



E-ISSN: 2320-7078

P-ISSN: 2349-6800

JEZS 2020; 8(1): 992-996

© 2020 JEZS

Received: 01-11-2019

Accepted: 05-12-2019

**Arjun Gurung**

Research Scholar, Institute of  
Agriculture and Animal Science,  
Tribhuvan University, Chitwan,  
Nepal

**Rameshwor Pudasaini**

Assistant Professor, Institute of  
Agriculture and Animal Science,  
Tribhuvan University,  
Kathmandu, Nepal

**Bishal Gaire**

Research Scholar, Institute of  
Agriculture and Animal Science,  
Tribhuvan University,  
Kathmandu, Nepal

**Subash Sitaula**

Research Scholar, Institute of  
Agriculture and Animal Science,  
Tribhuvan University,  
Kathmandu, Nepal

**Corresponding Author:**

**Arjun Gurung**

Research Scholar, Institute of  
Agriculture and Animal Science,  
Tribhuvan University, Chitwan,  
Nepal

## Host preference of Bihar hairy caterpillar *Spilosoma obliqua* in laboratory condition

**Arjun Gurung, Rameshwor Pudasaini, Bishal Gaire and Subash Sitaula**

### Abstract

Bihar hairy caterpillar (*Spilosoma obliqua*) is a serious polyphagous insect pest. An experiment was conducted to determine the host preference of Bihar hairy caterpillar in Entomology laboratory, Lamjung Campus at room temperature from March to April, 2019. The experiment was conducted in CRD with seven treatments namely; Cotton (*Gossypium hirsutum*), Cauliflower (*Brassicae oleraceae var. botrytis*), Cabbage (*Brassicae oleraceae var. capitata*), Broccoli (*Brassicae oleraceae var. italica*), Mulberry (*Morus alba*), Castor (*Ricinus communis*), Broad bean (*Vicia faba*). Five pure cultured larvae were set in each plastic jar containing respective plant sample. All the host plants showed highly significant difference at 1% level of significance. The first instar larvae fed highly in cabbage (13.2g) and least on castor (6.07g) where as in the later stages the larvae preferred broad bean more followed by the cabbage and other crucifers where as the least consumption was seen in mulberry and castor throughout the entire larval period. The time taken to reach the pupal stage was shortest in broad bean and cabbage i.e. within 14 days conversely the pupal stage was reached a lot later in castor and mulberry i.e. after 18 days.

**Keywords:** Bihar hairy caterpillar, crucifer, feeding, host preference, lepidoptera & pupation

### Introduction

Bihar hairy caterpillar, *Spilosoma obliqua* (Walker) is a polyphagous pest that comes under lepidoptera order of class insect consisting of chewing type of mouthpart which is confined to the oriental region known to cause severe damage to several crops of agricultural and horticultural importance. Besides Nepal, this pest has also been reported in various other countries too such as India, Myanmar, Pakistan, China (Kabir & Khan, 1968; Singh & Seghal, 1992) [19, 30]. *S. obliqua* is a sporadic in nature. The adult female lays eggs in clusters on lower surface of the epidermis of the leaves in the crowded condition. The young caterpillars are gregarious and they feed on green, soft tissues and leave behind the veins, thus skeletonizing the leaves whereas in later stages the larvae eat the leaves from the margin thereby giving the leaves of plant an appearance of net or web. *S. obliqua* larvae pass through six instars (Adsule & Kadam, 1979) [2]. (Gyawali, 1988) [15] found that the sixth instars larvae are the most damaging ones. They are capable of attacking 126 plant species belonging to more than 24 plant families which includes a number of economically important plant as such cereals, grams, oilseeds and pulses (Singh & Varatharajan, 1999) [44]. Jute has been one of its major hosts causing severe economic damage (Gupta & Bhattacharya 2008) [14]. The pest causes yield loss up to 30% in jute (Bandyopadhyay *et al.* 2014) [43]. This pest are profoundly found to infest bean, cotton, groundnut, brinjal, cabbage, cauliflower, linseed, pea, soybean and other leguminous crops (Kabir, 1966) [18]. Of late it has also become the major pest of mulberry (Geetha & Marimadaiah, 2006) [42]. This pest has become the second most serious pest of cole crops mostly cabbage being preferred as reported by Bhatia *et al.*, (1995) [41]. Kadam *et al.*, (1995) [2] observation recorded that the damage by *Spilosoma obliqua* and concluded that the fourth to sixth in stars are highly destructive in nature and shall require chemical control. Host plant species are those which slow or increase the development of the insect which have considerable relevance to the management method Tange *et al.*, (2013) [34]. These are plants upon which an organism lodges and subsists (Marriam-Webster). By this definition host plants are those biologically active plants that provide food and shelter to caterpillar to complete their lifecycle and reproduce. During the process of evolution, plants and insects have developed different strategies to dodge SSSeach other harmful effects. Plants respond to those harmful effects of insect through various morphological, biochemical, and molecular mechanisms.

