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### Biology, feeding potential and functional response of *Coccinella septempunctata* L. against *Aphis* gossypii Glover infesting cucumber

#### Deeksha Kumari, SC Verma and PL Sharma

#### Abstract

The study on biology, feeding potential and functional response of the coccinellid, Coccinella septempunctata L. to Aphis gossypii Glover was carried out in the Biological Control Laboratory of Department of Entomology, Dr YS Parmar University of Horticulture and Forestry, Nauni, Solan, Himachal Pradesh, India. The study reveals that average incubation period of C. septempunctata was 3.20 days. The mean duration of 1st, 2nd, 3rd, 4th larval instar, total larval, pre pupal and pupal period was 2.24, 3.12, 3.32, 4.10, 13. 34, 1.90 and 3.90 days, respectively. Females laid on an average of 189.60 eggs in their life span of 43.84 days. Mean number of aphids consumed per day by 1st, 2nd, 3rd and 4th instars were 15.08, 31.54, 55.94 and 63.10 aphids, respectively. The predator consumed on an average 579.92 aphids during the entire larval stage. The number of aphids consumed increased with advancement of larval instars and increased in prey density. The mean number of aphids consumed by 4th instar of C. septempunctata was 23.30 at prey density of 25 and 65.40 aphids at the prey density of 125, respectively. Adults of C. septempunctata consumed maximum aphids (78.30) at prey density of 125. Predator's attack rate was lowest (0.075  $h^{-1}$ ) for the 1<sup>st</sup> instar and highest (0.175  $h^{-1}$ ) with the adult stage followed by attack rate of 4<sup>th</sup> instar (0.152 h<sup>-1</sup>). All the stages as well as adult of C. septempunctata followed a Type II functional response. Thus, the 4<sup>th</sup> instar and adult of C. septempunctata could be utilised in biological control of A. gossipii.

Keywords: Aphis gossypii, biology, coccinellid, functional response, attack rate, handling time

#### Introduction

The cotton aphid, Aphis gossypii Glover (Homoptera: Aphididae) is a cosmopolitan, polyphagous species infesting 569 host plants in tropical, subtropical and temperate regions (Sakaki and Sahragard, 2011; Singh et al., 2014; Tazerouni et al., 2016) <sup>[22, 29, 32]</sup>. Both nymphs and adults damage the crop by sucking the cell sap, resulting in curling of leaves or appearance of discoloured spots on the foliage. As the aphid become more abundant in juvenile stage, the plants gradually wilts, become yellowish to brownish and ultimately die. Heavy infestation of the aphid, results in reduction of the yield and quality of fruit by contaminating with honeydew which encourages sooty mould that disturbs the normal physiology of the leaves and also act as vector of 76 plant viruses (Escriu et al., 2000; Pinto et al., 2008; Srinivasulu et al., 2010; Bayoumy, 2011) <sup>[7, 20, 30, 4]</sup>. For the control of *A. gossypii*, many insecticides have been recommended (Mhaske *et al.*, 2007; Misra, 2013) <sup>[13, 14]</sup> but, the indiscriminate use of synthetic pesticides leads to the development of insecticide resistance (Tomquelski et al., 2007; Denholm and Devine, 2013; Sarwar et al., 2014) <sup>[33, 5, 22]</sup>, pest resurgence, insecticide residue, environment pollution and killing of natural enemies. Cotton aphid, A. gossypii developed resistance against many insecticides (Herron and Wilson, 2010; Seyedebrahimi et al., 2015; Pan et al., 2018) [8, 25, 19] which forces the farmers to increase the dose and/or frequency of pesticide application which further aggravates the problem (Denholm and Devine 2013)<sup>[5]</sup>. Therefore, there is a need to find out some alternatives of chemical pesticides to protect cucumber from infestation of cotton aphid (Kianpour et al., 2010) [11]. Under such circumstances, biological control can be alternative, ecofriendly and sustainable approach to control this pest (Joshi et al., 2010)<sup>[9]</sup>. A large number of predators of A. gossypii have been reported to naturally suppress the aphid populations in different cropping systems belonging to coccinellidae, syrphidae, chrysopidae, and hemerobiidae (Ebert and Cartwright, 1997; Ahmadov and Hasanova, 2016)<sup>[6, 1]</sup>. Before using a predator in a biological control programme, it is essential to evaluate its predatory efficiency against the target species.

