

Journal of Entomology and Zoology Studies

Journal of Entomology and Zoology Studies

Available online at www.entomoljournal.com

E-ISSN: 2320-7078 P-ISSN: 2349-6800

$\underline{www.entomoljournal.com}$

JEZS 2020; 8(2): 1557-1561 © 2020 JEZS Received: 25-01-2020 Accepted: 27-02-2020

Dr. Alok Khemraj Wankar

Assistant Professor, Department of Veterinary Physiology, College of Veterinary & Animal Sciences, (MAFSU), Parbhani, Maharashtra, India

Dr. Gyanendra Singh

Sr. Principle Scientist, Nuclear Research Laboratory Physiology & Climatology Division, Indian Veterinary Research Institute, Izatnagar, Bareilly, Uttar Parades, India

Dr. Brijesh Yadav

Assistant Professor, Department of Physiology, College of Veterinary Science & Animal Husbandry, Veterinary University, Mathura, Uttar Parades, India

Corresponding Author:
Dr. Alok Khemraj Wankar
Assistant Professor, Department
of Veterinary Physiology,
College of Veterinary & Animal
Sciences, (MAFSU), Parbhani,
Maharashtra, India

Physio-metabolic responses in buffaloes exposed to sustained thermal stress: Acclimatization dynamics

Dr. Alok Khemraj Wankar, Dr. Gyanendra Singh and Dr. Brijesh Yadav

Abstrac

The study was conducted at Physiology & Climatology Division, IVRI on four dry adult *Murrah* buffaloes to observe the physio-metabolic alterations and acclimatization process during thermal stress [HS]. The animals were exposed in climatic chambers at 25, 30, 35 and 40 $^{\circ}$ C; 21 days at each treatment. Daily exposure period was fixed from 10.00-15.00 hrs. Rectal temperature [RT], respiration rate [RR], pulse rate [PR] were measured daily in morning and afternoon. Blood was collected on day one and day 21. There was a significant increase in RR, RT & PR at 35 and 40 $^{\circ}$ C, similarly enzymes AST, SOD, serum sodium, potassium also increased during heat stress period (P<0.05). No alteration was observed for other electrolytes, lipid and protein metabolites in the study. The results of the present study indicate that HS severely compromises thermoregulation, alters physio-metabolism and animals require more than 21 days to adapt to short/long term thermal stress.

Keywords: Physio-metabolic, acclimatization, thermal stress, thermoregulation, adapt

Introduction

The buffalo is mainly reared for its milk, meat and other agricultural purposes for centuries. Despite its natural selection to various geographical regions, still it is more adversely affected to high ambient temperatures and humidity [high THI]. Primary reasons for this low heat sustenance are its black colored skin and sparse sweat glands as compared to cattle [1]. There are immediate alterations in physiological responses, followed by short and long term metabolic adaptations during heat stress [HS]. These adaptations help in acclimatization to single/multiple stressors, essential for animal survival [2] but, at the cost of energy diversion leading to suboptimal production and reproductive performance. The negative impact of HS on animal husbandry will be further aggravated due to climate change and global warming in coming years. It is predicted that there will be a 1 to 6 °C increment in global temperature in 21 century [3].

There are several studies reporting decrease in feed intake, milk yield, reproduction, overall performance and alterations in physio-endocrinal axis during HS in livestock ^[4-7]. The impact of HS is more in tropical and sub-tropical regions where summer is characterized by high ambient temperatures and humidity for prolonged durations coupled with occasional heat waves. The actual acclimatization process and duration required are still not clearly recorded for different livestock species. In this scenario the present study was planned to observe the physio-metabolic responses and their duration in adult dry buffaloes during prolonged heat stress [low to high temperatures].

Materials and Methods

Experimental animals and treatments

The study was conducted on four adult, dry *Murrah* buffaloes (age: 5 > yrs; body weight: > 490 kg) at psychrometric chambers at Division of Physiology & Climatology, Nuclear Research Laboratory, Indian Veterinary Research Institute, Bareilly, India. All animals were kept under identical managemental and feeding conditions, with *ad libitum* water throughout the experiment ^[8]. The animals were exposed serially to increasing temperatures of 25, 30, 35 and 40 °C in psychrometric chamber, 21 days at each temperature. Humidity was fixed at comfortable 60% in the chamber at all the treatments and daily exposure period was form 10.00 to 15.00 hrs.

