



E-ISSN: 2320-7078

P-ISSN: 2349-6800

JEZS 2018; 6(3): 1287-1292

© 2018 JEZS

Received: 26-03-2018

Accepted: 27-04-2018

P Konwar

Department of Animal Nutrition, College of Veterinary Science and Animal Husbandry Anjora, Chhattisgarh Kamdhenu Vishwavidyalaya, Durg, Chhattisgarh, India

SP Tiwari

Professor and Head, Department of Animal Nutrition, College of Veterinary Science and Animal Husbandry Anjora, Chhattisgarh Kamdhenu Vishwavidyalaya, Durg, Chhattisgarh, India

M Gohain

Department of Animal Genetics and Breeding, College of Veterinary Science, Assam Agricultural University, Guwahati, India

M Buragohain

Department of Pathology, College of Veterinary Science, Assam Agricultural University Guwahati, India

Effect of protein supplementation on growth, FCE, nutrient utilization and rumen fermentation pattern in goat with natural subclinical nematodiasis

P Konwar, SP Tiwari, M Gohain and M Buragohain

Abstract

The study was conducted on twenty indigenous non-descript goat kids allocated into four different groups. T₁ was the negative control without concentrate supplement. T₂, T₃ and T₄ were supplemented with different levels of concentrate mixtures having 16, 20 and 24% DCP respectively. The animals were naturally infected with gastrointestinal nematodes and the experiment lasted for 60 days. The DM, DCP and TDN intake were found significant ($P < 0.01$) and being highest in T₃ group. The body weight gain and feed conversion efficiency, N and P balance were significantly ($P < 0.01$) highest in T₃ and lowest in T₁ whereas Ca balance was highest in T₃ and lowest in T₄. At the 45th day of experiment, ruminal pH significantly ($P < 0.05$) increased in protein supplemented groups. The total nitrogen, TCA preceptible nitrogen, soluble nitrogen, microbial protein and total bacterial count were significantly ($P < 0.01$) higher in T₃ while the total protozoal counts were significantly ($P < 0.01$) lower in T₃.

Keywords: Goat, subclinical nematodiasis, protein supplement, nutrient utilization

1. Introduction

Gastrointestinal parasitism is a major problem in small ruminant production worldwide, due to its impact on animal health and productivity and the associated costs of control measures [1]. The nutritional status of the host has a significant effect on its ability to withstand parasite infection, and it has also been frequently observed that the mortality resulting from gastrointestinal nematodes is more severe in malnourished animals than in well-nourished animals [2]. The usual mode of control of gastrointestinal nematodes based on the repeated use of anthelmintics is strongly questioned because of the increasing development of resistance to these molecules [3]. The manipulation of host nutrition is the alternative method to anthelmintics to improve the host resistance and/or resilience to parasitic infections. The presence of worms usually induces decreased appetite, decreased digestibility of food and a diversion of nutrients from production sites towards the repair of tissue-damage causing assimilation of nematode parasitism to a nutritional disease [4]. The patho-physiological studies reveal that protein metabolism is much more disturbed by the presence of gastrointestinal nematodes than other components of the diet, including energy [5]. Recent studies have underlined the need to consider both the protein and energy components, particularly in tropical conditions. Changes in protein metabolism as a major consequence of infection with gastrointestinal nematodes have focused attention on the effects of modifications in protein supply on outcome of infection [6]. There is a need to define further nutrition-parasite interactions in infected goats.

2. Materials and Methods

Twenty non-descript goat kids of 3-5 months were selected for the study were reared in semi-intensive method. They were randomly allocated into 4 groups (T₁, T₂, T₃ and T₄) of 5 animals each on the body weight basis. Group T₁ was left as negative control without any concentrate supplement. Three concentrate diets were prepared; T₂ {Normal Protein Normal Energy (NPNE)}, T₃ {Medium Protein Normal Energy (MPNE)} and T₄ {High Protein Normal Energy (HPNE)}. NPNE diet contains 16% DCP and 70% TDN, MPNE diet contains 20% DCP and 70% TDN and HPNE diet contains 24% DCP and 70% TDN.

