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Floral preferences for pollen by leaf cutter bees (Hymenoptera: Megachilidae) in Bangalore, India

SD Pradeepa and VV Belavadi

Abstract

Understanding the floral preferences of solitary bee is *sine qua non* for enhancing the knowledge on bee-flower association. Keeping this in mind, the present study was undertaken to assess the foraging preferences of leaf cutter bees belonging to the family Megachilidae. Floral species were identified by comparing the pollen samples (collected from the nest) with the reference pollen slides prepared beforehand. Further, with the help of scanning electron microscopic analysis (SEM), we written morphological description for all the pollen species collected from megachilid nests. The analysis of the pollen sample collected from 373 nests belonging to species revealed that, the bees preferentially foraged on the flowers of 18 species for pollen. Of this, majority (12 species) belonged to the family Fabaceae and accounted for more than 95 percent of the food provided to brood. The remaining species were from Asteraceae, Lamiaceae, Malvaceae, Convolvulaceae and Acanthaceae.

Keywords: leaf cutter bees, megachilidae, pollen morphology, floral-specialization

1. Introduction

Plants have evolved to protect against excessive pollen harvesting by bees, pollen of certain taxonomic groups possess protective properties that hamper digestion and thus challenge the general view of pollen as an easy-to-use protein source, such flowers narrow their window for access only to specialist bees for pollination ^[1].

Many solitary bees are resource specialists and depend on pollen often from a single plant species or from related species belonging to the same genus, or species of the same family because nutritional composition of pollen is very important in host-plant selection of bees. The composition of amino acids varied strongly among plant species, but taxonomically related species had similar nutritional compositions ^[2]. A study enlightening the bee-flower relationships through imprinting theory, tells that host recognition has a genetic basis in specialist bees ^[3]. But the risk of oligolectic bees declining is more due to its physiological adaptations to the fewer host range, such as short seasons, more or less synchronized with blooming ^[4, 5].

Leaf cutter bees (LCB) (Megachilidae) are solitary bees that nest in existing linear cavities. They are oligolectic ^[6, 7]. Scope located on the ventral aspect of their abdomen helps in better transfer of pollen in such plants which are specially adapted for pollination by LCB ^[8]. These bees are resource specialists have a short foraging range, demanding extreme patch fidelity ^[9, 10]. Hence, pollen from nests can be used for establishing a positive association of different bee species with the available flora in the vicinity ^[11].

The knowledge of palynology can be used as an instrument of multiple research for systematic botany. Pollen characters such as size, shape, colour and exine thickness, and aperture type are found to be considerably important ^[12]. By analyzing the diversity of pollen carried by the bees and by sampling pollen from the nests in different times of the year one can get an idea about the floral diversity and changes in flowering phenology in a given location. The present study was taken up with the hypothesis that examination of pollen from nests of leaf cutter bees (LCBs), would give an idea about their floral specialization.

2. Material and Methods**2.1 Study area**

The study was conducted at the Gandhi Krishi Vignana Kendra (GKVK) campus of the University of Agricultural Sciences, Bangalore, Karnataka state, India. The campus is spread over 526 ha (5.3 Km²) (13°04' 37" N, 77° 34' 39.99" E; 930 msl) and receives a mean annual

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rainfall of 915.8 mm. The study area comes under the eastern dry Agro-climatic zone of Karnataka state and it has diverse vegetation including cultivated crops (agricultural fields), plantations (horticultural blocks), medicinal plants, and several species of garden plants, wild trees, shrubs and weeds (Appendix 1) that would be sources of nectar and pollen for the bees.

2.2 Sampling and identification of pollen

Species of plants flowering in the study area was recorded at monthly intervals and reference pollen slides were prepared from pollen samples collected from freshly opened flowers. For each species a set of slides were prepared by the standard acetolysis method [13, 14, 15]. Photomicrographs of pollen were made using ZEISS Axio Scope A1 Trinocular Microscope and Scanning Electron microscope [16].

These are basically cavity nesting bees that construct a linear nest with a number of brood cells in preformed cavities using various materials like leaves, mud and resins etc. The mother LCB constructs a brood cell by lining the cell walls of the nest cavity with leaf pieces or soil or resin and then provisions this "cell" with a mixture of pollen and nectar above which an egg is laid before sealing the cell. New cells were added in front of completed cells. After completion of the nest, the mother closed the nest entrance by making a thick wall.

In ten selected locations on the campus, where there was LCBs activity, two artificial nesting materials were provided. Each artificial nest consisted of a 30 cm long PVC pipe (15 cm dia) containing 15 hollow reeds (one foot length) of *Ipomea carnea* and wooden blocks with number of holes having a width of 8-10mm (Table 1). Between August 2015 and June 2017, closed nests were collected (512 brood cells from 373 nests) at monthly intervals and in place of each collected nest an empty hollow read was kept for new nest construction. In the study period a total of 373 nests were collected and 512 brood cells were examined. The closed nests were brought to the laboratory and after cutting open the nests, observations were made on characterization and identification of pollen in the brood cells by comparing with pollen reference slides, under Olympus BX40 Trinocular Light Microscope. The Scanning Electronic Microscopic (SEM) studies were employed to provide the details of morphology beyond the resolution of Light Microscope and morphological description were written based on SEM images [14].