Direct defense mechanism includes plants affecting host plant preference or survival and reproductive success of the insects, and indirectly by natural enemies. These defenses are wide-ranging, highly dynamic, and mediated by biochemical mechanisms. Secondary metabolites as such terpenoids, alkaloids, anthocyanins, phenols, and quinones are produced in response to the damage of the crops which ultimately affect the survival and feeding habit of the pest (Howe & Jander 2008; Arimura *et al.* 2009; War *et al.* 2012) [12, 1, 37]. Different insects have their own type of host preferences.

Agriculture is the backbone of the Nepal economy because 65.6% of Nepalese's population depends on agriculture for the livelihood. However, it is disheartening to know that Nepalese's economic growth is not keeping up to the expectations because its agricultural growth is lagging. The state is worsened by the fact that 35% of the produce goes to the waste in pre and post harvest operation due to the insect pest attacks in the field and storage (Palikhe, 2002) [40]. The 3<sup>rd</sup> instars onward larvae of this pest is the most damaging one and cause heavy reduction in the yield (Hussain & Begum, 1995; Gupta & Bhattacharya, 2008) [37, 14]

Being a sporadic and polyphagous pest it has wide range of the host, the rate of larval survival and development vary greatly on different host plants. Nevertheless, polyphagy offers *S. obliqua* an avenue for the population survival and persistence. However in Nepal the polyphagous nature of *S. obliqua* larvae has been scarcely studied. The knowledge on host plant preference and vulnerable stages will ultimately help to combat the pest in terms of effective measures of controlling its population and reducing the pest damage.

### Materials and Methods

The experiment was conducted in entomology lab of lamjung campus during March-April 2019. The study site is located at 28 7' to 28 10' North and longitude from 84 24' to 84 28' east at an altitude of 675m above sea level.

This place has humid sub-tropical climate. The total annual rainfall in this area is reported as 2800 mm maximum and minimum temperature is 28-30 °C and 20-22 °C respectively. The experiment was conducted in CRD with seven treatments namely; Cotton (*Gossypium hirsutum*), Cauliflower (*Brassicae oleraceae var. botrytis*), Cabbage (*Brassicae oleraceae var. capitata*), Broccoli (*Brassicae oleraceae var. italica*), Mulberry (*Morus alba*), Castor (*Ricinus communis*), Broad bean (*Vicia faba*).

### Collection and Rearing of insect

A pure culture is a population of cells or multicellular organisms growing in the absence of other species or types which originate from a single cell or single organism, in which the cells are genetic clones of one another (Morello).

The eggs of Bihar hairy caterpillar was collected from research field of Gunjanagar, Chitwan. The eggs, was then allowed to hatch in the laboratory of department of Entomology, IAAS lamjung campus and the larvae was given a 10% sugar solution as a food for their growth and development. The male and female adults of *Spilosoma obliqua* were separated and allowed to mate in a breeding cage. On the completion of mating female adults started laying eggs in clusters. Thus, the eggs obtained were left undisturbed for hatching. In this way, pure culture larvae were used in the experiments.

### Inoculation of stock culture

Larvae were starved for 12 hrs before its inoculation. Five pure cultured larvae were inoculated in a plastic jar containing the respective sample leaves. Fresh sample leaves was collected from the periphery of lamjung campus and each of 5g was put in a plastic jar and the leaves was changed in each 2 days. The jar was then covered with fine muslin cloths for the aeration. The moisture loss of the leaves was checked as a control by placing the respective sample leaves in a jar without larvae on each 2 days.

