp. (3.69%), Raillietina sp. (2.15%), S. trachea (1.54%) and C. infundibulum (1.54%). In winter season also Eimeria sp. (27.78%) was recorded highest followed by A. galli (21.53%), H. gallinarum (17.36%), S. avium (13.89%), Raillietina sp. (9.03%), Capillaria sp. (8.33%) and S. trachea (2.08%). Different species of G.I. parasites are prevalent throughout the year in hilly region of Meghalaya which may be due to sufficient moisture in the litter, humidity and ambient temperature for growth and development of parasitic egg/ova throughout the year.

Age wise prevalence of G.I. parasitic infections in backyard poultry of Meghalaya

Age wise G.I. parasitic infections was recorded in all age groups of poultry *viz.* < 8 weeks (25.24%), 8-28 weeks (48.17%) and > 28 weeks (38.77%) (Table 2). The prevalence of *Eimeria* sp. was recorded highest in the birds of < 8 weeks

(68.18%) followed by 8-28 weeks (25.86%) and > 28 weeks (15.08%). In birds of < 8 weeks age, *Eimeria* sp. (68.18\%), *H*. gallinarum (18.18%) and Capillaria sp. (13.64%) were only observed. While in the birds of 8-28 weeks age, all the eight species of G.I. parasites were observed viz. Eimeria sp. (25.86%), A. galli (25.52%), H. gallinarum (14.13%), Capillaria sp. (6.89%), Raillietina sp. (6.89%), S. trachea (3.45%), C. infundibulum (3.45%) and S. avium (13.79%). However in the birds of > 28 weeks age, the percentage of infection is lower in comparison to 8-28 weeks old birds i.e. A. galli (27.38%), S. avium (17.46%), Eimeria sp. (15.08%), Raillietina sp. (15.08%), H. gallinarum (11.90%), S. trachea (5.56%), Capillaria sp. (5.16%) and C. infundibulum (2.38%). Age wise variation in the prevalence of G.I. parasitic infection in poultry was also reported by Islam et al. [22]; Wokem and Obiyor^[29]; Sheikh et al. ^[30] and Hembram et al. ^[19]. High rate of infection in young birds may be due to decreased immunity as well as continuous exposure to infections from the contaminated litter. In the present study, Eimeria sp. was recorded in all age groups and highest in birds of < 8 weeks (68.18%) which is responsible for causing coccidiosis. It is characterized by dysentery, bloody diarrhoea, enteritis, poor growth, drooping wings, emaciation and decreased production ^[31]. According to Bera et al. ^[32] approximately US\$ 20 million/annum coccidiosis associated economic losses were recorded in India. Sharma et al. [33] and Debbou-Iouknane et al. [34] from Jammu and Algeria reported 58.86% and 54.28% Eimeria sp. infection in young poultry birds, respectively. However, Badran and Lukesouna ^[35] reported *Eimeria* sp. infection in all ages. In the present study, A. galli infection was recorded highest in > 28 weeks (27.38%) birds in comparison to 8-28 weeks (25.52%) old birds, which may be due to frequent contact with the intermediate host and external environment. Earlier Rashid et al. [36], Fatima et al. ^[37] and Zada et al. ^[38] also observed A. galli infection more in adults, than young birds which corroborates with the present findings.

The present study has significance because eight species of G.I. parasites were recorded for the first time in the different age groups of backyard poultry in the hilly region of Meghalaya. Usually birds pick up infection from contaminated litter having parasitic eggs/ova or intermediate host ^[39] and heavy infection in birds will decrease egg production, weight gain and haemoglobin depression ^[40].

Table 2: Age wise prevalence of	f G.I. parasites in back	yard poultry of Meghalaya
---------------------------------	--------------------------	---------------------------

Age (weeks)	Sample examined	Sample positive	<i>Eimeria</i> sp.	H. gallinarum	A. galli	<i>Capillaria</i> sp.	S. trachea	<i>Raillietina</i> sp.	Choanotaenia sp.	S. avium
< 8	523	132 (25.24)	90 (68.18)	24 (18.18)	-	18 (13.64)	-	-	-	-
8-28	602	290 (48.17)	75 (25.86)	41 (14.13)	74 (25.52)	20 (6.89)	10 (3.45)	20 (6.89)	10 (3.45)	40 (13.79)
> 28	650	252 (38.77)	38 (15.08)	30 (11.90)	69 (27.38)	13 (5.16)	14 (5.56)	38 (15.08)	6 (2.38)	44 (17.46)
Total	1775	674 (37.97)	203 (30.12)	95 (14.09)	143 (21.22)	51 (7.57)	24 (3.56)	58 (8.61)	16 (2.37)	84 (12.46)

Figures in parentheses indicates percent positivity

Conclusion

The present study revealed that different species of G.I. parasites are prevalent throughout the year in backyard poultry of Meghalaya. Highest infection observed during rainy season and young age groups are more susceptible. Regular screening and deworming of bird is necessary for profitable backyard poultry farming.