Correspondence**P Konwar**

Department of Animal Nutrition, College of Veterinary Science and Animal Husbandry Anjora, Chhattisgarh Kamdhenu Vishwavidyalaya, Durg, Chhattisgarh, India

All kids were herd grazed for four hours in field with mixed grass predominant with sola grass (*Aeschynomene indica*) in the morning time (8:00 am to 12:00 noon), followed by supplementary feeding of 100 g concentrate mixture. All kids were weighed fortnightly for three consecutive days during the entire experiment. Feed conversion in kids was calculated as the ratio of feed consumed over body weight gain. Daily gain in body weight of kids was determined by subtracting the initial body weight from the final body weight. Daily feed intake of all kids was recorded for 60 days. It was determined by subtracting leftover DM from the amount of dried feed offered.

2.1 Sample collections

The fresh faecal samples were collected per rectum and faecal examination was done by modified McMaster Technique [7]. Dried faecal samples were used for analysis of calcium and phosphorus. Nitrogen estimation was done by mixing the sample with 25% sulphuric acid. Urine samples were collected using toluene as a preservative for balance study of nitrogen, calcium and phosphorus. Rumen liquor was collected through the stomach tubes at 45th day of the experiment, strained through 4 coats of muslin cloths to analyze the rumen metabolites.

2.2 Nutritional parameters

2.2.1 Proximate analysis of feed and faeces

Samples of feed offered, residue left and faeces voided were analyzed for various proximate principles viz., dry matter, organic matter, nitrogen, ether extract, crude fibre, total ash and acid insoluble ash [8]. The samples were also analyzed for calcium [9] and inorganic phosphorus [10].

2.2.2 Van Soest analysis of fibre

The sola grass was analyzed for neutral detergent fibre (NDF), acid detergent fiber (ADF), cellulose, hemicellulose and lignin [11].

3. Ethical consideration

The ethical approval was granted by the Institutional Animal Ethics Committee.

4. Statistical analysis

All statistical analyses were performed with standard software (IBM SPSS Statistics 20 software).

5. Results and Discussions

5.1 Effect on Dry Matter Intake (Growth trial)

The average daily DM intake (g/day) of the entire feeding trial at 15 d interval in various treatment groups has been presented in table 1. Significant ($P<0.01$) difference were obtained between different treatment groups. In this experiment, the highest dry matter intake was observed in the T₃ and lowest in the T₄.

The lower DM intake in T₄ might be due to higher density of nutrient. Previous workers established significant relationship between the severity of infection and the nutrient intake by the animals [12]. The present study revealed depressed feed intake as the level of infection increased in different treatments which might be due to the panic reaction by the kids. The blood sucking gastrointestinal parasites sucks blood from deeper abomasal and intestinal mucosa causing profuse loss of N in the form of desquamated epithelial cells of damaged tissues, which consequently caused aggravated pain resulting into reduced overall nutrient intake [13]. The findings were in accordance to the earlier report [14]. Voluntary feed intake/dry matter intake of *H. contortus* infected animals increased with increasing dietary crude protein content than those fed lower protein diets [15]. In earlier report, inappetence was observed in *H. contortus* infected lambs receiving low protein diet with lesser inappetence in lighter infections [16]. The protein/amino acids requirement increases due to increase in protein synthesis to overcome the loss of endogenous amino acids through the process of deamination and/or fermentation in the small and large intestine.