Biology and predatory potential of different predators against *A. gossypii* on different crops have been studied by many workers in India (Nehare *et al.*, 2004; Omkar and Pervez, 2004; Nagamallikadevi *et al.*, 2012; Naruka *et al.*, 2017; Sharma *et al.*, 2017) <sup>[17, 18, 15, 16, 27]</sup>. However, the information on the biological parameters and feeding potential of *coccinella septempunctata* against *A. gossypii* on cucumber under mid-hill conditions of Himachal Pradesh is lacking. Therefore, the objective of the study was to develop a better strategy for the biological control of *A. gossypii* on cucumber under mid-hill conditions of Himachal Pradesh.

#### Materials and Methods

#### **Raising of cucumber crop**

The cucumber crop was raised in the Experimental Farm of the Department of Entomology by following the recommended package of practices of vegetable crops (Anonymous, 2014)<sup>[3]</sup>. The variety of the cucumber was *Cucumis sativus* var. *malini*.

#### Laboratory rearing of Cotton aphid, A. gossypii

The cotton aphid, *A. gossypii* was collected from cucumber field and was maintained in the polyhouse throughout the year on cucumber plants. Cucumber leaves having nymphs were brought to laboratory for the further experiments. Before maturity of the old plants new plants were planted to maintain the culture of cotton aphid throughout the year.

#### Developmental biology of C. septempunctata

Developmental biology of the C. septempunctata was studied on the cotton aphid, A. gossypii on cucumber leaves in culture room fixed at 12:12 h photoperiod, at  $25\pm 1^{\circ}C$  and  $70\pm 5$  per cent relative humidity. In this experiment, one pair of newly emerged adults were confined in the Petri plates containing wet blotting paper on base. Cucumber leaves (6×6cm) having mixed population of second and third instar of A. gossypii were placed on the wet blotting paper in Petri plates. The experiment was carried out with ten replications. The food was changed daily and the Petri plates under observation were examined at 24 h interval for egg laying and moulting of the nymph till all nymphs developed into adults. The duration of development of egg, each nymphal instar, total nymphal duration, pre pupal, pupal and total duration from egg to adult emergence were recorded. After emergence, the adults were identified as male and female under a stereozoom microscope and confined in pairs in Petri plates containing ample food. The pre-oviposition period and fecundity were recorded and then observations were recorded regularly until the first egg was laid which indicated the end of pre-oviposition period. Oviposition period, post- oviposition period, male and female longevity were recorded.

#### **Feeding potential**

Feeding potential of different developmental stages of C. *septempunctata* was studied against *A. gossypii*. Newly hatched larvae of the same stage were shifted individually to a single Petri plate having counted number of aphids of equal age. In order to select the aphids of equal age the cucumber leaves were carefully examined under the stereo zoom binocular microscope and the unwanted stages were removed carefully with the help of camel hair brush. The base of the Petri plate was covered with the wet blotting paper. The experiment was carried out in the insect culture room fixed at 12:12 h photoperiod, at  $25\pm 1^{\circ}$ C and  $70\pm 5$  per cent relative

humidity. The data on aphid consumption by the predator was recorded after every 24 h till they enter the next stage and the feed of the predator was changed. The experiment was replicated ten times. The number of aphids consumed per day by each larval instar was calculated. The data obtained was analysed by using one-way analysis of CRD without any transformation through online statistical software OPSTAT (Sheron *et al.*, 1998) <sup>[28]</sup>.

#### Functional response of C. septempunctata

Functional response of different stages of C. septempunctata to varying densities of A. gossypii was studied in Biological Control Laboratory of the Department of Entomology. Five densities (each replicated 10 times) of nymphs of second or third stage of A. gossypii were offered to the individuals of different stages of C. septempunctata on cucumber leaves in Petri plates separately. The experimental arena consisted of a 6 cm cucumber leaf disk placed in a Petri plate. Prey densities offered were 10, 15, 20, 25 and 30 for first instar; 15, 30, 45, 60 and 75 for second instar; 20, 40, 60, 80 and 100 for third instar; 25, 50, 75, 100 and 125 for fourth instar as well as for adult female of C. septempunctata. In order to select the aphids of equal age the cucumber leaves were examined carefully under the microscope and the excess density and unwanted life stages were carefully removed with the help of the camel hair brush. The Petriplates were placed in culture room at 25 °C, 70± 5 per cent relative humidity and a photoperiod of 12:12 (L:D)h. Observations were made after every 24 h. For studying the functional response of adults only females of C. septempunctata were used.