Sampling

Physiological recordings were made twice daily i.e. at 10.10 and 15.10 hrs, respectively. Rectal temperature [RT: °C] was recorded by inserting digital thermometer in rectum of animals and removed when the temperature reached maximum. Respiration rate [RR: breaths/min] were recorded by observing the flank movements for one minute. Pulse rate [PR:beats/min] was recorded by palpation of coccygeal artery at base of tail for one minute.

Blood samples were drawn by jugular venipuncture on day one and day 21 in morning (10.20 hrs) and afternoon (15.20 hrs) period at each treatment. Serum was harvested [3000 g x 20 min] and stored at -20 °C, till further analysis.

Biochemical parameters

In all the serum samples sodium, potassium, calcium, chloride, total protein, albumin, globulin, urea, creatinine, cholesterol, triglyceride, aspartate aminotransferase [AST], alanine aminotransferase [ALT] were estimated by commercial kits (span diagnostics). While reactive oxygen molecules [ROS] and superoxide dismutase [SOD] were analysed by other protocol [9, 10].

Statistical analysis

Data were analyzed by the by one way analysis of variance model using SPSS 20.00 software [10] and indicated by their probability value (P). Differences among treatments were determined using Tukey's b test and indicated by the superscripts a, b, c, d. All data are presented as means \pm SE and level of significance was declared at P<0.05.

Results

The means for physiological indices i.e., RT, RR & PR at different treatments are presented in Table. 1. There was significant increase in rectal temperature on all the days at 35 and 40 °C as compared to 25 and 30 °C (P<0.05) and highest RT was recorded on day 14 at 40 °C (38.98±0.05). On other days at 25 and 30 °C no significant variation was recorded for RT (P>0.05). Similar to RT, the respiration rates were also elevated during 35 and 40 °C treatments as compared to lower exposure temperatures (P<0.05). Higher RR was observed on, day 14 at 35 °C, days one and 21, respectively at 40 °C [RR: breaths/min: 56.50±17.17; 82.00±3.46 & 58.00±6.00]. Pulse rate did not varied statistically at different days at 25, 30 or 35 °C but a positive increase was noted at 40 °C as compared to 25 and 30°C (P<0.05).

Table 1: Physiological variables, RT, RR & PR at different treatments in buffaloes (Means±SE)

Parameter		Time	25°C	30°C	35°C	40°C
	Day 1	M	37.54±0.04	37.27±0.16 a	37.70±0.14	37.73±0.07
		A	37.58±0.17	37.58±0.22	38.05±0.08b	38.76±0.18 ^b
Rectal temperature	D 14	M	37.22±0.31 a	37.10±0.33a	37.59±0.12 ^b	37.87±0.06
(°C)	Day 14	A	37.23±0.15 a	37.60±0.15	37.99±0.15	38.98±0.05 b
	Day 21	M	37.53±0.15	37.69±0.13	37.84±0.12	37.57±0.16
	Day 21	A	37.63±0.07	37.87±0.17	38.04±0.08 b	38.22±0.06 b
	Day 1	M	11.00±1.00a	9.00±1.00 a	14.00±1.41 a	20.00±2.30a
Dii		A	14.00±1.15 a	15.00±1.00 a	25.00±4.35	82.00±3.46 ^b
Respiration Rate	Day 14	M	10.50±0.95 a	12.00±1.63 a	18.00±2.58 b	13.50±1.50
(breaths/min)		A	14.00±1.15 a	23.00±5.25	56.50±17.17 ^b	37.50±16.64
(breams/mm)	Day 21	M	13.00±1.91 a	20.00±2.82	20.50±1.70	19.00±4.12 a
		A	16.00±1.63 a	18.00±1.15 a	46.00±9.30 ^b	58.00±6.00 ^b
	Day 1	M	45.00±3.41	46.00±2.00	48.00±4.89	51.00±3.00
Pulse Rate (beats/min)		A	36.00±2.82a	39.00±1.91	44.00±4.32	46.00±2.58
	Day 14	M	49.50±1.50	51.00±3.00	50.50±3.94	53.50±2.50 ^b
		A	36.00±1.63 a	36.00±1.63 a	48.00±5.88	41.00±1.91
	Day 21	M	44.50±3.09	52.00±2.82	48.00±0.00	57.00±3.00 b
		A	45.00±2.51	48.00±2.82	41.00±4.12	41.00±2.51