Table 1: Effect of protein supplementation on dry matter intake (g/day) of kid

Period (day)	Treatment				Overall Mean
	T ₁	T ₂	T ₃	T ₄	
0-15	451.32±0.52 ^b	454.16±0.15 ^c	458.17±0.15 ^d	400.01±0.18 ^a	440.92±5.45**
16-30	456.67±0.65 ^b	458.98±0.12 ^c	463.34±0.31 ^d	405.08±0.18 ^a	446.02±5.45**
31-45	463.36±1.84 ^b	463.97±0.25 ^b	468.85±0.49 ^c	410.12±0.28 ^a	451.58±5.53**
46-60	472.98±1.68 ^b	473.81±0.24 ^b	478.55±0.37 ^c	420.17±0.18 ^a	461.37±5.49**

(** $P<0.01$)

5.2 Nutrient Utilization (Growth trial)

5.2.1 Digestible Crude Protein (DCP) Intake and Total Digestible Nutrient (TDN) Intake

The mean DCP intake (g/d) and TDN intake (g/d) values at the end of experiment was highly significant ($P<0.01$). The highest DCP intake (g/d) and TDN intake (g/d) were recorded in T₃ (Table 2).

The voluntary feed intake of *H. contortus* infected lambs increased with increasing dietary crude protein than those fed lower protein diets [17]. As the crude protein content of the diet was higher, there was higher feed intake that caused higher DCP intake but T₄ had lesser intake which might be due to impaired energy protein ratio which is very much important in digestion of nutrients.

Table 2: Effect of protein supplementation on DCP and TDN intake in kid

Groups	DCP intake (g/d)	TDN intake (g/d)
T ₁	40.40±0.25 ^a	247.25±0.28 ^b
T ₂	47.32±0.18 ^b	265.18±0.23 ^c
T ₃	52.39±0.21 ^d	268.38±0.23 ^d
T ₄	50.99±0.15 ^c	235.25±0.18 ^a

5.3 Growth Performance

The mean fortnightly body weight (kg), fortnightly body weight gain (g) and body weight gain (g/day) were presented in table 3, 4 and 5. The fortnightly body weight of kids, fortnightly body weight gain and daily body weight gain were significant ($P<0.05$, $P<0.01$) among the treatments during the entire experiments where the T₃ group revealed highest body weight and T₁ was found to be lowest.

The result of the present study suggested that protein supplementation to kids reduced the adverse effect of *H. contortus* infection by improving body weight, body weight gain and feed conversion efficiency and also increase the resistance and/or resilience. T₃ group revealed better performance as compared to other groups which might be due to the fact that high proteinous diet produced the fermentative product, the NH₃-N, more rapidly as compared to the low protein diet in the alimentary tract, which caused inhibition of moulting of pathogenic larvae (L₃) to adult one, leading to

reduction of total eggs in the faeces [13]. In the present study, T₄ group containing higher level of protein showed lower performance than T₃, which might be due to much more protein level leading to lowered intake by kids to maintain energy equilibrium. Increased dietary protein leads to increase amino acids in gut leading to increase deposition as muscles resulting in live wt gain in goats [18]. Higher growth performance with increased protein supplementation was also reported by earlier workers [19].

Table 3: Effects of protein supplementation on fortnightly bodyweight (Kg) of kids

Period (day)	Treatment				Overall Mean	Significance
	T ₁	T ₂	T ₃	T ₄		
0	8.91±0.12	8.90±0.12	8.89±0.15	8.90±0.09	8.90±0.06	NS
15	9.12±0.12 ^a	9.30±0.12 ^{ab}	9.59±0.15 ^b	9.44±0.10 ^{ab}	9.36±0.07	*
30	9.36±0.12 ^a	9.73±1.11 ^{ab}	10.31±0.15 ^c	10.00±0.12 ^{bc}	9.85±0.10	**
45	9.62±0.13 ^a	10.19±0.11 ^b	11.07±0.15 ^d	10.60±0.13 ^c	10.37±0.14	**
60	9.89±0.13 ^a	10.70±0.11 ^b	11.85±0.16 ^d	11.23±0.15 ^c	10.92±0.18	**

(NS- Non significant, * $P < 0.05$, ** $P < 0.01$)

Table 4: Effects of protein supplementation on fortnightly bodyweight gain (Kg) of kids