#### Data analysis of functional response

The data generated on functional response were analysed using the predation data for 24h. A logistic regression between proportion of prey consumed and prey density offered were fitted to determine the shape (e.g., type II or type III) of functional response:

$$N_{e}/N_{0} = \frac{\exp(p_{0} + p_{1}N_{0} + p_{2}N_{0}^{2} + p_{3}N_{0}^{3})}{1 + \exp(p_{0} + p_{1}N_{0} + p_{2}N_{0}^{2} + p_{3}N_{0}^{3})}$$

Where  $N_e$  is the number of prev eaten;  $N_0$  is the initial number of preys,  $p_0$  is intercept,  $p_1$  is linear coefficient,  $p_2$  is quadratic coefficient and p<sub>3</sub> is cubic coefficient. Significant negative and positive linear coefficients (i.e.  $p_1$ ) from the regression indicate type II or type III functional response, respectively (Juliano, 2001) <sup>[10]</sup>. After determining the functional response the random predator equation of Rogers (1972) <sup>[21]</sup> was used to describe the functional responses. The form of the equation is as follows:  $N_e = N_0 [1 - exp (aT_hN_e - aT)]$ . Where  $N_e$  is the number of prey eaten per predator, N<sub>0</sub> is the prey density offered, T is the duration of the experiment (24h), T<sub>h</sub> is the handling time i.e. time required by the predator to pursue, kill and digest the prey and a is the predation coefficient or predators attack rate. The value a/ Th indicates the effectiveness of predator which was calculated by dividing a by  $T_h$  and maximum predation rate  $K = T/T_h$  was also calculated.

#### **Results and Discussion**

#### Developmental biology of C. septempunctata

Data contained in Table 1 reveal that the average incubation period of *C. septempunctata* was 3.20 days. Subail *et al.* (1999) <sup>[31]</sup> recorded that the incubation period of *C.* 

septempunctata was of 3.53 days which was nearly close to the present study. The mean duration of 1st, 2nd, 3rd and 4th larval instar was 2.24, 3.12, 3.32 and 4.10 days, respectively. Total larval, pre pupal and pupal period was 13.34, 1.90 and 3.90 days, respectively. The present findings agree with the findings of Suhail et al. (1999)<sup>[31]</sup> who observed that the total larval and pupal period of C. septempunctata was 11.58 and 3.64 days, respectively. Similarly, Ali et al. (2017) [2] recorded that larval period of C. septempunctata preying on A. gossypii was 13.33 days and pupal period was 6.33 days. In the present investigation, time taken from egg to adult emergence was 22.66 days which was nearly same as reported by Sattar et al. (2008) <sup>[24]</sup> where egg to adult emergence period was 25.6 days. The male longevity of the adult beetles of C. septempunctata was 35.80 days. Females laid an average of 189.60 eggs in their life span of 43.84 days. Pre oviposition, oviposition and post oviposition periods were 11.38, 21.54 and 8.78 days, respectively (Table 1). In a similar study Sharma et al. (2013)<sup>[26]</sup> recorded that male and female longevity of adult beetles of C. septempunctata was 37.71 and 45.29 days, respectively at 25 °C temperature on A. fabae. Slight difference in findings of both the studies may be due to the different aphid species offered. Ali et al. (2017)<sup>[2]</sup> observed that on an average adult beetles of C. septempunctata survived for 33 days on cotton aphid.

**Table 1:** Duration of developmental stages (day's  $\pm$  SE) of C.septempunctata reared on A. gossypii

Development Stage	C. septempunctata (days ± SE)
Incubation period	$3.20 \pm 0.18$
1 <sup>st</sup> instar	$2.24 \pm 0.05$
2 <sup>nd</sup> instar	$3.12 \pm 0.22$
3 <sup>rd</sup> instar	$3.32 \pm 0.23$
4 <sup>th</sup> instar	$4.10 \pm 0.31$
Total larval period	$13.34 \pm 0.28$
Pre-pupa	$1.90 \pm 0.11$
Pupa	$3.90 \pm 0.12$
Egg-adult emergence	$22.66 \pm 0.25$
Female longevity	$43.84 \pm 3.41$
Male longevity	$35.80 \pm 1.03$
Pre oviposition period	$11.38\pm0.92$
Oviposition period	$21.54 \pm 1.12$
Post oviposition period	$8.78\pm0.90$
Fecundity (Eggs/female)	$189.60 \pm 7.46$

## Predatory potential of C. septempunctata against A. gossypii

Predatory potential of *C. septempunctata* against *A. gossypii* was studied by assessing the feeding potential of different developmental stages and functional response of the *C. septempunctata* to varying densities of cotton aphid.