^{*}Means with different superscript a, b vary significantly within row/column for respective parameter (P<0.05)

The results of biochemical estimation for electrolytes; enzymes, triglyceride and reactive oxygen molecules; protein metabolism are presented in Table 2, 3 and 4, respectively. Serum sodium did not varied significantly at 25 °C (P>0.05), but was lower at 30 °C as compared to 35 °C and 40°C

(P<0.05). Similarly, higher values for potassium were obtained at 40 °C than at 25 °C (P<0.05), while no significant changes were noted at 30 and 35 °C (Table 2). There was no notable variation for calcium and chloride during exposure at any of the treatments (P>0.05).

Table 2: Serum electrolyte profile at different treatments in buffaloes (Means±SE)

Parameter		Time	25 °C	30 °C	35 °C	40 °C
	Day 1	M	143.47±6.68	146.26±3.04a	145.88±4.50a	134.37±4.32
Sodium	Day 1	A	132.02±3.84	112.84±3.41 ^b	126.01±11.80	117.71±13.39
(mmol/L)	Day 21	M	133.09±5.13	147.35±4.64a	138.73±6.94	129.82±4.43
	Day 21	A	124.50±1.78	123.51±8.83	115.34±3.43	145.94±2.94a
	Day 1	M	4.46±0.20	5.23±0.26	4.66±0.34	5.30±0.29
Potassium		A	5.37±0.60	4.48±0.33	5.92±0.44	5.26±0.42
(mmol/L)	Day 21	M	4.55±0.16	5.75±0.42	4.95±0.14	6.85±0.26 a
		A	4.21±0.17 b	4.88±0.38	4.74±0.09	4.90±0.21
Calcium (mmol/L)	Day1	M	1.94±0.25	2.36±0.28	2.13±0.14	2.25±0.16
		A	1.97±0.23	2.56±0.28	2.37±0.36	2.69±0.27
	Day 21	M	2.09±0.16	2.28±0.19	1.93±0.19	2.45±0.11

		A	2.10±0.14	2.75±0.29	2.38±0.25	2.06±0.18
Choride (mmol/L)	Day 1	M	73.41±6.11	101.35±3.19	92.82±3.60	78.74±8.52
		A	69.88±8.67	113.76±9.72	109.30±10.96	99.43±18.80
	Day 21	M	102.04±8.00	93.48±0.54	79.32±12.56	88.43±4.01
		A	96.49±13.88	115.03±10.93	103.00±2.90	107.11±3.80

^{*}Means with different superscript a, b vary significantly within row/column for respective parameter (P<0.05)

Superoxide dismutase enzyme was highest on day 1 at 40 °C but decreased again on day 21. Significant increase in SOD activity were observed at 40 °C as compared to other temperatures (P<0.05). No concurrent significant increase was recorded for reactive oxygen species [ROS] during exposure at any days at different treatments (Table 3). Serum AST levels increased significantly on day 21 at 35 °C as compared to 25, 30 and 40 °C (P<0.05), but, no such variation was observed for serum ALT at any of the

temperatures during exposure (P>0.05). Serum lipid derivatives like cholesterol and triglycerides did not varied amongst different days at any treatment during exposure (P>0.05), but numerically lower and higher values were obtained for triglyceride and cholesterol on day 21 at 35 and 40°C during exposure than at 25 and 30 °C, respectively [triglyceride: 0.69 ± 0.08 ; 0.66 ± 0.05 vs 0.74 ± 0.12 ; 0.80 ± 0.06 & cholesterol: 7.48 ± 2.86 ; 6.79 ± 0.94 vs 4.45 ± 0.09 ; 5.26 ± 0.46].