Period (day)	Treatment				Overall Mean	Significance
	T ₁	T ₂	T ₃	T ₄		
0-15	208±8.00 ^a	400±13.04 ^b	696±12.88 ^d	536±13.64 ^c	460±41.50	**
16-30	244±11.66 ^a	426±21.12 ^b	724±4.00 ^d	564±15.03 ^c	489.5±41.06	**
31-45	258±9.17 ^a	472±10.20 ^b	756±5.10 ^d	596±14.35 ^c	520.5±42.01	**
46-60	274±8.72 ^a	506±7.48 ^b	786±5.10 ^d	632±15.94 ^c	549.5±43.25	**

(** $P < 0.01$)

Table 5: Effect of protein supplementation on daily body weight gain (g/day) of kids

Period (day)	Treatment				Overall Mean	Significance
	T ₁	T ₂	T ₃	T ₄		
0-15	13.86±0.53 ^a	26.67±0.87 ^b	46.40±0.86 ^d	35.73±0.91 ^c	30.67±2.27	**
16-30	16.27±0.78 ^a	28.40±1.41 ^b	48.27±0.27 ^d	37.60±1.00 ^c	32.63±2.74	**
31-45	17.20±0.61 ^a	31.47±0.68 ^b	50.40±0.34 ^d	39.73±0.96 ^c	34.70±2.80	**
46-60	18.26±0.58 ^a	33.73±0.50 ^b	52.40±0.34 ^d	42.13±1.06 ^c	36.63±2.88	**

(** $P < 0.01$)

5.4 Feed Conversion Efficiency (FCE)

The influence of protein supplementation on feed conversion efficiency (FCE) was presented in table 6. There was significant ($P < 0.01$) difference amongst treatments on FCE during the entire growth trial having highest value in T₃ and lowest in T₁.

The results of the present study may be due to the same reason as the increase in growth performance of the kids at this stage. The reduced FCE in kids with lower protein supplementation showed the impact of parasites on nutrient utilization, the efficiency of digestion and absorption of nutrients. The parasite competes with host for the availability of proteinous material in intestine, thus decreasing the

availability of amino acids to the kids affecting the overall growth of animals [20]. A significant amount of protein is also redirected for the repair of the damaged mucosa caused by GI nematodes, synthesis of extra amount of specific serum protein and for production of immunoglobulin reducing the amount of protein deposition in the musculature required for collagen fiber synthesis [13]. This leads to the reduction of the body weight gain. The presence of *Haemonchus* larvae induces abomasal gland hyperplasia, the higher abomasal pH decreases the conversion of pepsinogen to pepsin which leads to reduced protein digestion, increases mucosal permeability and endogenous protein losses in the abomasum.

Table 6: Effect of protein supplementation on feed conversion efficiency of kids

Period (day)	Treatment				Overall Mean
	T ₁	T ₂	T ₃	T ₄	
0-15	0.0307±0.0012 ^a	0.0587±0.0019 ^b	0.1013±0.0019 ^d	0.0893±0.0023 ^c	0.0700±0.0064**
16-30	0.0356±0.0017 ^a	0.0619±0.0030 ^b	0.1041±0.0006 ^d	0.0928±0.0025 ^c	0.0736±0.0062**
31-45	0.0371±0.0015 ^a	0.0678±0.0014 ^b	0.1075±0.0008 ^d	0.0969±0.0024 ^c	0.0773±0.0063**
46-60	0.0386±0.0013 ^a	0.0712±0.0010 ^b	0.1095±0.0008 ^d	0.1002±0.0025 ^c	0.0799±0.0064**

5.5 Balance Study

5.5.1 Nitrogen Balance

The nitrogen intake, outgo, balance and percent retained (g/d)

were statistically significant ($P < 0.01$). The highest nitrogen intake and retention were recorded in T₃ and outgo in T₄ whereas the lowest were recorded in T₁ group. The data has

been presented in table 7.