3. Results and Discussion

Though the study area had over 150 species of flowering plants, the LCBs preferentially collected pollen from only 18 species of plants of which 12 species belonged to Fabaceae, two species to Asteraceae and one species each to Convolvulaceae, Malvaceae, Lameaceae and Acanthaceae.

3.1 Description of pollen collected by LCBs

The detailed morphology of the pollengrains belonging to the 18 species are given below. Pollen taxa had been given along with SEM images, which are served significantly useful for characterization of species.

Fabaceae

Medicago sativa (Plate-1a, A)

Monad, medium (28µm), isopolar, prolate, polar view triangular, tricalporate, calpi broad and taper to the end and exine microreticulate/porous.

Cajanus cajan (Plate-1a, B)

Monad, medium (31µm), isopolar, prolate, polar view triangular, tricalporate, calpi long and broad at pore, exine thicker and striate reticulate.

Cassia auriculata (Plate-1a, C)

Monad, medium (28µm) isopolar, prolate, spheroidal, tricalporate, calpi long and taper to end, exine fossulate and perforate.

Crotalaria juncea (Plate-1a, D)

Monad, small (19µm), heteropolar, prolate, dumbbell shaped, tri-zonocalporate, calpi long and tapered, exine perforate/coarsely reticulate.

Caesalpinia decapetala (Plate-1a, E)

Monad, large (51 µm), isopolar, prolate, polar view triangular/spheroid, tricalporate, calpi long and broad, exine reticulate.

Cicer arietinum (Plate-1a, F)

Monad, small (19 µm), polar view triangular, oblate, tricalporate, exine striated.

Dolichos lablab (Plate-1b, A)

Monad, small (21 µm), isopolar, prolate, tricalporate, calpi long, exine punctate.

Mimosa pudica (Plate-1b, B)

Tetrad, very small (7µm), oblate/spheroid, inaperturate, exine areola/psilate.

Peltophorum pterocarpum (Plate-1b, C)

Monad, medium (39 µm), isopolar, oblate, tricalporate, calpi broader near pores, calpi broader in center, exine heterobracteate reticulate.

Millettia pinnata (Plate-1b, D)

Monad, small (23 µm), isopolar, oblate, tricalporate, calpi broad near pores, exine regulate and punctate.

Senna siamea (Plate-1b, E)

Monad, medium (40 µm), isopolar, spheroid, tricalporate, calpi broader at center, exine punctate/perforate.

Vigna sinensis (plate-1b, F)

Monad, medium (38 µm), isopolar, polar view triangular, tricalporate, exine homobrachteate reticulate.

Asteraceae

Helianthus annuus (Plate-1c, A)

Monad, small (14 µm), spheroid, tricalporate, exine is echinate, spines are short and sharp.

Guizotia abyssinica (Plate-1c, B)

Monad, medium (29 µm), spheroid, tricalporate' exine is echinate, spines are long with conical base.

Convolvulaceae

Ipomoea sp. (Plate-1c, C)

Monad, large (108 µm), spheroid, pantaporate, exine echinate, blunted spines had funnel base with perforations.

Malvaceae

***Abelmoschus esculentus* (Plate-1c, D)**

Monad large (120 µm), spheroid, pantaporate, exine is echinate, spines are blunted, cylindrical and wider apart.

Lameaceae

***Ocimum sp.* (Plate-1c, E)**

Monad medium (35 µm) spheroid, hexacalplate, calpi broad at middle and tapered towards end, exine is heterobrachate reticulate.

Acanthaceae

***Justicia sp.* (Plate-1c, F)**

Monad, medium (30 µm), prolate, dicalporate, calpi longer than wide, Exine is perforate.

3.2 Foraging preference

During the study period 512 brood cells from 373 nests were analyzed. From these 76.87 per cent brood cells contained pollen grains from only one plant species per nest and 19.96 per cent brood cells had pollen from two plant species per nest (Fig. 1). Whenever the cells had two species of pollen there was a clear demarcated deposition in the cells, which could be due to shortage of pollen of one species in the local area which was compensated by foraging on a taxonomically related plant species [17] because nutritional composition in closely related plant species might be similar to the first species [2]. Only 3.17 per cent brood cells contained more than two species of pollen in mixed condition, among these one was present in a trace amount and it might have come along with nectar source. Because, these bees rarely specialize on nectar collection and they are expected to collect nectar from many plants species [17]. In this study, we observed LCBs nests containing trace amounts of pollen from unrelated species like, *Justicia*, *Ocimum*, *Ipomoea*, *Abelmoschus* pollen species as the bees may have visited these species for collecting nectar.

Pollen collected by LCBs gives valuable information on bee floral association. Among the floral family encountered in the LCBs nest Fabaceae was the dominant one accounting for 67 per cent followed by Asteraceae (11%) and each of the remaining families accounted for 6 per cent each (Fig. 2). This was probably because, many LCB species are pollen specialists (oligolectic) and restrict their pollen foraging to few taxonomically related plant species [17, 18]. Which is supported by an earlier study [19] found that *M. lanata* was an oligolectic solitary bee mainly associated with some leguminous crops.