#### Feeding potential of C. septempunctata

The number of the aphids consumed increased at each

successive stages of the predator. The larva of the predator consumed on an average of 33.80, 98.70, 185.70 and 258.72 aphids during 1st, 2nd, 3rd and 4th instars, respectively (Table 2). Mean number of aphids consumed per day by 1<sup>st</sup>, 2<sup>nd</sup>, 3<sup>rd</sup> and 4<sup>th</sup> instars was 15.08, 31.54, 55.94 and 63.10 aphids/day, respectively. The predator consumed on an average 579.92 aphids during the entire larval stage. Female and male beetles consumed 3376.94 and 2457.84 aphids in their entire life span and the average number of aphids consumed per day by each female or each male beetle was 77.02 or 68.66 aphids. respectively. The results recorded in the present study were supported by the study of Suhail et al. (1999) <sup>[31]</sup> who recorded that 1<sup>st</sup>, 2<sup>nd</sup>, 3<sup>rd</sup> and 4<sup>th</sup> in stars of *C. septempunctata* consumed on an average 15.23, 36.40, 42.23 and 47.15 individuals of A. gossypii/day, respectively. Ali et al. (2017) <sup>[2]</sup> recorded that at temperature of 18 <sup>0</sup>C and 65 per cent relative humidity  $1^{st}$ ,  $2^{nd}$ ,  $3^{rd}$  and  $4^{th}$  instar of C. septempunctata consumed 55.33, 102.66, 172.00 and 315.00 aphids, respectively. Slight differences in the results of the present study and those of Ali et al. (2017)<sup>[2]</sup> may be due to the stage of the aphids offered to the predator and the temperature conditions provided during the experimental period. The present results also agree with the findings of Sattar *et al.* (2008) <sup>[24]</sup> who recorded that the average number of the A. gossypii consumed by an adult female of C. septempunctata per day was 77.80.

 Table 2: Feeding potential of different developmental stages of C.

 septempuncata against A. gossypii

Stage	Number of a Consum	Number of aphids consumed/ day		
_	Mean ± SE	Range	Mean ± SE	Range
1 <sup>st</sup> instar	$33.80\pm3.70$	25-45	$15.08\pm0.47$	13-17
2 <sup>nd</sup> instar	$98.70 \pm 4.10$	88-108	$31.54\pm0.83$	28-34
3 <sup>rd</sup> instar	$185.70 \pm 7.75$	155-198	$55.94 \pm 1.57$	50-61
4 <sup>th</sup> instar	$258.72 \pm 10.76$	235-287	$63.10\pm4.14$	58-78
1 <sup>st</sup> - 4 <sup>th</sup> instar	$579.92 \pm 12.33$	538-604	$43.48 \pm 3.65$	36-52
Adult female	$3376.94 \pm 58.56$	3156-3490	$77.02 \pm 1.52$	73-83
Adult male	$2457.84 \pm 21.07$	2375-2490	$68.66 \pm 1.36$	65-73

#### Functional response of C. septempunctata

The data presented in Table 3 reveal that cubic polynomial fit between prey density offered (N) and proportion of prey consumed (Na/N) by all the stages of the predator resulted significant negative linear coefficients (1<sup>st</sup> instar: -0.05; 2<sup>nd</sup> instar: -0.02; 3<sup>rd</sup> instar: -0.008; 4<sup>th</sup> instar: -0.005 and adult: -0.002) confirming a Type II functional response for all the developmental stages of the predator. Previously many workers have reported a Type II functional response in coccinellids (Xue *et al.* 2011; Kumar *et al.* 2014; Sharma *et al.* 2017) <sup>[34, 12, 27]</sup> to their prey. A predator with a Type II functional response has an inverse density-dependent action against its prey.