As the level of nutrition increased the nitrogen intake, outgo and their balance were found to be increased. The kids with lesser infection retained more nitrogen as compared to highly infected kids. However in T₄ group, although with high protein level the balance was less which might be due to impaired digestion and absorption due to impaired energy-protein ratio. In the present study parasites caused extensive protein losses and redirect protein synthesis away from skeletal muscle and therefore help in the repair of gut tissues leading to reduced nitrogen balance [21]. However, several workers suggested that loss of endogenous nitrogen into the intestine was the major reason for reduced nitrogen gain [22]. Increasing dietary protein intake and abomasal infusion of protein resulted in the animal being much better, able to tolerate these infections and improved nitrogen retention [5]. Moreover, increased nitrogen outgo was reported on sheep infected with *H. contorts* [14].

Table 7: Effect of protein supplementation on N, Ca and P balance in kid

Groups	Nitrogen (g/d)				Calcium (g/d)				Phosphorus (g/d)			
	Intake	Out go		Balance	Intake	Out go		Balance	Intake	Out go		Balance
		Faecal	Urinary			Faecal	Urinary			Faecal	Urinary	
T ₁	9.42± 0.02 ^a	2.18± 0.07 ^a	0.93± 0.03 ^a	6.38± 0.02 ^a (67.19%)	6.77± 0.13 ^d	3.60± 0.02 ^d	0.28± 0.01 ^a	2.88± 0.02 ^c (42.56%)	0.76± 0.00 ^a	0.11± 0.00 ^d	0.04± 0.00 ^b	0.61± 0.00 ^a (79.36%)
T ₂	11.16± 0.04 ^b	2.25± 0.02 ^c	1.10± 0.03 ^b	7.81± 0.02 ^b (69.96%)	6.31± 0.01 ^b	3.21± 0.01 ^c	0.31± 0.01 ^b	2.79± 0.01 ^b (44.29%)	1.02± 0.01 ^c	0.10± 0.00 ^c	0.05± 0.00 ^d	0.88± 0.01 ^c (85.50%)
T ₃	12.28± 0.05 ^d	2.23± 0.01 ^b	1.28± 0.01 ^d	8.77± 0.05 ^d (71.41%)	6.40± 0.02 ^c	3.11± 0.01 ^b	0.32± 0.01 ^c	2.97± 0.01 ^d (46.41%)	1.11± 0.00 ^d	0.07± 0.00 ^a	0.03± 0.00 ^a	1.01± 0.00 ^d (90.44%)
T ₄	11.98± 0.05 ^c	2.35± 0.01 ^d	1.17± 0.01 ^c	8.45± 0.05 ^c (70.55%)	5.52± 0.01 ^a	2.71± 0.01 ^a	0.31± 0.01 ^b	2.50± 0.01 ^a (45.35%)	0.91± 0.00 ^b	0.08± 0.00 ^b	0.04± 0.00 ^c	0.79± 0.00 ^b (86.27%)

5.6 Rumen Fermentation Pattern

5.6.1 Ruminal pH

The results of the average pH of the experimental animals under different treatments have been presented in table 8. The average pH values of SRL drawn at 45th day of experiment were significant ($P<0.05$) amongst the treatments. The highest pH was recorded in T₃ where as the lowest in T₁ group.

In the present study, pH was slightly higher in T₃ where diet was supplemented with high protein than normal. This showed that inspite of maximum nitrogen intake in T₃ the pH was not much affected. This could not therefore adversely affect the buffering capacity of rumen required for normal growth of rumen microbes and fermentation pattern. Previous study revealed that protein supplementation also increases the pH of the rumen, which might have affected the exsheathment of the infective larvae [17]. The present study was supported by the findings of different workers [13].

5.6.2 Total Nitrogen, TCA Preceptible Nitrogen and Soluble Nitrogen

The results of the average total nitrogen, TCA preceptible nitrogen and soluble nitrogen of the experimental animals under different treatments have been presented in table 8.