Among the Fabaceae family, *Milletia pinnata* was seasonal in flowering from end of the January to June and peak flowering in *Peltophorum* was observed in April but it continued to flower staggardly till the end of year and crops like *Crotalaria juncea*, *Cajanus cajan* and *Dolichos lablab* were available year around in patches accounting for 86.50 per cent of total flora frequently encountered by bees. The remaining species were found to be meager/low (Fig. 3). The frequency of remaining six species likes *Helianthus annuus*, *Guizotia abyssinica* constituted 11%, while species of *Ocimum*, *Abelmoschus*, *Ipomoea* and *Justicia* were present in trace amounts (<1%).

This indeed is expected because LCBs being solitary and having short life cycle, have a foraging distance of 100-200m with high patch fidelity [10]. Because, host shifting or new host finding is very expensive for an LCB and it has to complete its nest construction within a stipulated period of time. Otherwise, the nest might become prone to attack of biotic factors like predators and parasites.

Further, mass provisioning nature demands maintenance of nutritional status that leads to foraging LCBs on closely related species [2]. Many studies have showed that there is no interspecific competition with respect to foraging range and overlap in resource use of flower-visiting ground and trap-nesting wild bees with honey bees [20].

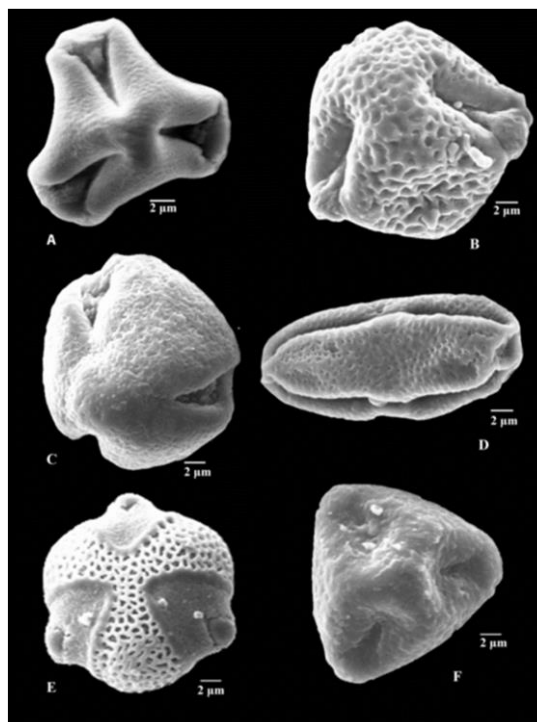


Plate 1a: (A) *Medicago sativa*, (B) *Cajanus cajana*, (C) *Cassia auriculata*, (D) *Crotalaria juncea*, (E) *Caesalpinia decapetala*, (F) *Cicer arietinum*

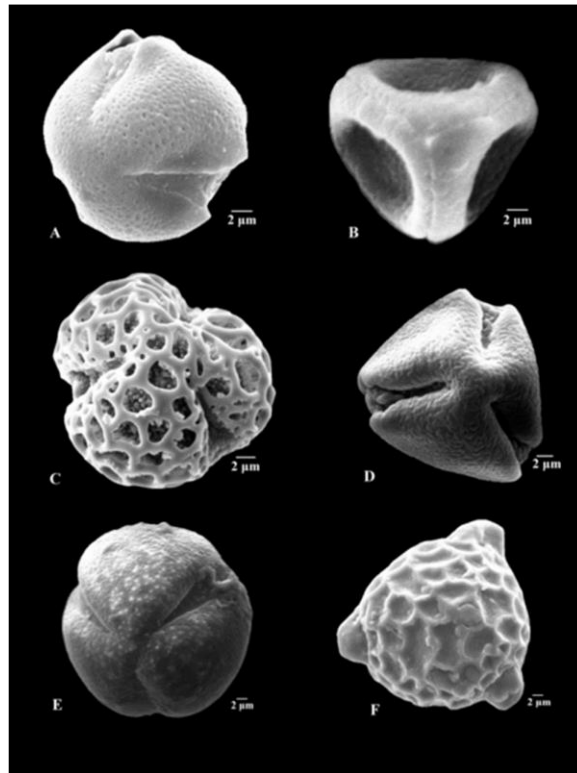


Plate 1b: (A) *Dolichos lablab*, (B) *Mimosa pudica*, (C) *Peltophorum pterocarpum*, (D) *Millettia pinnata*, (E) *Senna siamea*, (F) *Vigna sinensis*