Table 3: Serum enzymes, ROS and lipid profile at different treatments in buffaloes (Means±SE)

Parameter		Time	25 °C	30 °C	35 °C	40 °C
SOD	D 1	M	141.12±15.69 ^b	208.74±40.37	128.82±8.82 b	334.83±35.20a
	Day 1	A	219.77±10.84	262.71±21.80	212.42±12.43 b	222.31±25.46
(U/mL)	Day 21	M	178.55±37.84 b	159.28±6.75 b	146.99±33.82 b	167.68±6.71 b
	Day 21	A	212.85±13.60	196.18±61.78ba	190.11±10.76 b	188.27±37.09
DOG	Day 1	M	0.05 ± 0.00	0.04 ± 0.00	0.05±0.00	0.04±0.00
ROS	Day 1	Α	0.05 ± 0.00	0.04 ± 0.00	0.04±0.00	0.06±0.00
(mg H ₂ O ₂ /dL)	Day 21	M	0.05 ± 0.00	0.04 ± 0.00	0.04±0.00	0.06±0.00
/uL)	Day 21	Α	0.04 ± 0.00	0.04 ± 0.00	0.06±0.00	0.06±0.00
	Doy 1	M	147.37±11.72a	142.50±15.00 a	174.85±14.83	175.62±14.88
AST	Day1	Α	119.85±20.45 a	121.88±20.59 a	147.34±18.99 a	138.78±9.76 a
(U/L)	Day 21	M	162.01±2.85	191.28±9.49	224.14±15.22 ^b	160.21±26.24
		A	124.81±3.48 a	131.34±8.76 a	154.32±8.83 a	138.33±3.72 a
	Day 1	M	66.27±20.57	78.48±14.67	127.67±24.45	99.06±12.67
ALT		Α	46.65±13.31	52.18±14.90	109.56±31.71	112.32±11.83
(U/L)	Day 21	M	92.79±25.33	105.69±15.87	129.06±28.41	113.72±22.24
		A	75.34±25.55	83.98±18.58	111.98±32.39	101.26±22.55
	Day 1	M	3.60±0.44	4.70±0.75	4.36±0.40	4.87±0.45
Cholesterol		Α	3.76±0.38	4.81±1.17	4.75±0.32	5.65±0.50
(mmol/L)	D 21	M	2.56±0.61	5.49±0.54	12.61±2.85	4.69±0.63
	Day 21	Α	4.45±0.09	5.26±0.46	7.48±2.86	6.79±0.94
Triglyceride	Day 1	M	1.02±0.04	1.05±0.26	0.97±0.08	0.87±0.04
		Α	0.73±0.04	0.86±0.11	0.56±0.05	0.66±0.02
(mmol/L)	Doy 21	M	0.94±0.09	1.01±0.14	0.91±0.01	0.94±0.08
	Day 21	A	0.74±0.12	0.80 ± 0.06	0.69±0.08	0.66±0.05

^{*}Means with different superscript a, b vary significantly within row/column for respective parameter (P<0.05)

It was observed that serum total protein, albumin and globulin did not altered at any of the treatments during the study (Table 4). Similarly, no significant variation was recorded for

serum urea and creatinine during exposure at different temperatures in buffaloes (P>0.05).

Table 4: Serum protein, albumin, globulin, urea and creatinine levels at different treatments in buffaloes (Means±SE)

Parameter		Time	25°C	30 °C	35°C	40°C
	D 1	M	63.19±6.27	64.41±7.60	61.07±5.11	70.95±1.77
Total protein	Day 1	A	60.27±7.36	56.51±3.84	56.21±4.72	58.80±4.05
(gm/L)	Day 21	M	61.50±6.77	60.44±4.94	70.99±5.03	64.18±7.38
	Day 21	A	52.96±5.05	64.84±4.34	60.02±7.14	79.62±2.64
	Doy 1	M	25.43±2.68	30.28±2.18	32.26±2.15	29.91±2.98
Albumin	Day 1	A	19.35±1.72	23.44±1.12	30.66±4.00	34.71±3.72
(gm/L)	Day 21	M	29.11±1.85	30.34±2.75	29.53±1.78	31.27±4.26
		A	23.92±2.43	29.05±3.83	32.05±1.14	30.53±7.24
	Day1	M	37.76±8.19	34.13±7.10	28.81±3.69	41.03±2.03
Globulin		A	40.92±6.98	33.07±3.05	25.54±5.04	24.09±5.52
(gm/L)	Day 21	M	32.38±6.69	30.09±7.58	41.45±6.31	49.09±5.35
		A	29.03±4.23	35.79±4.73	27.96±7.55	28.64±9.17
Urea (mmol/L)	Day 1	M	14.97±0.84	10.54±0.55	11.01±2.05	12.43±1.39
		A	13.52±2.11	11.09±1.02	10.21±2.00	8.54±1.68
	Day 21	M	13.41±1.02	15.35±0.57	11.26±2.88	12.39±1.28