Table 3: Parameters of polynomial regression between proportions of prey consumed (Na/N) and initial prey density (N) of C. septempunctata

Dradator stage	Parameter	Coefficient	t volue	р
Predator stage		Mean ± SE	t- value	ſ
1 <sup>st</sup> instar	PO	$1.05 \pm 0.13$	8.23	< 0.001
	P1	$-0.05 \pm 0.02$	-2.73	0.023
	P2	$0.003 \pm 0.0009$	2.67	0.025
	P3	$-0.00005 \pm 1.8E-05$	-2.66	0.026
2 <sup>nd</sup> instar	PO	$1.11 \pm 0.03$	33.05	< 0.001
	P1	$-0.02 \pm 0.003$	-7.63	< 0.001
	P2	$0.0004 \pm 7E-05$	5.46	< 0.001

	P3	$-0.000003 \pm 4.5E-07$	-6.02	< 0.001
2rd in store	PO	$1.04 \pm 0.03$	30.92	< 0.001
	P1	$-0.008 \pm 0.002$	-4.15	0.0024
5 mistai	P2	$0.0001 \pm 0.00001$	3.56	< 0.001
	P3	$-0.0000009 \pm 1.9E-07$	-4.68	0.0012
4 <sup>th</sup> instar	PO	$1.04\pm0.02$	67.85	< 0.001
	P1	$-0.005 \pm 0.0006$	-9.12	< 0.001
	P2	0.00005±8.4E-06	5.71	< 0.001
	P3	$-0.0000003 \pm 3.9E-08$	-7.61	< 0.001
Adult	PO	$0.99 \pm 0.01$	98.81	< 0.001
	P1	$-0.002 \pm 0.0004$	-4.84	< 0.001
	P2	$0.00002 \pm 6.2E-06$	3.49	< 0.001
	P3	$-0.0000002 \pm 4.2E-08$	-4.50	0.0014

P0= Constant, P1= Linear coefficient, P2= Quadratic coefficient, P3= Cubic coefficient

Data presented in Table 4 reveal that the number of aphids consumed by 1<sup>st</sup> instar of *C. septempunctata* was maximum at the prey density of 30 which was statistically on par with the prey density of 25. But, the proportion of prey consumed was

maximum at the lowest prey density (73.00%) and proportion of prey consumed decreased with increasing prey density. The proportion of prey consumed was minimum at the prey density of 30 (48.30%).

Instars/Adult	Prey density	Mean number of aphids consumed (Mean ± SE)	Proportion of aphids consumed (%)
	10	$7.30\pm0.15^{d}$	73.00
	15	$9.80 \pm 0.13^{\circ}$	65.30
1 st :	20	$12.90 \pm 0.28^{b}$	64.50
1 <sup>st</sup> instar	25	$14.30\pm0.15^{\rm a}$	57.20
	30	$14.50 \pm 0.17^{a}$	48.30
	CD (p= 0.05)	0.53	
2 <sup>nd</sup> instar	15	$12.90 \pm 0.10^{d}$	86.00
	30	$21.00\pm0.26^{\rm c}$	70.00
	45	$29.90 \pm 0.31^{b}$	66.44
	60	$33.00 \pm 0.26^{a}$	55.00
	75	$33.10 \pm 0.23^{a}$	44.13
	CD (p= 0.05)	0.70	
3 <sup>rd</sup> instar	20	$18.60\pm0.16^d$	93.00
	40	$33.60 \pm 0.31^{\circ}$	84.00
	60	$50.00 \pm 0.33^{b}$	83.33
	80	$56.10 \pm 0.23^{a}$	70.12
	100	$56.30 \pm 0.26^{a}$	56.30
	CD (p= 0.05)	0.76	
	25	$23.30\pm0.21^d$	93.20
	50	$41.50 \pm 0.34^{\circ}$	83.00
4th instan	75	$59.70 \pm 0.30^{b}$	79.60
4 <sup>th</sup> instar	100	$65.30\pm0.26^{a}$	65.30
	125	$65.40 \pm 0.22^{a}$	52.32
	CD (p= 0.05)	0.77	
Adult female	25	$23.90\pm0.10^d$	95.60
	50	$46.50\pm0.16^{\rm c}$	93.00
	75	$65.70 \pm 0.26^{b}$	87.60
	100	$78.10 \pm 0.28^{a}$	78.10
	125	$78.30 \pm 0.21^{a}$	62.64
	CD(p=0.05)	0.61	