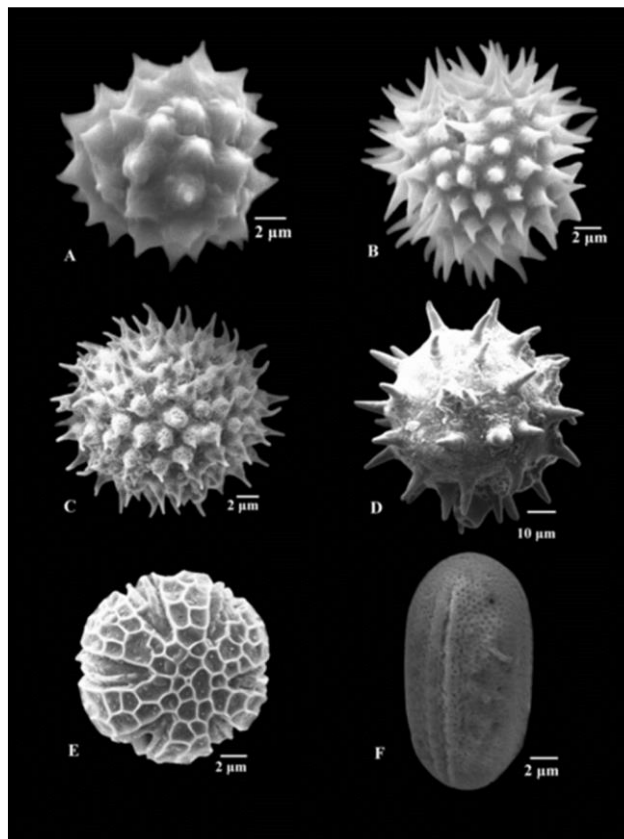


Plate: 1c: (A) *Helianthus anus*, (B) *Guizotia abyssinica*, (C) *Ipomoea sp.*, (D) *Abelmoschus sp.*, (E) *Ocimum sp.*, (F) *Justicia sp.*

Table 1: Flora collected by bee species

Sl. No.	Flora	Family
1	<i>Millettia pinnata</i>	Fabaceae
2	<i>Cajanus cajan</i>	Fabaceae
3	<i>Crotalaria juncea</i>	Fabaceae
4	<i>Peltophorum pterocarpum</i>	Fabaceae
5	<i>Dolichos lablab</i>	Fabaceae

6	<i>Vigna sinensis</i>	Fabaceae
7	<i>Mimosa pudica</i>	Fabaceae
8	<i>Senna siamea</i>	Fabaceae
9	<i>Cicer arietinum</i>	Fabaceae
10	<i>Medicago sativa</i>	Fabaceae
11	<i>Casia auriculata</i>	Fabaceae
12	<i>Caesalpinia decapetala</i>	Fabaceae
13	<i>Justicia sp</i>	Acanthaceae
14	<i>Helianthus annuus</i>	Asteraceae
15	<i>Guizotia abyssinica</i>	Asteraceae
16	<i>Ocimum sp</i>	Lamiaceae
17	<i>Ipomoea sp</i>	Convolvulaceae
18	<i>Abelmoschus esculentus</i>	Malvaceae

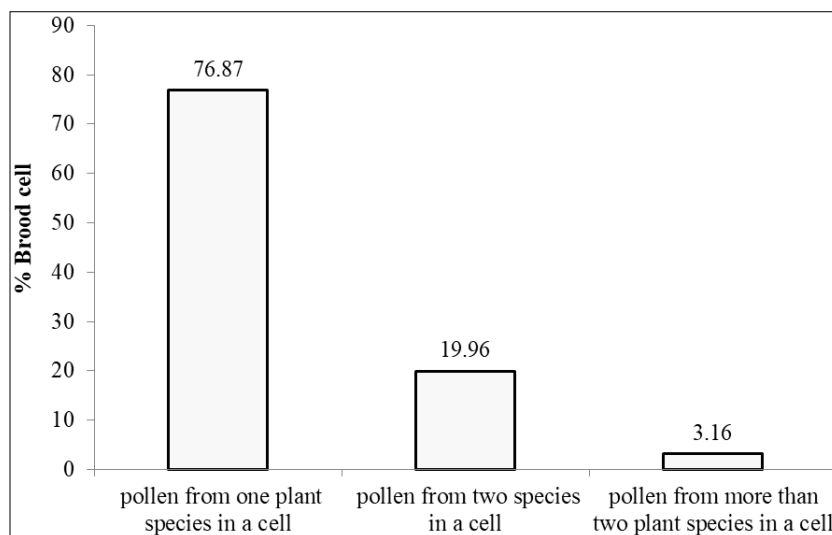


Fig 1: Pollen species in a brood cell of LCBs

Note: - Pollen from one plant species- At the time of sampling all the cells of each nest contained with pollen from any one plant species.
 Pollen from two plant species- At the time of sampling all the cells of each nest contained with pollen from any two of the plant species and had a cleared demarcated deposition.
 Pollen from more than two plant species- At the time of sampling all the cells of each nest contained with pollen from more than two plant species.