		A	13.82±0.92	8.48±1.82	8.61±1.44	9.89±2.84
Creatinine	Day 1	M	1.24±0.22	0.89±0.27	0.95±0.06	1.05±0.24
		A	0.75±0.07	1.00±0.09	1.38±0.20	1.19±0.19
(mmol/L)	(mmol/L) Day 21	M	1.13±0.22	1.47±0.12	1.15±0.26	0.99±0.25
		A	1.02±0.10	1.34±0.36	1.22±0.13	1.19±0.07

Discussion

Immediate response to acute heat stressor is secretions of catecholamines and activation of thermoregulatory mechanisms. Respiration rate increased on all the days (P<0.05) at higher temperatures of 35 °C and 40 °C as compared to 25 °C and 30 °C, which indicated evaporative cooling was activated to dissipate the excess heat load. Similarly, other studies have confirmed increased RR in animals during heat stress which confirm present findings [12, ^{13]}. There was concurrent increase in rectal temperature in buffaloes on all the days at 35 °C and 40 °C than lower exposure treatments (P < 0.05). This increment in core body temperature confirms slow, accumulating heat load on the animals and that thermoregulation was altered. Although evaporative cooling was activated, but the raise in RT could not be prevented completely, thus increasing the core body temperature during HS [14, 15]. As compared to RR and RT, the cardiovascular systems response is variable during HS. It can be altered by changes in metabolism, catecholamines and autonomic nervous system [16]. The increased PR during HS than ambient temperatures in this study might be due to increased thermoregulatory activity or higher catecholamine secretions [17, 18]. The physiological responses are immediately activated during HS allowing essential biochemical modulations necessary for stress adaptation. As the RR, RT and PR were increased till day 21 in buffaloes, it can be concluded that more than 21 days of acclimatization is essential for systemic stress adaptation.

Serum sodium decreased on day one at 30 °C during exposure, thereafter an increase was recorded at 35 and 40 °C (P<0.05). This reflects active sodium conservation, probably under aldosterone action which is responsible for sodium conservation and potassium excretion in sweat, urine during HS. Another mechanism can be an isotonic expansion of extracellular fluid volume resulting in higher sodium levels in buffaloes [19]. Similarly, potassium levels increased (P<0.05) at 40 °C than at 25 °C, which might be due to higher exchange from intracellular compartment to extracellular fluids to replace and conserve sodium during thermal stress. This finding of higher potassium levels during HS in buffaloes is contradictory to previous reports and needs further exploration [20]. No alterations in chloride and calcium concentration at any treatments might be due to optimum retention for the minerals during HS period and our results are supported by similar findings in Omani sheep and Holstein *cattle* exposed to hot environmental conditions [21, 22].

Superoxide dismutase increase significantly on day 21 at 30 °C, didn't varied at 35 °C and again increased on day 1 at 40 °C. This increase in SOD levels in buffaloes indicates elevated antioxidant activity during hyperthermia to prevent/reduce cellular damage [23, 24]. Contrastingly, the ROS levels did not altered (*P*>0.05) during HS in our study possibly due to optimum cellular antioxidant activity, which contradicts the previous results in cattle [25]. Although, serum aspartate aminotransferase [AST] decreased on day 21 during exposure at 35 °C, but was higher than at 25, 30 and 40 °C, respectively, probably due to increased gluconeogenesis during HS. This increase at 35 °C might be due to immediate

hepatic/cellular injury or stress, followed by gradual adaptation at 40 °C as recorded in present study. Our findings are supported by other workers who also reported, an increased AST activity during heat stress period [26, 27]. However no such variation was recorded for serum ALT at any treatment during the study, probably indicating that ALT might not be an accurate biomarker of HS in cattle/buffaloes. The concentrations of serum cholesterol and triglyceride did not altered at any exposure temperature in buffaloes, particularly during HS, probably due to optimum energy balance, thus sparing the lipid metabolites. Similarly, no variation in serum triglyceride was noted in crossbred Holstein cattle exposed to different temperatures [28].