The number of aphids consumed by the  $2^{nd}$  instar of *C.* septempunctata at prey densities of 15, 30, 45, 60 and 75 was 12.90, 21.00, 29.90, 33.00 and 33.10, respectively. The number of aphids consumed at the prey density of 75 were statistically on par with the prey density of 60. The maximum proportion of the aphids were consumed at the prey density of 15 (86.00%) and minimum proportion was consumed at prey density of 75 (44.13%). The  $3^{rd}$  instar of *C. septempunctata* consumed 18.60, 33.60, 50.00, 56.10 and 56.30 aphids on the prey densities of 20, 40, 60, 80 and 100, respectively. The number of aphids consumed at the prey density of 100 was statistically on par with the prey density of 80. The proportion of aphids consumed was maximum (93.00%) at the lowest prey density and minimum (56.30) at the highest prey density. The proportion of the aphids consumed were maximum at the prey density of 25 (93.20%) and minimum at the prey density of 125 (52.32%) (Table 4). The mean number of aphids consumed by 4<sup>th</sup> instar of *C. septempunctata* was 23.30, 41.50, 59.70, 65.30 and 65.40 aphids at the prey densities of 25, 50, 75, 100 and 125, respectively. The number of aphids consumed at the prey density of 100 were statistically on par with the prey density of 80. Prey densities offered to adults of *C. septempunctata* were 25, 50, 75, 100 and 125 and mean number of aphids consumed at these prey densities was 23.90, 46.50, 65.70, 78.10 and 78.30, respectively (Table 4). The number of aphids consumed at the prey density of 125 was

statistically on par with the prey density of 100. The proportion of aphids consumed was highest at the prey density of 25 (96.00%) and lowest at the prey density of 125 (62.64%).

In the present findings, the proportion of prey consumed at higher density decreased which may be due to satiation of the predator at increased number of prey. These findings were in agreement with the findings of Pervez and Omkar (2005) <sup>[18]</sup> who found that there was a decline in the consumption rate of *C. septempunctata* at higher prey densities. The present study (Table 4) reveal that the number of the aphids consumed increased with the increase in prey densities for all the developmental stages of the predator and when satiation was attained by the predator there was no significant increase in the number of aphids consumed and there was no effect of prey density (Omkar and Pervez, 2005) <sup>[18]</sup>. The proportion of the aphids consumed was maximum at the lowest density and minimum at the highest prey density.

The predation efficiency of C. septempunctata against A. increased with the advancement of the gossypii developmental stages of the predator which is numerically illustrated by parameters of attack rate (a) and handling time (T<sub>h</sub>) (Table 9). Predator's attack rate was lowest (0.075  $h^{-1}$ ) for the 1<sup>st</sup> instar and highest (0.175 h<sup>-1</sup>) with the adult stage followed by attack rate of 4<sup>th</sup> instar (0.152 h<sup>-1</sup>). Handling time of the adult beetle (0.205 h<sup>-1</sup>) was statistically on par with the 4<sup>th</sup> instar grub (0.262 h<sup>-1</sup>) and was shorter than rest of the stages. The present study predicts that the 1st, 2nd, 3rd, 4th instar and adult of C. septempunctata could predate a maximum of 26.70, 46.20, 80.27, 91.60 and 117.07 aphids in a 24 h period. Functional response parameters indicated that the adult and  $4^{th}$  instar grubs were the effective stages of C. septempunctata against A. gossypii. In the present investigations, the attack rate of the predator increased and handling time decreased with the gradual increase in stage/ age of predator. Similar to the present investigations, Sharma et al. (2017) [27] also observed that the attack rate of Harmonia dimidiata increased with the advancement of stages against A. gossypii and handling time shortened with the increase in stages of the predator.

 Table 5: Parameters of functional response of C. septempunctata to

 A. gossypii over 24 h period

<b>D</b> radator stage	Parameter				
r reuator stage	a ± SE	$T_h \pm SE$	a/T <sub>h</sub>	K	<b>R</b> <sup>2</sup>
1 <sup>st</sup> instar	$0.075 \pm 0.004^{e}$	$0.903 \pm 0.053^{d}$	0.09	26.70	0.952
2 <sup>nd</sup> instar	$0.109\pm0.003^d$	$0.521\pm0.012^c$	0.21	46.20	0.984
3 <sup>rd</sup> instar	$0.141\pm0.006^c$	$0.281\pm0.008^b$	0.47	80.27	0.989
4 <sup>th</sup> instar	$0.152\pm0.008^{b}$	$0.262\pm0.007^a$	0.58	91.60	0.991
Adult	$0.175 \pm 0.001^{a}$	$0.205 \pm 0.008^{a}$	0.85	117.07	0.995
CD (p= 0.05)	0.014	0.071			

a= Coefficient of attack rate,  $T_{h}{=}$  Handling time, K= maximum theoretical predation rate,  $R^2{=}$  Coefficient of determination

#### Conclusion

*Coccinella septempunctata* can be utilized as a component of integrated pest management programmes for the management of cotton aphid, *A. gossypii* in cucumber under mid-hill conditions of Himachal Pradesh.