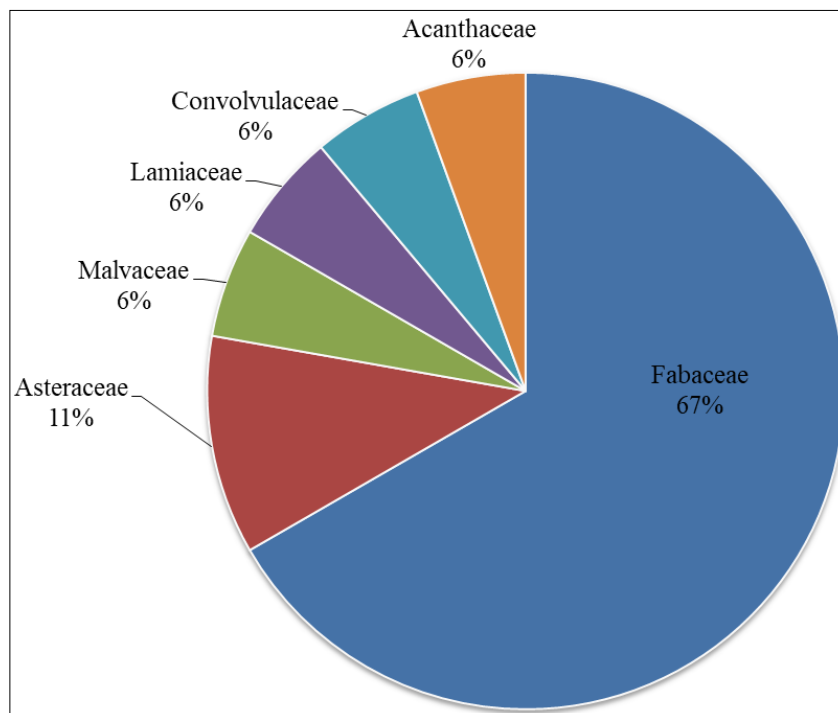


Fig 2: Frequency of floral family found in LCB nests

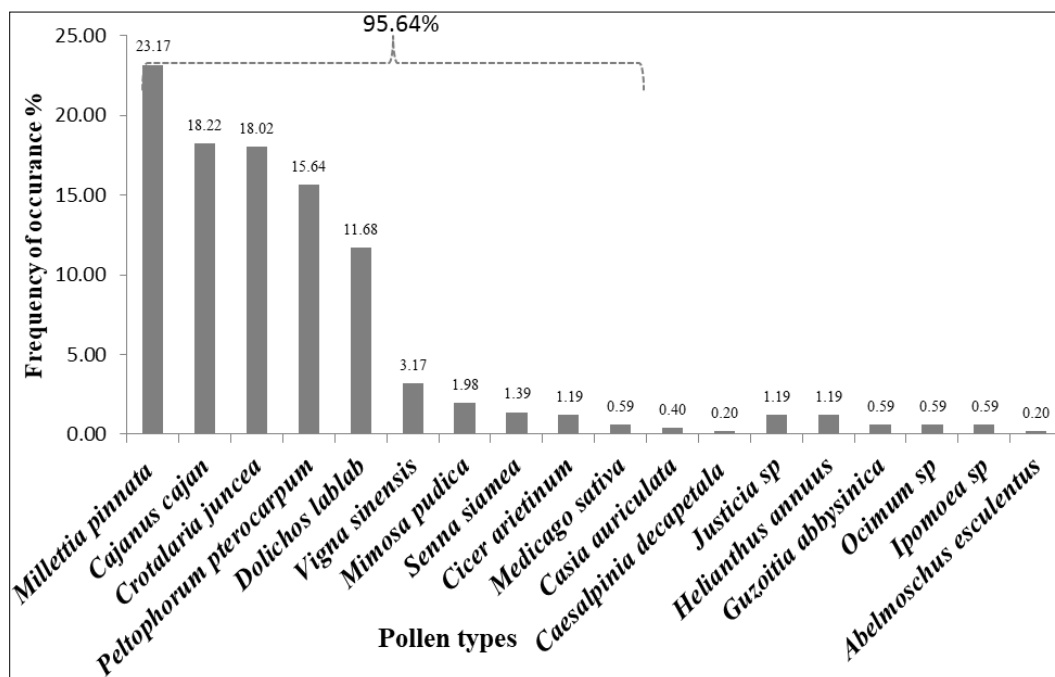


Fig 3: Frequency of floral species found in LCB nests

Appendix 1: Flora of GKVK Bengaluru.