In present study, lack of variation for total protein, albumin, globulin, urea and creatinine (Table 4) in buffaloes, might be due to optimum hepatic protein metabolism during HS period. It also reflects balanced nitrogen metabolism as serum urea and creatinine levels did not increased. Our findings are corroborated by other similar animals studies reporting no significant variation for protein and other protein metabolites at optimum and heat stress conditions [29-31].

Conclusion

Due to our geographical location and semi modernized practices till date, the animals are definitely exposed to heat stress [moderate/severe; acute/chronic]. Immediate alteration is seen in physiological variables, followed by changes in enzymes, electrolytes, hormones and overall metabolism during HS. This enables the animals to sustain the acute or chronic heat stress conditions but at the cost of higher energy expenditure for thermoregulation. The present study confirmed the negative impact of HS on buffaloes and also established that a period of more than 21 days is essential for physio-metabolic acclimatization and homeostasis during thermal stress.

References

- 1. Moran JB. Heat tolerance of Brahman cross, buffalo, bauteng and shorthorn steers during exposure to sun and as a result of exercise. Australian Journal of Experimental Agriculture. 1973; 24:775-782.
- 2. Gudev D, Popova RS, Moneva LP, Aleksiev P, Peeva TZ, Penchev P *et al.* Physiological indices in buffaloes exposed to sun. Archiva Zootechnica. 2007; 10:127-132
- 3. Intergovernmental Panel on Climate Change. IPCC. 2018. 2018- special report: global warming of 1.5 °C and Climate change synthesis report.
- 4. Marai IFM, Haeeb AAM. Buffalo's biological functions as affected by heat stress A review. Livestock Science. 2010; 127:89-109.
- 5. West JW. Effects of heat-stress on production in dairy cattle. Journal of Dairy Science. 2003; 86:2131-2144.
- 6. Ganaie AH, Gauri Shanker, Nazir AB, Ghasura RS, Mir NA, Wani SA *et al.* Biochemical and Physiological Changes during Thermal Stress in Bovines. Journal of Veterinary Science and Technology. 2013; 4(1):1-6.
- 7. West W. Effects of heat-stress on production in dairy cattle. Journal of Dairy Science. 2003; 86:2131-2144.