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#### References

- 1. Ahmadov BA, Hasanova SS. The role of coccinellids (Coccinellidae) in suppression of the number of *Aphis craccivora* Koch, 1854, and *Aphis gossypii* Glover, 1877. J Ent. Zool. Stud. 2016; 4:234-37.
- Ali A, Memon SA, Mastoi AH, Narejo MN, Azizullah, Afzal M, Ahmed S *et al.* Biology and feeding potential of ladybird beetle (*Coccinella septempunctata* L.) against different species of aphids. Sci. Intl. 2017; 29:1261-63.
- Anonymous. Package of Practices of Vegetable crops. Directorate of Extension Education. Dr. YS Parmar University of Horticulture and Forestry, Nauni, Solan-173230 HP, 2014.
- 4. Bayoumy MH. Foraging behavior of the coccinellid *Nephus includes* (Coleoptera: Coccinellidae) in response to *Aphis gossypii* (Hemiptera: Aphidiae) with particular emphasis on larval parasitism. Environ. Ent, 40:835-43.
- Denholm I, Devine G. Insecticide resistance. Encyclopedia of Biodiversity. 2<sup>nd</sup> ed. Academic Press, Waltham, 2013, 298-307.
- 6. Ebert TA, Cartwright B. Biology and ecology of *Aphis gossypii* Glover (Homoptera: Aphididae). Southwest Entgst. 1997; 22:115-53.
- 7. Escriu F, Perry KL, Garcia-Arenal F. Transmissibility of cucumber mosaic virus by *Aphis gossypii* correlates with viral accumulation. Viral. 2000; 90:1069-72.
- 8. Herron GA, Wilson LJ. Neonicotinoids resistance in *Aphis gossypii* Glover (Aphididae: Hemiptera) from Australian cotton. Australian J Entomol. 2010; 50:93-98.
- 9. Joshi S, Rabindra RJ, Rajendran TP. Biological control of aphids. Journal of Biol. Contl. 2010; 24:185-202.
- 10. Juliano SA. Non-linear curve fitting: Predation and functional response curves. In: Design and analysis of ecological experiments (Scheiner SM and Gurevitch J, eds). Chapman and Hall, New York, 2001, 178-96.
- 11. Kianpour R, Fathipour Y, Kamali K, Naseri B. Bionomics of *Aphis gossypii* (Homoptera: Aphididae) and its predators *Coccinella septempunctata* and *Hippodamia variegata* (Coleoptera: Coccinellidae) in natural conditions. J Agric. Sci. Technol. 2010; 12:1-11.
- Kumar B, Mishra G, Omkar. Functional response and predatory interactions in conspecific and heterospecific combinations of two congeneric species (Coleoptera: Coccinellidae). European J Entomol. 2014; 111:257-65.
- 13. Mhaske BM, Pardesh SR, Bhoite KD, Rasal PN. Biosafety of coccinellids predators and chemical control of wheat aphids. Agric. Sci. Digt. 2007; 27:264-66.
- 14. Misra HP. Newer insecticides for the management of aphid, *Aphis gossypii* Glover in gherkins (*Cucumis anguria* L.) and their effect on the predator, *Coccinella septempunctata* L. Pest Managt Horti. Ecosyst. 2013; 19:123-27.
- Nagamallikadevi M, Undirwade DB, Reddy NB, Ramadevi A, Srasvankumar G. Biology of *Mallada boninensis* (Okamoto) [Chrysopidae: Neuroptera] on Aphids and Neonate Noctuids. Biosci. Trnds. 2012; 6:827-30.
- Naruka P, Meena A, Meena BM. Feeding potential of *Chrysoperla zastrowi* arabica on different prey hosts. J Entomol. Zool. Stud. 2017; 5:608-12.
- 17. Nehare SK, Deotale VY, Deotale RO, Dawane PN. Biology and predatory potential of *Mallada boninensis* (Okamoto) against sucking pests. J Soils and Crops. 2004; 14:427-32.