### Data collection

The data was taken on every 2 days i.e. when the fresh leaves was placed. The leaves was weighed on every 2 days till the larvae reached to the pupation. The other plastic jar containing only the leaves was also weighed to determine the moisture lost. The food consumption by the larvae was calculated as;

Food consumption: 5g-(weighed of leaves after feeding by larvae + weighed of leaves due to moisture loss)

### Statistical analysis

All the data were recorded and tabulated based on replications and treatments with the help of MS Excel. Recorded data were managed and then subjected to analyze by using R-studio. Microsoft word 2010 was used for word processing, Excel for tabulating the data and finding the correlation between different parameters, R studio for running statistical analysis. Calculation of the significant critical difference at 1% level of significance was made by mean comparisons.

### Results and Discussion

The present study dealt with the host suitability and food consumption of *Spilosoma obliqua* (Walker). The results of different experiments are discussed below:

### Food consumption

**Table 1:** Influence of different host on consumption index of *Spilosoma obliqua*

Treatment	Mean consumption index of larvae									
	Day2	Day4	Day6	Day8	Day10	Day12	Day14	Day16	Day18	Day20
Cotton	9.05 <sup>c</sup>	2.52 <sup>e</sup>	12.81 <sup>bc</sup>	15.09 <sup>bcd</sup>	26.24 <sup>b</sup>	24.57 <sup>b</sup>	6.28 <sup>e</sup>	24.95 <sup>d</sup>	37.9 <sup>c</sup>	45.71 <sup>b</sup>
Cauliflower	11.13 <sup>abc</sup>	7.87 <sup>d</sup>	13.87 <sup>bc</sup>	17.13 <sup>bc</sup>	40.6 <sup>a</sup>	22.6 <sup>b</sup>	41.27 <sup>a</sup>	60 <sup>a</sup>	60.6 <sup>a</sup>	26.27 <sup>d</sup>
Cabbage	13.2 <sup>a</sup>	8.8 <sup>d</sup>	15.33 <sup>b</sup>	17.6 <sup>b</sup>	37.6 <sup>a</sup>	26.93 <sup>b</sup>	26.93 <sup>c</sup>	60 <sup>a</sup>	61.6 <sup>a</sup>	41.27 <sup>c</sup>
Broccoli	11.67 <sup>ab</sup>	18.8 <sup>a</sup>	11.73 <sup>c</sup>	14.2 <sup>cde</sup>	38.87 <sup>a</sup>	36.05 <sup>a</sup>	33.53 <sup>b</sup>	37.53 <sup>c</sup>	60.2 <sup>a</sup>	11.87 <sup>e</sup>
Mulberry	10.81 <sup>bc</sup>	16.05 <sup>b</sup>	11.19 <sup>cd</sup>	13.19 <sup>de</sup>	24.33 <sup>b</sup>	21.48 <sup>b</sup>	12.67 <sup>d</sup>	13.86 <sup>f</sup>	9.62 <sup>d</sup>	28.86 <sup>d</sup>
Castor	6.07 <sup>d</sup>	10 <sup>d</sup>	8.2 <sup>d</sup>	11.47 <sup>e</sup>	21.47 <sup>b</sup>	21.47 <sup>b</sup>	7.13 <sup>e</sup>	18.8 <sup>e</sup>	34.13 <sup>c</sup>	1.8 <sup>f</sup>
Broad bean	9.76 <sup>bc</sup>	13.29 <sup>c</sup>	21.76 <sup>a</sup>	24 <sup>a</sup>	45.23 <sup>a</sup>	39.05 <sup>a</sup>	34.28 <sup>b</sup>	51.43 <sup>b</sup>	51.43 <sup>b</sup>	51.43 <sup>a</sup>
F value	***	***	***	***	***	***	***	***	***	***
LSD	2.37	2.37	2.99	3.06	9.86	6.65	4.45	3.62	6.49	3.88
CV%	13.24	12.27	12.58	10.84	16.81	13.83	10.98	5.4	8.22	7.49

The feed consumption of *S. obliqua* on seven different host plants are presented in the above table. All the host plants showed highly significant difference at 1% level of significance. The first instar larvae fed highly in cabbage and least on castor where as in the later stages the larvae preferred broad bean more followed by the cabbage and other crucifers where as the least consumption was seen in mulberry and castor throughout the entire larval period. The higher consumption rate in broad bean and cabbage was due to the higher nutrient contents such as crude protein and fat (Taleb, 2008)<sup>[33]</sup>.

Lower consumption in the castor was due to the fact that castor has toxic and anti feedant property which changes the internal metabolic processing of the larvae that donot inhibits the food uptake rather slow down (Ali M. Ali & Ahmed Ibrahim, 2018)<sup>[38]</sup>.