The mean total nitrogen values of SRL drawn at 45th day of experiment were significant ($P<0.01$) among themselves where highest value shown in T₃ and lowest in T₁ group. It was suggested that total-N concentration of rumen contents increased in response to increase in nitrogen intake in sheep [26]. In our experiment similar trends were observed. Similar results were also observed by various workers in small

5.5.2 Calcium and Phosphorus Balance

The calcium and phosphorus intake, outgo, balance and percent retained by the kids during the metabolic trial were significant ($P<0.01$) (Table 7).

The results of the present study might be due to the higher content of calcium or proper utilization of feed. Lowest calcium balance was observed in T₄ and highest in T₃ which might be due to the effect of the gut nematodes. Higher Ca balance in sheep fed with soyabean meal as compared to non supplemented group was reported by previous workers [23].

The phosphorous balance was highest in T₃ and the lowest in T₁ group. Higher P balance might be due to the higher content of P in diet or better utilization of feed. As the infection increased the P balance was reduced which might be due to the reduced intake and impairment of P absorption [24]. The intestinal infections with nematodes reduced the absorption and retention of P and the mineralization of the skeleton [25].

ruminants [13].

The average TCA preceptible nitrogen value of SRL drawn at 45th day of experiment in T₃ were significantly ($P<0.01$) higher than other treatments groups where T₁ had the lowest value. Hume *et al.* (1970) Earlier studies said that the concentration of TCA preceptible N increased in the rumen of sheep with an increase in the nitrogen content of the diet. Similar trends were obtained by other workers [13].

The average soluble nitrogen values of SRL drawn at 45th day of experiment were significantly ($P<0.01$) differed among the treatments. The value was highest in T₃ and the lowest in T₂ group.

5.6.3 Microbial Protein

The results of the average microbial protein of the experimental animals under different treatments have been presented in table 8. The average microbial protein of SRL drawn at 45th day of experiment was significantly ($P<0.01$) higher in T₃ followed by T₄, T₂ and T₁ respectively.

Microbial protein synthesis was reflected by the availability of fermentable N and soluble carbohydrate in the rumen. Microbial protein was significantly higher in groups supplemented with urea molasses mineral granules (UMMG) in goat kids [27]. Results of the present study might be due to availability of fermentable-N from supplemented high protein diet to the microbial growth and multiplication leading to higher microbial protein. Similar findings were also reported in goats where increased N increased the microbial protein level [13].

5.6.4 Total Bacterial and Protozoal Count

The results of the average total bacterial count of the experimental animals under different treatments have been presented in table 8. The average total bacterial count of SRL drawn at 45th day of experiment were significantly ($P<0.01$) higher in T₃ followed by T₄, T₂ and T₁.

The result of the present study might be due to availability of fermentable-N from supplemented high protein diet to the bacteria for their growth and multiplication. Similar findings

were also reported in goats [13].

The results of the average protozoal count of the experimental animals under different treatments have been presented in table 8. The average protozoal count of SRL drawn at 45th day of experiment was significantly ($P<0.01$) higher in T₁ followed by T₂, T₄ and T₃ group respectively. Protozoal count was reduced in kids which might be due to feeding of high protein diet [13] and is in agreement with the previous report.

Table 8: Effect of protein supplementation on rumen fermentation pattern of kids

Parameters	Treatment				Overall Mean
	T ₁	T ₂	T ₃	T ₄	
pH	7.13±0.03 ^a	7.19±0.03 ^{ab}	7.29±0.06 ^b	7.23±0.06 ^{ab}	7.21±0.02*
Total Nitrogen (mg/dl)	77.89±0.23 ^a	83.24±0.18 ^b	92.14±0.13 ^d	87.97±0.31 ^c	85.31±1.22**
TCA Precipitable N (mg/dl)	42.14±0.24 ^a	48.80±0.19 ^b	55.55±0.22 ^d	52.19±0.29 ^c	49.67±1.14**
Soluble N (mg/dl)	35.76±0.13 ^b	34.44±0.28 ^a	36.58±0.19 ^c	35.77±0.14 ^b	35.64±0.20**
Microbial Protein (%)	38.93±0.14 ^a	45.26±0.18 ^b	50.94±0.28 ^d	48.76±0.33 ^c	45.97±1.05**
Total Bacterial count (x10 ⁹ /ml SRL)	7.13±0.04 ^a	8.13±0.05 ^b	10.13±0.04 ^d	9.13±0.05 ^c	8.63±0.26**
Total Protozoal count (x10 ⁵ /ml SRL)	3.52±0.04 ^d	3.01±0.04 ^c	2.54±0.03 ^a	2.70±0.04 ^b	2.94±0.09**