- Omkar, Pervez A. Functional and numerical responses of *Propylea dissecta* (Col., Coccinellidae). J Appl. Entomol. 2004; 128:140-46.
- 19. Pan Y, Tian F, Wei X, Wu Y, Gao X, Xi J *et al.* Thiamethoxam Resistance in *Aphis gossypii* Glover Relies on Multiple UDP-Glucuronosyltransferases. Frontrs. Physiol. 2018; 9:322-27.
- 20. Pinto ZV, Rezende JAM, Valdir A, Yuki VA, Piedade SMS. Ability of *Aphis gossypii* and *Myzus persicae* to transmit cucumber mosaic virus in single and mixed infection with two poty viruses to zucchini squash summa. *Phytopathol. Botu.* 2008; 34:183-85.
- 21. Rogers D. Random search and insect population models. J Animal Ecol. 1972; 41:369-83.
- 22. Sakaki S, Sahragard A. A new method to study the functional response of *Scymnus syriacus* (Coleoptera: Coccinellidae) to different densities of *Aphis gossypii*. J Asia-Pacific Entomol. 2011; 14:459-62.
- 23. Sarwar MK, Azam I, Iram N, Iqbal W, Rashda A, Anwer F, *et al.* Cotton aphid, *Aphis gossypii* L. (Homoptera; Aphididae); A challenging pest; biology and control strategies: A review. Intnl. J Appl. Biol. Pharmtl. Technol. 2014; 5:288-94.
- Sattar M, Hamed M, Nadeem S. Biology of *Coccinella* septempunctata L. (Coleoptera: Coccinellidae) and its predatory potential on cotton aphids, *Aphis gossypii* Glover (Hemiptera: Aphididae). Pakistan J Zool. 2008; 40:239-42.
- 25. Seyedebrahimi SS, Jahromi KT, Imani S, Naveh VH, Hesami S. Characterization of imidacloprid resistance in *Aphis gossypii* Glover (Hemiptera: Aphididae) in Southern Iran. Turkish J Entomol. 2015; 39:413-23.
- 26. Sharma R, Sood A, Sharma KC, Sood M. Biology and predatory potential of *Coccinella septempunctata* on *Aphis fabae* in the mid hills of Himachal Pradesh. Journal of Insect Science. 2013; 26:220-24.
- 27. Sharma PL, Verma SC, Chandel RS, Shah MA, Gavkare O. Functional response of *Harmonia dimidiata* (fab.) to melon aphid, *Aphis gossypii* Glover under laboratory conditions. Phytoparasitica. 2017; 45:373-79.
- 28. Sheoran OP, Tonk DS, Kaushik LS, Hasija RC, Pannu RS. Statistical Software Package for Agricultural Research Workers. Department of Mathematics Statistics, CCS Haryana Agric. Univ. Hissar, Haryana, India, 1998.
- 29. Singh G, Singh NP, Singh R. Food plants of a major agricultural pest, *Aphis gossypii* Glover (Homoptera: Aphididae) from India: An updated checklist. Intnl. J Life Sci. Biotechnol. Pharma Res. 2014; 3:1-28.
- Srinivasulu M, Sarovar B, Anthony Jhonson AM, Sai Gopal DVR. Association of poty virus with mosaic virus disease of gherkin (*Cucumis anguira* L.) in India. Indian J Microbiol. 2010; 50:221-24.
- 31. Suhail A, Sabir AM, Hussain A, Saeed A. Predatory efficacy of *Coccinella septempunctata* L. on cotton aphid, *Aphis gossypii* Glov. Pakistan J Biol. Sci. 1999; 2:603-05.
- 32. Tazerouni Z, Talebi AA, Fathipour Y, Soufbaf M. Bottom-up effect of two host plants on life table parameters of *Aphis gossypii* (Hemiptera: Aphididae). J Agric. Sci. Technol. 2016; 18:179-90.
- 33. Tomquelski GV, Martins GLM, Papa G. Efeitos dos indutores de resistência acibenzolar- metile silíciona biologia de *Alabama argilacea* (Lepidoptera: Noctuidae) emalgodoeiro. Rev de Agric. 2007; 82:170-75.

 Xue JJ, Fengliang L, Ying C. Functional response of Coccinella septempunctata to Aphis craccivora. Agris. 2011; 37:68-71.

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