Sl. No.	Common name	Scientific name	Family	Flowering period	Life form
1	Coconut	<i>Cocos nucifera</i>	Arecaceae	Year round	T
2	Royal palm	<i>Roystonea regia</i>	Arecaceae	Year round	T
3	Eucalyptus	<i>Eucalyptus obliqua</i>	Myrtaceae	Aug-Jan	T
4	Bottle brush	<i>Calistamen viminalis</i>	Myrtaceae	Year round	T
5	Jamun	<i>Syzygium cumini</i>	Myrtaceae	Feb-Apr	T
6	Rose apple	<i>Syzygium jambos</i>	Myrtaceae	May-July	T
7	Teek	<i>Tectona grandis</i>	Lamiaceae	Jun-Oct	T
8	Soap nut	<i>Sapindus emarginatus</i>	Sapindaceae	Oct-Nov	T
9	Pichkari/Nandi flame	<i>Spathodea companulata</i>	Bignoniaceae	Year round	T
10	Yellow trumpetbush	<i>Tecoma capensis</i>	Bignoniaceae	Year round	T
11	Sky jasmine	<i>Millingtona hortensis</i>	Bignoniaceae	Sept-Jan	T
12	Trumpet tree	<i>Tabebuia rosea</i>	Bignoniaceae	Nov-Jan	T
13	Trumpet tree	<i>Tabebuia argentea</i>	Bignoniaceae	Feb-May	T
14	Nile tulip	<i>Markhamia platycalyx</i>	Bignoniaceae	Aug-Sept	T
15	Indian-almond	<i>Terminalia catapa</i>	Combretaceae	Aug-Oct	T
16	Arjun tree	<i>Terminalia arjuna</i>	Combretaceae	Mar-May	T
17	Amla	<i>Phyllanthus emblica</i>	Phyllanthaceae	Feb-Mar	T
18	Breadfruit	<i>Artocarpus sp</i>	Moraceae	Dec-Jan	T
19	Copper pod	<i>Peltophorum pterocarpum</i>	Fabaceae	Mar-Jan	T
20	May flower	<i>Delonix regia</i>	Fabaceae	Apr-Jun	T
21	Cassia tree	<i>Senna siamea</i>	Fabaceae	July-Nov	T
22	Tamarind	<i>Tamarindus indica</i>	Fabaceae	Mar-Jun	T
23	Camel's foot tree	<i>Bauhinia variegata</i>	Fabaceae	Oct-Dec	T
24	Camel's foot tree	<i>Bauhinia perpurea</i>	Fabaceae	Feb-Sept	T
25	Baage mara	<i>Albizia lebeck</i>	Fabaceae	Mar-Nov	T
26	Bilvaara	<i>Albizia odoratissima</i>	Fabaceae	Mar-Nov	T
27	Earleaf acacia	<i>Acacia auriculiformis</i>	Fabaceae	Aug-Sept	T
28	Bili jaali	<i>Acacia leucophloea</i>	Fabaceae	Jul-Nov	T
29	Kaadu seege	<i>Acacia pennata</i>	Fabaceae	Jul-Oct	T
30	Mugali	<i>Acacia polyacantha</i>	Fabaceae	Sept-Dec	T
31	Avarice tree	<i>Cassia auriculata</i>	Fabaceae	Year round	T
32	Honge	<i>Millettia pinnata</i>	Fabaceae	Feb-Apr	T
33	Rosewood	<i>Dalbergia sp</i>	Fabaceae	Feb-Mar	T
34	Siver oak	<i>Grevillea robusta</i>	Proteaceae	Mar-Apr	T
35	Neem	<i>Azadirachta indica</i>	Meliaceae	Jan-Mar	T
36	Singapore cherry	<i>Muntingia calabura</i>	Muntingiaceae	Year round	T
37	Drumstick	<i>Moringa oleifera</i>	Moringaceae	Year round	T
38	Sandal wood	<i>Santalum album</i>	Santalaceae	Dec-June	T
39	Cinnamon tree	<i>Cinnamomum sulphuratum</i>	Lauraceae	Nov-Feb	T
40	Copper-wood	<i>Bursera simaruba</i>	Burseraceae	Dec-Jan	T