- 8. NRC. Nutrient Requirements of Dairy Cattle, National Academy Press, Washington, DC, 2001.
- 9. Madesh M, Balasubramanian KA. Microtiter plate assay for superoxide dismutase using MTT reduction by superoxide. Indian Journal of Biochemistry and Biophysics. 1998; 35:184-8.
- 10. Brambilla G, Fiori M, Archetti LI. Evaluation of the oxidative stress in growing pigs by microplate assays. Journal of Veterinary Medical Science. 2001; 48:33-38.
- 11. SPSS, Inc. SPSS (Statistical Package for Social Sciences) for Windows© 1997; Version 20.00 SPSS Inc Chicago, IL. 1993.
- 12. Brown-Brandl TM, Eigenberg RA, Nienaber JA, Hahn GL. Dynamic response indicators of heat stress in shaded and nonshaded feedlot cattle, Part 1: analyses of indicators. Biosystem Engineering. 2005; 90:451-462.
- 13. Yadav B, Singh G, Wankar A. Acclimatization dynamics to extreme heat stress in crossbred cattle. Biological Rhythm Research. 2019; DOI: 10/1080/09291016.2019.16127.
- 14. Sejian V, Bhatta R, Soren NM, Malik PK, Ravindra JP, Prasad C et al. Introduction to Concepts of Climate Change Impact on Livestock and Its Adaptation and Mitigation. In Climate Change Impact on Livestock: Adaptation and Mitigation; Sejian, V., Gaughan, J., Baumgard, L., Prasad, C., Eds.; Springer: New Delhi, India, 2015, 1-23.
- 15. Mehla K, Magotra A, Choudhary J, Singh AK, Mohanty AK, Upadhyay RC *et al.* Genome wide analysis of heat stress response in zebu (sahiwal) cattle. Gene. 2014; 533:500-507.
- 16. Janzekovic M, Mursec B, Janzekovic I. Techniques of measuring heart rate in cattle. Tehnicki Vjesnik. 2006; 13:31-37.
- 17. Banerjee D, Ashutosh. Circadian changes in physiological responses and blood ionized sodium and potassium concentrations under thermal exposure in Tharparkar and Karan Fries heifers. Biological Rhythm Research. 2011; 42(2):131-139.
- 18. Krishnan G, Singh G, Shukla DC. The Effect of Electrolyte Supplementation on Physiological Responses in Heat Stressed Male Buffalo Calves. Indian Journal of Animal Science. 2009; 79(1):34-37.
- 19. Nessim MG. Heat-induced biological changes as heat tolerance indices related to growth performance in buffaloes. PhD Thesis, Faculty of Agriculture Animal Shams University, Cairo, Egypt, 2004.
- 20. Hooda OK, Singh G. Changes in Serum Electrolytes, Mineralsand Physiological Responses in Buffalo Heifers Exposed to High Temperature in a Psychrometric Chamber. Indian Journal of Animal Nutrition. 2009; 26(4)
- 21. Srikandakumar A, Johnson EH, Mahgoub O. Effect of heat stress on respiratory rate, rectal temperature and blood chemistry in Omani and Australian Merino sheep. Small Ruminant Research. 2003; 49(2):193-198.
- Chaiyabutr N, Chanpongsang S, Suadsong S. Effects of evaporative cooling on the regulation of body water and milk production in crossbred Holstein cattle in a tropical environment. International Journal of Biometeorology. 2008; 52:575-585.
- 23. Kumaraguruparan R, Balachandran C, Murali MB, Nagini S. Altered oxidant–antioxidant profile in canine mammary tumors. Veterinary Research Communications.

- 2005; 29:287-296.
- 24. Kumar A, Singh G, Kumar BVS, Meur SK. Modulation of antioxidant status and lipid peroxidation in erythrocyte by dietary supplementation during heat stress in buffaloes. Livestock Science. 2011; 138:299-303.
- 25. Bernabucci U, Ronchi B, Lacetera N, Nardone A. Markers of oxidative status in plasma and erythrocytes of transition dairy cows during hot season. Journal of Dairy Science. 2002; 85:2173-2179.
- 26. Boots LR, Crist WL, Davis DR, Bruin EW, Ludwick IM. Effect of age, body weight, stage of gestation and sex on plasma glutamic oxaloacetic and glutamic pyruvic transaminase activity in immature cattle. Journal of Dairy Science. 1969; 52:211-216.
- 27. Marai IFM, Habeeb AAM, Daader AH, Yousef HM. Effects of diet supplementation and body cooling on Friesian calves reared in high ambient temperature in the Eastern Desert of Egypt. Tropical Animal Health Production. 1997; 4:201-208.
- 28. Chaiyabutr N, Chanpongsang S, Suadsong S. Effects of evaporative cooling on the regulation of body water and milk production in crossbred Holstein cattle in a tropical environment. International Journal of Biometeorology. 2008; 52:575-585.
- 29. Scharf B, Carroll JA, Riley DG, Chase CC, Jr Coleman SW, Keisler DH *et al.* Evaluation of physiological and blood serum differences in heat-tolerant (Romosinuano) and heat-susceptible (Angus) Bos taurus cattle during controlled heat challenge. Journal of Animal Science. 2010; 88(7):2321-36.
- 30. Yokus B, Cakir UD. Seasonal and Physiological Variations in Serum Chemistry and Mineral Concentrations in Cattle. Biological Trace Element Research. 2006; 109:255-266.
- 31. Nonaka I, Takusari N, Tajima K, Suzuki Higuchi T, Kurihara KM. Effects of high environmental temperatures on physiological and nutritional status of prepubertal Holstein heifers. Livestock Science. 2008; 113:14-23.