As the concentrations and proportion of nutrients differ greatly among different host plants, this variation determines the regulation of the pest population (Slansky & Rodriquez 1987; Schoonhoven *et al.*, 2005)<sup>[32, 27]</sup>.

Roy & Barik (2012, 2013)<sup>[25]</sup> found that when 4 host plants were tested with respect to food quality, various parameters as such the growth duration, consumption rate, utilization efficiency, development time, longevity, fecundity and survival of *Diacrisia casignetum* Kollar were found to be significant.

**Days to pupation**

The result on influence of host plants on its days to reach the pupation has been discussed below:

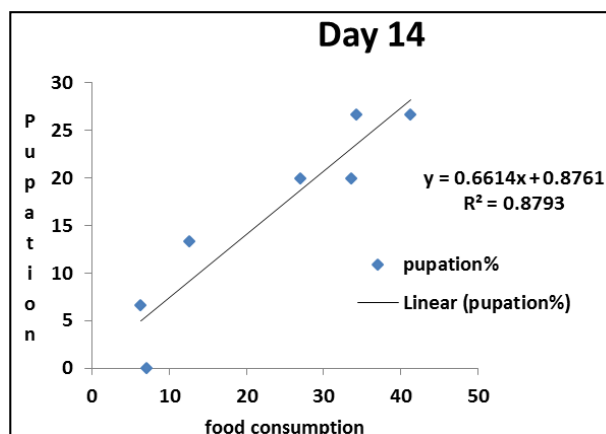
**Table 2:** Influence of different host on the pupation of *Spilosoma obliqua*

Treatment	Pupation index of larvae				
	Day 14	Day 16	Day 18	Day 20	Day 22
Cotton	6.67 <sup>a</sup>	20 <sup>bc</sup>	33.33 <sup>bc</sup>	60 <sup>cde</sup>	100 <sup>a</sup>
Cauliflower	26.67 <sup>a</sup>	40 <sup>a</sup>	60 <sup>a</sup>	86.67 <sup>ab</sup>	100 <sup>a</sup>
Cabbage	20 <sup>ab</sup>	40 <sup>a</sup>	60 <sup>a</sup>	93.33 <sup>a</sup>	100 <sup>a</sup>
Broccoli	20 <sup>ab</sup>	26.67 <sup>abc</sup>	53.33 <sup>a</sup>	73.33 <sup>bc</sup>	100 <sup>a</sup>
Mulberry	13.33 <sup>abc</sup>	26.67 <sup>abc</sup>	26.67 <sup>c</sup>	53.33 <sup>de</sup>	73.33 <sup>b</sup>
Caster	0 <sup>c</sup>	13.33 <sup>c</sup>	26.67 <sup>c</sup>	46.67 <sup>e</sup>	73.33 <sup>b</sup>
Broad bean	26.67 <sup>a</sup>	33.33 <sup>ab</sup>	46.67 <sup>ab</sup>	66.67 <sup>cd</sup>	100 <sup>a</sup>

The above table shows that the pest took shortest days to pupate in the broad bean and cabbage followed by other crucifers (started within 14 day) such that all the 5 larvae reached pupation within 22 days in these respective host plants whereas in the castor and mulberry days to reach the

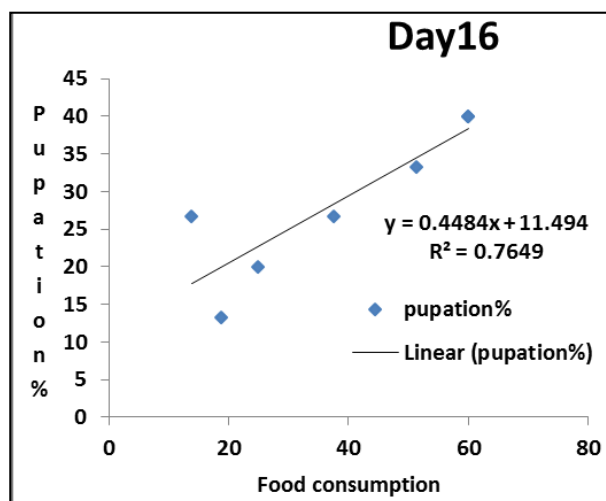
pupation was found to be longest such that all the larvae reached pupation in 24 days or more.