41	Star-fruit	<i>Averrhoa carambola</i>	Oxalidaceae	Year round	T
42	Phalsa	<i>Grewia asiatica</i>	Malvaceae	Feb-Mar	T
43	Mango	<i>Mangifera indica</i>	Anacardaceae	Nov-Jan	T
44	Hebbevu	<i>Melia dubia</i>	Meliaceae	Mar-Feb	T
45	Yenna Mara	<i>kingiodendron pinnatum</i>	Fabaceae	Apr-Aug	T
46	Indian rose chestnut	<i>Mesua ferrea</i>	Calophyllaceae	Mar-July	T
47	Cashew	<i>Anacardium occidentale</i>	Anacardaceae	Dec-Jan	T
48	Rain tree	<i>Samania saman</i>	Fabaceae	Mar-June	T
49	White teak	<i>Gmelina arborea</i>	Verbenaceae	Feb-Apr	T
50	Blue jacaranda	<i>Jacaranda mimosifolia</i>	Bignoniaceae	Apr-Oct	T
51	Pride of India	<i>Lagerstroemia speciosa</i>	Lythraceae	Mar-Apr	T
52	Red sandle	<i>Pterocarpus santalinus</i>	Fabaceae	Mar-Apr	T
53	Badminton Ball Tree	<i>Parkia biglandulosa</i>	Fabaceae	Dec-Jan	T
54	Bendi	<i>Kydia calycina</i>	Malvaceae	Sept-Nov	T
55	Kadamb tree	<i>Anthocephalus cadamba</i>	Rubiaceae	Sept-Oct	T
56	Baeladamara	<i>Limonia acidissima</i>	Rutaceae	Mar-May	T
57	Arasina buruga	<i>Cochlospermum religiosum</i>	Bixaceae	Feb-Apr	T
58	Andimurugalu	<i>Carallia brachiata</i>	Rhizophoraceae	Dec-Jun	T
59	Garudaphala	<i>Hydnocarpus wightianus</i>	Achariaceae	Jan-Apr	T
60	Indian Elm	<i>Holoptelea integrifolia</i>	Ulmaceae	Feb-Mar	T
61	Bombay ebony	<i>Diospyros montana</i>	Ebenaceae	March-April	T
62	True indigo	<i>Indigofera sp</i>	Fabaceae	May-Aug	SH
63	Indian Redwing	<i>Pterilobium sp</i>	Fabaceae	Sept-Oct, Mar-Apr	SH
64	Sun hemp	<i>Crotalaria juncea</i>	Fabaceae	Year round	SH
65	Pigeon pea	<i>Cajanus cajana</i>	Fabaceae	July-Mar	SH
66	Powder puff plant	<i>Calliandra haematocephala</i>	Fabaceae	Year round	SH
67	Feather-leaved Cassia	<i>Chamaecrista mimosoides</i>	Fabaceae	Nov-Mar	H
68	Field bean	<i>Dolichus lablab</i>	Fabaceae	Sept-Mar	H
69	Cow pea	<i>Vigna sinensis</i>	Fabaceae	June-Aug, Feb-Mar	H
70	Touch-me-not	<i>Mimosa pudica</i>	Fabaceae	Year round	H
71	Alfa alfa	<i>Medicago sativa</i>	Fabaceae	Mar-Apr	H
72	Chick pea	<i>Cicer arietinum</i>	Fabaceae	Oct-Dec	H
73	Mysore thorn	<i>Caesalpinia decapetala</i>	Fabaceae	Year round	SH
74	Peacock flower	<i>Caesalpinia pulcherima</i>	Fabaceae	Year round	SH
75	Divi Divi	<i>Caesalpinia coriaria</i>	Fabaceae	Jul-Aug	SH
76	Subabul	<i>Leucaena leucocephala</i>	Fabaceae	Year round	SH
77	Natkadeodar	<i>Erythroxylum monogynum</i>	Erythroxylaceae	Aug-Sep	SH
78	Chilly	<i>Capsicum annum</i>	Solanaceae	Year round	H
79	Brazilian Nightshade	<i>Solanum seafortianum</i>	Solanaceae	Year round	C
80	Butterfly Pea	<i>Clitoria ternatea</i>	Fabaceae	Apr-Jun	C
81	American black nightshade	<i>Solanum americanum</i>	Solanaceae	Year round	H
82	Egg plant	<i>Solanum melongena</i>	Solanaceae	Year round	SH
83	Bavanchi	<i>Psoralea corylifolia</i>	Fabaceae	Jul-Aug	H
84	Purple bush-bean	<i>Macroptilium atropurpureum</i>	Fabaceae	Year round	C
85	Calico-plant	<i>Alternanthera betzickiana</i>	Amaranthaceae	Year round	H
86	Cock's comb	<i>Celosia argentea</i>	Amaranthaceae	Year round	H
87	Amaranth	<i>Amaranthus sp. 1</i>	Amaranthaceae	Year round	H
88	Amaranth	<i>Amaranthus sp. 2</i>	Amaranthaceae	Year round	H
89	Indian plum	<i>Flacourtia sp</i>	Salicaceae	Nov-Mar	SH
90	Loranthus	<i>Dendrothoe falcata</i>	Loranthaceae	Year round	E
91	Mexican clover	<i>Richardia scabra</i>	Rubiaceae	Year round	H
92	Fameflower	<i>Talinum portulacifolium</i>	Portulacaceae	Year round	H
93	Moss roses	<i>Portulaca sp.</i>	Portulacaceae	Year round	H
94	Wireweed	<i>Sida sp</i>	Malvaceae	Year round	H
95	Bagflower	<i>Clerodendrum sp</i>	Lamiaceae	Year round	SH
96	Tulsi	<i>Ocimum sp</i>	Lamiaceae	Year round	H
97	Spurge	<i>Euphorbia hypericifolia</i>	Euphorbiaceae	Year round	C
98	Wild Poinsettia	<i>Euphorbia heterophylla</i>	Euphorbiaceae	Year round	H
99	Croton	<i>Croton sp.</i>	Euphorbiaceae	Year round	H
100	Castor	<i>Ricinus cummunis</i>	Euphorbiaceae	Dec-July	SH
101	Croton	<i>Codiaeum variegatum</i>	Euphorbiaceae	Year round	SH
102	Yellow bells	<i>Tecoma stans</i>	Bignoniaceae	Year round	SH
103	Orange trumpetvine	<i>Pyrostegia venusta</i>	Bignoniaceae	Jan-Feb	H
104	Goat weed	<i>Ageratum conyzoides</i>	Asteraceae	Year round	H
105	Blazing star	<i>Maulua spicata</i>	Asteraceae	Year round	H
106	Cosmos	<i>Cosmos sp</i>	Asteraceae	Year round	H
107	False Mallow	<i>Malvestrum coromandelianum</i>	Malvaceae	Year round	H
108	Bhendi	<i>Abelmoschus esculentus</i>	Malvaceae	Year round	H