Acknowledgement

We are thankful to the Director, ICAR Research Complex for NEH Region, Umiam, Meghalaya for providing financial assistance and other facilities to carry out this research work under the Institute project (IXX15052).

References

- 1. Frantovo D. Some parasitic nematodes (Nematoda) of birds (Aves) in the Czech Republic. Acta Societatis Zoologicae Bohemicae. 2000; 66(1):13-28.
- Bachaya HA, Raza MA, Khan MN, Iqbal Z, Abbas RZ, Murtaza S *et al.* Predominance and detection of different *Eimeria* species causing coccidiosis in layer chickens. Journal of Animal and Plant Sciences. 2012; 22:597-600.
- 3. Livestock Census. 20th Livestock Census. Department of Animal Husbandry, Dairying and Fisheries, Ministry of Agriculture, Government of India, 2019.
- 4. Soulsby EJL. Helminths, arthropod and protozoa of domestic animals. 7th Edn. The English Language Book Soc., Bailliere Tindal, London, 1982.
- 5. Swaton HK, Tshovhote J, Nesamvumi E, Ranwedzi NN, Fouric C. Characterization of indigenous free ranging poultry production system under traditional management conditions in the Vhembe district of the Limpopo Province, South Africa. Corpus ID: 54922369, 2004.
- Luka SA, Ndams IS. Gastrointestinal parasites of domestic chicken *Gallus Gallus domesticus* Linnaeus 1758 In Samaru, Zaria Nigeria. Science World Journal. 2007; 2(1):27-29.
- 7. Gauly M, Homann T, Erhardt G. Age related differences of *Ascaridia galli* egg output and worm burden in chickens following a single dose infection. Veterinary Parasitology. 2005; 128(1-2):141-148.
- 8. Bhowmik MK, Sasmal NK, Chakraborty AK. Effect of *Raillietina cesticellus* infection on the meat and egg production of fowl. Indian Veterinary and Medicine Journal. 1982; 6(2):100-102.
- Jegede OC, Asadu IA, Opara M, Obeta SS, Olayemi DO. Gastrointestinal parasitism in local and exotic breeds of chickens reared in Gwagwalada Guinea Savannah zone of Nigeria. Sokoto Journal of Veterinary Sciences. 2015; 13(3):25-30.
- Afolabi OJ, Simon-Oke IA, Olasunkanmi AO. Intestinal parasites of domestic chicken (*Gallus gallus domesticus*) in Akure, Nigeria. Journal of Biomedicine. 2016; 1(4):1-4.
- 11. Kumari B, Pal S, Sanyal PK, Verma SK. Studies on

Prevalence of Gastrointestinal Helminthic Infections in Poultry of Durg (Chhattisgarh). International Journal of Pure and Applied Bioscience. 2018; 6(3):570-574.

- Kalita A, Sarmah PC, Borah MK, Hussain L, Bhattacharjee K. Magnitude of Coccidia Infection in Small Scale Broiler Chicken Farms of Rural Assam (India). International Journal of Current Microbiology and Applied Science. 2018; 7(10):3399-3403.
- Kumar S, Garg R, Ram H, Maurya PS, Banerjee PS. Gastrointestinal parasitic infections in chickens of upper Gangetic plains of India with special reference to poultry coccidiosis. Journal of Parasitic Disease. 2015; 39(1):22-26.
- 14. Naphade ST, Chaudhari KV. Studies on the Seasonal Prevalence of Parasitic Helminths in Gavran (Desi) Chickens from Marathwada Region of Maharashtra. International Journal of Fauna and Biological Studies. 2013; 1(2):4-7.
- 15. Shukla S, Mishra P. Gastro Intestinal Helminths Parasites of Local Chickens Samples from Tribal Areas of Madhya Pradesh. International Journal of Life Sciences. 2013; 1(4):284-287.
- 16. Katoch R, Yadav A, Godara R, Khajuria JK, Borkataki S, Sodhi SS *et al.* Prevalence and impact of gastrointestinal helminths on body weight gain in backyard chickens in subtropical and humid zone of Jammu, India. Journal of Parasitic Disease. 2012; 36:49-52.
- 17. Puttalakshmamma GC, Ananda KJ, Prathiush PR, Mamatha GS, Rao S. Prevalence of Gastrointestinal parasites of Poultry in and around Banglore. Veterinary World. 2008; 1(7):201-202.
- MAFF. Ministry of Agriculture, Fisheries and Food. Manual of veterinary parasitological techniques, Her Majesty's Stationery Office, London, 1986.
- 19. Hembram A, Panda MR, Mohanty BN, Pradhan CR, Dehuri M, Sahu A *et al.* Prevalence of gastrointestinal helminths in Banaraja fowls reared in semi-intensive system of management in Mayurbhanj district of Odisha. Veterinary World. 2015; 8(6):723-726.
- Nguyen T BV, Nguyen VC, Nguyen TPY, Nguyen TH N, Bach TK, Nguyen VH *et al.* Characterisation of gastrointestinal helminths and their impact in commercial small-scale chicken flocks in the Mekong Delta of Vietnam. Tropical Animal Health and Production. 2020; 52:53-62.
- Wamboi P, Waruiru RM, Mbuthia PG, Nguhiu JM, Bebora LC. Haemato-biochemical changes and prevalence of parasitic infections of indigenous chicken sold in markets of Kiambu County, Kenya. International Journal of Veterinary Science and Medicine. 2020; 8(1):18-25.
- 22. Islam MS, Dey AR, Parvin S, Farjana T, Alam MZ. Intestinal parasitic infection in commercial chickens in Sirajgonj. Journal of Bangladesh Agricultural University. 2020; 18(1):111-116.