Correlation between the food consumption by larvae and the pupation is given below: After 14 days



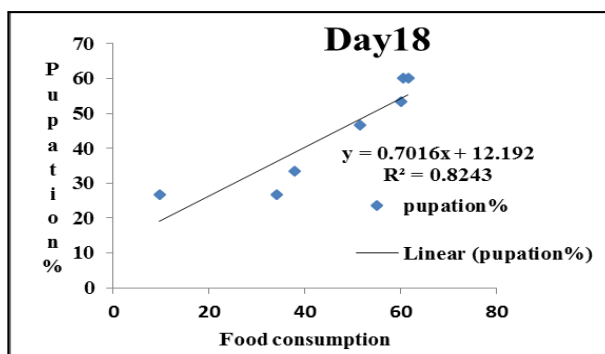
**Fig 1:** Correlation between food consumption and pupation% after 14 days

The correlation between them was found to be 0.93755 which is close to the 1 such that they are nearly perfectly correlated.



**Fig 2:** Correlation between food consumption and pupation % after 16 days

The correlation between them was found to be 0.8741 thus has a positive correlation.



**Fig 3:** Correlation between food consumption and pupation% after 18 days

The correlation was found to be 0.91 thus are nearly perfectly correlated.

Therefore, the above correlation graph shows that the food consumption of the pest in the larval developmental period

has a positive effect on the transformation of these larvae to the pupation phase as well as the pupal weight of the pest (Goel & Kumar, 1983)<sup>[10]</sup>.

This finding is also supported by the fact that the food types helps in the increment of developmental rate which allows this species to reach the pupal period faster and finally to the adulthood and increase their population size (Miller *et al.*, 2014)<sup>[39]</sup>.

As the larval period increases, the consumption index and approximate digestibility is also increased and inversely related is the efficiency of conversion of digested food (ECD) and efficiency of conversion of ingested food (ECI) with the ageing of the larval (Kumar *et al.*, 2011)<sup>[22]</sup>.

During the later stage, the increase in ECD is attributed to the fact that the more food is utilized in the assimilation of various tissues resulting in the faster pupation as reported by the findings of Reynolds *et al.*, (1985)<sup>[24]</sup> and Sharma *et al.*, (1999)<sup>[29]</sup>.

The presence of various compounds as such phenol and peroxidase in the host plants adversely affect the larval growth of the insects thereby delaying the pupation period of the insects Gotyal *et al.*, (2015)<sup>[11]</sup>.

Different growth parameter as such the larval period, pupal period is in direct relation with the presence of nutrients in the host plants as reported by Hockensmith *et al.*, 1986<sup>[17]</sup>.

### Summary and Conclusion

The statistical result showed that the different host plants showed highly significance results at 1% level of significance. The pest, polyphagous and causing more damage have high level of preferences on cabbage and other crucifers in initial instars followed by the broad bean and cabbage in the later instar of larvae till formation of pupa such that the pupation was reached early in the same host plants that the pest fed the most.

The experiment concluded that the larvae preferred highly on the cabbage, broad bean and other crucifers whereas the least preferred host plants were mulberry and castor.

Thus, the effective measures for controlling pest population are to be adopted so as to reduce its damage extent either by chemical use or by botanical extracts as an alternative.

### Acknowledgments

We are thankful to the advisory committee for providing better guidance in the completion of the research work also the entomology lab of lamjung campus for providing necessary equipments required for the successful completion of the study and all my friends, juniors and my family for their continuous help and support.