109	White Buttercup	<i>Turnera subulata</i>	Passifloraceae	Year round	H
110	Yellow Buttercups	<i>Turnera ulmifolia</i>	Passifloraceae	Year round	H
111	Passion Flowers	<i>Passiflora sp.</i>	Passifloraceae	Year round	C
112	Morning Glories	<i>Ipomea sp.</i>	Convolvulaceae	Year round	H
113	Creeping-Oxeyes	<i>wedelia sp.</i>	Asteraceae	Year round	H
113	Coat buttons	<i>Tridax procumbens</i>	Asteraceae	Year round	H
114	Knapweeds	<i>Centaurea sp</i>	Asteraceae	Year round	H
115	Black-jack	<i>Bidens pilosa</i>	Asteraceae	Year round	H
116	Quick Weed	<i>Galinsoga parviflora</i>	Asteraceae	Year round	H
117	Node weed	<i>Synedrella vialis</i>	Asteraceae	Year round	H
118	Node Weed, Pig Grass	<i>Synedrella nodiflora</i>	Asteraceae	Year round	H
119	Tassel flower	<i>Emilia sanchifolia</i>	Asteraceae	Year round	H
120	Niger	<i>Guizotia abyssinia</i>	Asteraceae	July-Nov, Jan-Mar	H
121	Sunflower	<i>Helianthus annus</i>	Asteraceae	Year round	SH
122	Congress weed	<i>Parthanium hysterophorus</i>	Asteraceae	Year round	H
123	Dandotapala	<i>Vernonia sp</i>	Asteraceae	Year round	H
124	Blanketflower	<i>Gaillardia sp</i>	Asteraceae	Year round	H
125	Spider flowers	<i>Cleome monophylla</i>	Cleomaceae	Year round	H
126	Mustard	<i>Brasica sp</i>	Brassicaceae	Jun-July, Dec-Jan	H
127	Nut grass	<i>Cyprus sp.</i>	Poacea	Year round	S
128	Maize	<i>Zea maize</i>	Poacea	Year round	SH
129	Buffalo grass	<i>Bouteloua sp</i>	Poacea	Year round	H
130	Horsewood	<i>Clausena sp</i>	Rutaceae	Jun-July	SH
131	Coriander	<i>Coriandrum sp</i>	Apiaceae	Year round	H
132	False heather	<i>Cuphea sp</i>	Lythraceae	Year round	H
133	Chuparosa	<i>Justicia sp</i>	Acanthaceae	Year round	H
134	Chinese violet	<i>Asystasis sp</i>	Acanthaceae	Year round	H
135	White-lady	<i>Thunbergia fragrans</i>	Acanthaceae	Year round	C
136	Coral vine	<i>Antigonum leptopus</i>	Polygonaceae	Year round	C
137	Narrow-Leaved Milkwort	<i>Polygala elongata</i>	Polygalaceae	Year round	H
138	Puncture Vine	<i>Tribulus terrestris</i>	Zygophyllaceae	Year round	H
139	Bana Chakunda	<i>Senna occidentalis</i>	Fabaceae	Year round	SH
140	Pomegranate	<i>Punica granatum</i>	Punicaceae	Apr-July	SH
141	Rattlebox Plant	<i>Crotalaria pallida</i>	Fabaceae	June-Nov	H
142	One leaf Senna	<i>Cassia uniflora</i>	Fabaceae	Year round	H
143	Pencil-Flower	<i>Stylosanthes fruticosa</i>	Fabaceae	Year round	H
144	Carpetweed	<i>Mollugo pentaphylla</i>	Molluginaceae	Year round	H
145	Snakeweed	<i>Stachytarpheta indica</i>	Verbenaceae	Year round	H
146	Girdlepod	<i>Mitracarpus hirtus</i>	Rubiaceae	Year round	H
147	Silkleaf	<i>Lagasea mollis</i>	Asteraceae	Year round	H
148	Golden dewdrop	<i>Duranta repens</i>	Verbenaceae	Year round	SH
149	Divi Divi	<i>Caesalpinia coriaria</i>	Fabaceae	Year round	SH
150	Burnet-saxifrage	<i>Pimpinella sp</i>	Apiaceae	Year round	H
151	Rosemary	<i>Rosmarinus officinalis</i>	Lamiaceae	Year round	H
152	Physic nut	<i>Jatropha curcas</i>	Euphorbiaceae	Year round	SH
153	Globe amaranth	<i>Gomphrena sp</i>	Amaranthaceae	Year round	H
154	Gliricidia	<i>Gliricidia sepium</i>	Fabaceae	Feb-Mar	SH
155	Ridge guard	<i>Luffa acutangula.</i>	Cucurbitaceae	Sept-Nov	C
156	Bitter guard	<i>Momordica charantia</i>	Cucurbitaceae	Sept-Nov	C
157	Dwarf Morning Glory	<i>Evolvulus alsinoides</i>	Convolvulaceae	Year round	C
158	Roundleaf Bindweed	<i>Evolvulus nummularius</i>	Convolvulaceae	Year round	C
159	Skyblue Clustervine	<i>Jacquemontia pentanthos</i>	Convolvulaceae	Year round	C
160	Shatavari	<i>Asparagus racemosus</i>	Asparagaceae	Year round	H
161	Sweet Clock-Vine	<i>Thunbergia fragrans</i>	Acanthaceae	Year round	H
162	Nelaberu	<i>Andrographis paniculata</i>	Acanthaceae	Sept-Oct	H
163	Indian mallow	<i>Abutilon hirtum</i>	Malvaceae	Year round	H
164	Thumbai	<i>Lucas sp</i>	Lamiaceae	Year round	H
165	Dayflowers	<i>Commelina sp</i>	Commelinaceae	Year round	H
166	Pignut	<i>Hyptis sp</i>	Lamiaceae	Year round	H
168	Coffee	<i>Coffea arabica</i>	Rubiaceae	Jan-Mar	SH

Note: T-Tree, H- Herb, SH- Shrub, C- Creeper/Climber

4. Conclusion

Our study has pointed out that megachilids exhibit floral preference to the members of the family Fabaceae. Flowers of these plants have keel petals, a structure that might have evolved to protect against pollen theft by generalists bees. And probably in some plant species flowers and pollen were

structurally or chemically protected and accessed only by a specialized pollinator. Hence generalist bees do not or avoid foraging on the flora preferred by specialist bees. We postulate that the enormous pollen requirements of bees might have led to selection of flowers and pollen with such protective properties. These conclusions open a new field of

research in the study of insect–flower relationships.

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