- Berhe M, Mekibib B, Bsrat A, Atsbaha G. Gastrointestinal Helminth Parasites of Chicken under Different Management System in Mekelle Town, Tigray Region, Ethiopia. Journal of Veterinary Medicine, 2019. <u>https://doi.org/10.1155/2019/1307582</u>.
- 24. Sreedevi C, Jyothisree C, Rama Devi V, Annapurna P, Jeyabal L. Seasonal prevalence of gastrointestinal parasites in desi fowl (*Gallus gallus domesticus*) in and around Gannavaram, Andhra Pradesh. Journal of Parasitic Disease. 2016; 40(3):656-661.
- 25. Salam TS, Mir S, Khan MR. The prevalence and pathology of *Raillietina cesticillus* in indigenous chicken (*Gallus gallus domesticus*) in the temperate Himalayan region of Kashmir-short communication. Veterinarski Arhiv. 2010; 80:323-328.
- 26. Catelli CTE, Poglayen G, Gadale ATO. Preliminary study of the helminthes of The chicken digestive tract in Somalia. Pathologie Infectieuse. 1999; 52:107-112.
- 27. Permin A, Bisgaard M, Frandsen F, Pearma M, Kold J, Nansen P *et al.* Prevalence of gastrointestinal helminths in different poultry production systems. British Poultry Science. 1999; 40(4):439-443.
- 28. Taylor M, Coop R, Wall R. Veterinary Parasitology. 4th Edn: Wiley Blackwell, 2016.
- 29. Wokem GN, Obiyor ET. Assessment of Intestinal Parasites of Commercial Layers in Selected Local Government Areas of Rivers State, Nigeria and Their Public Health Implications. Current Trends in Biomedical Engineering and Bioscience. 2018; 11(5). CTBEB.MS.ID.555822.
- Sheikh BA, Ahmad F, Sofi TA. Morphology and Prevalence of Some Helminth Parasites in *Gallus domesticus* from Gurez Valley of Jammu and Kashmir, India. Journal of Fisheries and Livestock Production. 2016; 4:159.
- Gerhold RW. Overview of coccidiosis in poultry. Merck Veterinary Manual. 10th Edn. Merck & Co., Inc., Kenilworth, NJ, USA, 2015.
- 32. Bera AK, Bhattacharya D, Pan D, Dhara A, Kumar S, Das S *et al.* Evaluation of economic losses due to coccidiosis in poultry industry in India. Agricultural Economics Research Review. 2010; 23:91-96.
- 33. Sharma S, Iqbal A, Azmi S, Mushtag I, Wani AZ, Ahmad S *et al.* Prevalence of poultry coccidiosis in Jammu region of Jammu and Kashmir State. Journal of Parasitic Disease. 2015; 39(1):85-89.
- 34. Debbou-Iouknane N, Benbarek H, Ayad A. Prevalence and aetiology of coccidiosis in broiler chickens in Bejaia province, Algeria. Onderstepoort Journal of Veterinary Research. 2018; 85(1):1-6.
- 35. Badran I, Lukesova D. Control of coccidiosis and different coccidia of chicken in selected technologies used in tropics and subtropics. Agricultura Tropica et Subtropica. 2006; 39(1):39-43.
- 36. Rashid M, Akbar H, Bakhsh A, Rashid MI, Hassan MA, Ullah R *et al.* Assessing the prevalence and economic significance of coccidiosis individually and in combination with concurrent infections in Pakistani commercial poultry farms. Poultry Science. 2019; 98:1167-1175.
- Fatima T, Sajid MS, Saleemi MK, Iqbal Z, Siddique RM. Descriptive epidemiology of endo-parasitic fauna in layer birds (*Gallus domesticus*) of central Punjab. Pakistan Journal of Agricultural Science. 2015; 52:815-820.

- 38. Zada L, Rehman T, Niaz S, Zeb M, Ruqia B, Salma KM *et al.* Prevalence of *Ascaridia galli* in some poultry farms of district Mardan. Journal of Advances in Parasitology. 2015; 2:75-79.
- 39. Janquera P. Parasites of dogs, cats, horses and livestock: Biology and control. Parasitipedia. Net, 2017.
- 40. Nair VK, Nadakal AM. Haematological changes in domestic fowl experimentally infected with the cestode (*Raillietina tetrogona* Molin, 1858). Veterinary Parasitology. 1981; 81:49-58.