### References

1. Arimura GI, Matsui K, Takabayashi J. Chemical and molecular ecology of herbivore-induced plant volatiles: proximate factors and their ultimate functions. *Plant Cell Physiology*. 2009; 50:911-923.
2. Adsule VM, Kadam MV. Studies on the bionomics of Bihar hairy caterpillar, *Spilosoma obliqua* Walker on sunflower (*Helianthus annuus* L.). *Journal of Maharashtra Agricultural University*. 1979; 4(3):249-252.
3. Chand P. Host preference of *Diacrisia obliqua* Walk. (Lepidoptera: Arctiidae) in the field. *Science & Culture*. 1975; 41(12):604-606.
4. Dahale PS, Puri SN, Bilapate GG. Biometrics, growth and development of *Diacrisia obliqua* Walk. On different hosts. *Journal of Maharashtra Agricultural University*. 1988; 13(2):164-168.
5. Deshmukh PD, Rathore YS, Bhattacharya AK. Larval survival of *Diacrisia obliqua* Walk. On several plant species. *Indian Journal of Entomology*. 1979; 41(1):5-12.
6. Deshmukh PD, Rathore YS, Bhattacharya AK. Effect of temperature on the growth and development of *Diacrisia obliqua* (Walk.) on five host plants. *Indian Journal of Entomology*. 1982; 44(1):21-33.
7. Dhaliwal JS. Development of *Diacrisia obliqua* Walk. On some fodder crops. *Journal of Research*. Punjab Agricultural University. 1988; 25(4):616-618.
8. Djou YW. A limbean leaf-hopper, *Diacrisia obliqua* Walk. (Lepidoptera: Arctiidae). *Lingnan Science of Journal*. 1938; 17(4):639-645.
9. Gargave VG, Katiyar OP. Some new host of *Diacrisia* at Raipur, M. P. *Indian Journal of Horticulture*. 1971; 24(4):316.
10. Goel SC, Kumar A. Morphology and weight loss in the pupae of *Diacrisia obliqua* Walk. (Lepidoptera: Arctiidae). *Annals of Entomological society of America*. 1983; 76(3):437-440.
11. Gotyal BS, Selvaraj K, Meena PN, Satpathy S. Host plant resistance in cultivated jute and its wild relatives towards jute hairy caterpillar *Spilosoma obliqua* (Lepidoptera: Arctiidae). *Florida Entomological Society*. 2015; 98(2):721-727.
12. Howe GA, Jander G. Plant immunity to insect herbivores. *Annual Review of Plant Biology*. 2008; 59:41-66.
13. Gupta BM, Dabi RK, Gupta SC, Sharma SK. Effect of host plants on the larval development of *Diacrisia obliqua* Walk. *Journal of Entomological Research*. 1979; 3(2):224-226.
14. Gupta G, Bhattacharya AK. Assessing toxicity of post emergence herbicides to the *Spilarctia obliqua* Walker (Lepidoptera: Arctiidae). *Journal of Pest Science*. 2008; 81:9-15.
15. Gyawali BK. Damage and yield loss of soybean from hairy caterpillar (Lepidoptera: Arctiidae). *Quarterly Newsletter*. 1988; 31(3):35-41.
16. Haque M, Karim ANMR, Alam S. Preliminary study or varietal reaction of cultivars to leaf defoliators. *Tropical Grain Legume Bull*. 1984; 29:35-37.
17. Hockensmith PE, Devine TL, Legg DE, Rodriguez JG. Energy Consumption and Food Utilization of the Indian Meal Moth (Lepidoptera: Pyralidae) on Different Corn Genotypes. *Journal of the Kansas Entomological Society*. 1986; 59(4):598-603.
18. Kabir AKMF. Phase variation in the jute hairy caterpillar, *Diacrisia obliqua* Walk. (Lepidoptera: Arctiidae). *Sindh University Research Journal*. 1966; 11:1-3.
19. Kabir AKMF, Khan SA. Bioassay of some insecticides for the control of jute, hairy caterpillar, *Diacrisia obliqua* Walk. *Indian Journal of Science and Technology*. 1968; 6(1-2):131-138.
20. Kabir AKMF, Khan SA. Biology and life-history of jute hairy caterpillar, *Diacrisia obliqua* Walk. In East Pakistan. *Pakistan Journal of Zoology*. 1969; 1(1):45-48.
21. Katiyar OP, Mukharji SP, Lal L. Effect of certain natural food plants on the larval development of *Diacrisia obliqua* Walk. *Indian Journal of Entomology*. 1976; 38(2):130-133.
22. Kumar VP, Harinath P, Ramana SP. Lifecycle and larval performance of the joker butterfly *Byblia ilithyia*

- (Lepidoptera: Rhopalocera: Nymphalidae) from southern Andhra Pradesh. Bulletin of Pure and Applied Sciences. 2011; 31(1):1-6.
23. Pandey ND, Yadava DR, Teotia TPS. Effect of different food plants on the larval development of *Diacrisia obliqua* Walk. Indian Journal of Entomology. 1968; 30(3):229-234.
  24. Reynolds SE, Nottingham SF, Stephans AE. Food and water economy and its relation to growth in fifth instar larvae of tobacco hornworm, *Manduca sexta*. Journal of Insect Physiology. 1985; 31:119-127.
  25. Roy N, Barik A. The impact of variation in foliar constituents of sunflower on development and reproduction of *Diacrisia caignetum* Kollar (Lepidoptera: Arctiidae). Psyche, 2012. 2012: article 812091.
  26. Roy N, Barik A. Influence of four host plants on feeding, growth and reproduction of *Diacrisia caignetum* (Lepidoptera: Arctiidae). Entomological Science. 2012; 16:112-118.
  27. Schoonhoven LM, Loon JJAV, Dicke M. Insect-Plant Biology. Oxford University Press, Oxford, United Kingdom, 2005.
  28. Sharma HC, Sujana G, Rao DM. Morphological and chemical components of resistance to pod borer, *Helicoverpa armigera* in wild relatives of pigeon pea. Arthropod-Plant Interactions. 2009; 3:151-161.
  29. Sharma R, Sheikher C, Sharma KC. Quantitative intake and utilization of cauliflower leaves by cabbage white butterfly, *Pieris brassicae* (L) (Lepidoptera: Pieridae). Entomon. 1999; 24(2):143-147.
  30. Singh S, Seghal SS. Studies on growth and development of *Spilosoma obliqua* (Walker) on different food plants. Indian Journal of Entomology. 1992; 54(4):471-482.
  31. Singh G, Singh I. Comparative development and survival of Bihar hairy caterpillar *Spilosoma obliqua* (Walker) at different temperatures and on different varieties of sunflower at Ludhiana. Indian journal of Agriculture Science. 1993; 63(7):447-450.
  32. Slansky JRF, Rodriquez JG. Nutritional Ecology of Insects, Mites, Spiders, and Related Invertebrates. John Wiley & Sons, Inc., New York, USA, 1987.
  33. Taleb MA. Growth indices of jute hairy caterpillar, *Spilarctia obliqua* on host plant spectrum. Bangladesh Journal of Entomology. 1998; 8(1, 2):9-19.
  34. Tanga CM, Ekesi S, Govender P, Mohamed SA. Effect of six host plant species on the life history and population growth parameters of *Rastrococcus iceryoides* (Hemiptera: Pseudococcidae). Florida Entomologist. 2013; 96:1030-1041.
  35. Usha Rani P, Jyothsna Y. Biochemical and enzymatic changes in rice as a mechanism of defense. Acta Physiologiae Plantarum. 2010; 32:695-701.
  36. Yadava RP, Singh R. Studies on the larval development of Bihar hairy caterpillar in relation to some host plants at Dholi (North Bihar). Science & Culture. 1997; 43(5):233-234.
  37. War AR, Paulraj MG, Ahmad T, Buhroo AA, Hussain B, Ignacimuthu S. Mechanims of plant defense against insect herbivore. Plant Signaling and Behaviour. 2012; 7:1306-1320.
  38. Ali M. Ali, Ahmed Ibrahim. Castor and camphor essential oils alter hemocyte populations and induce biochemical changes in larvae of *Spodoptera littoralis* (Boisduval) (Lepidoptera: Noctuidae). Journal of Asia-Pacific Entomology. 2018; 21:631-637.
  39. Miller SL, Mensah B, Wubah D. The effect of food type on the Development rate of the Danaid Eggfly. Journal of Young Investigators. 2014.
  40. Palikhe B. Challenges and options of pesticide use: In context of Nepal. Journal of Landschaftsokologie and Umweltforschung. 2002.
  41. Bhatia R, Verma A.K. Seasonal incidence of major insect pest of cabbage in Himachal Pradesh. Annals of Agricultural Research. 1995; 16(3):278-281.
  42. Geethabai M, Marimadaiah B. The spiraling whitefly, *Aleurodicus disperses* Russell (Homoptera: Aleyrodidae, A pest of mulberryin Karnataka. Sericologia. 2006; 41:121-127.
  43. Bandyopadhyay S, Gotyal B S, Satpathy S, Selvaraj K, Tripathi A N, Ali N. Synergistic effect of Azadiractin and Bacillus thuringiensis against bihar hairy caterpillar, *Spilosoma obliqua* Walker. Biopesticides International. 2014; 10(1):71-76.
  44. Singh YR, Varatharajan R. Hot range of bihar hairy caterpillar, *Spilosoma obliqua* (Walker) (Arctiidae, Lepidoptera). Hexapoda. 1999; 11:65-74.