(* $P<0.05$, ** $P<0.01$)

6. Economics

The economics of raising parasitized kids with different types of supplements is given in table 9. The total expenditure incurred in different groups was worked out to Rs. 0, 133, 149

and 165 for T₁, T₂, T₃ and T₄ groups, respectively. Net profit for the different groups were Rs. 344, 498, 887 and 646 for T₁, T₂, T₃ and T₄ groups, respectively and the maximum profit was recorded in T₃.

Table 9: Economics of raising kids on different diets affected with gastro intestinal nematodes

Particulars	Treatment			
	T ₁	T ₂	T ₃	T ₄
Total DMI (kg)	27.67	27.77	28.02	24.52
Cost of feed (Rs/kg)	0	22.19	24.79	27.64
Total cost(Rs) (cost of concentrate *total concentrate intake)	0	133.14	148.74	165.84
Total body weight gain (kg)	0.984	1.804	2.960	2.320
Total cost of meat (Rs)*	344.40	631.40	1036.00	812.00
Feeding cost/kg meat (Rs) (total cost of feed/total BW gain)	0	73.80	50.25	71.48
Profit / Loss (Rs) (total cost of meat-total cost of feed)	344.40	498.26	887.26	646.16
Total expenditure (Rs)	0	133.14	148.74	165.84
Net profit / loss (Rs) (profit/loss)	344.40	498.26	887.26	646.16

*Cost of meat was considered at the local market price of Rs 350/kg.

7. Conclusions

The present study revealed that kids infected with natural subclinical nematodiasis gains faster body weight and higher FCE when supplemented with 20% DCP. During the parasitic infestation the nutritional parameters and rumen fermentation pattern were adversely affected which could be overcome by increasing level of protein in the diet. The cost of rearing was most economical in 20% DCP supplemented group. Thus, kids having parasitic infestation would better be reared under semi-intensive system supplemented with medium protein diet.

7. References

1. Knox MR, Torres-Acosta JFJ, Aguilar-Caballero AJ. Exploiting the effect of dietary supplementation of small ruminants on resilience and resistance against gastrointestinal nematodes. *Veterinary Parasitology*. 2006; 139:385-393.
2. Coop RL, Kyriazakis I. Nutrition-parasite interaction. *Veterinary Parasitology*. 1999; 84:187-204.
3. Sanyal PK. Integrated parasite management in ruminants in India. A concept note. *Biological control of gastrointestinal parasites in ruminants using predacious fungi*. FAO Animal Production & Health Paper 141, FAO, Rome, 1998, 54-65.
4. Hoste H, Huby F, Mallet S. *Strongyloides gastrointestinales* des ruminants: consequences physiopathologiques et mecanismes pathogeniques. *Le Point Veterinaire*. 1997; 28:53-59.
5. Brown MD, Poppi DP, Sykes AR. The effect of post ruminal infusion of protein or energy on the pathophysiology of *Trichostrongylus colubriformis* infection and body composition in lambs. *Australian Journal of Agriculture*. 1991; 42:253-267.
6. Van Hountert MFJ, Barger IA, Steel JW. Supplementary feeding and gastrointestinal nematode parasitism in young grazing sheep. *Proceedings of New Zealand Society of Animal Production*. 1996; 56:94-98.
7. Skerman KD, Hillard JJ. *A handbook of studies of helminth parasites of ruminants*. Near East Anim. Hlth. Inst., Iran, FAO, Rome, 1966.
8. AOAC. *Official methods of analysis*. Edn 17, Association of official Analytical Chemists, Verginia, USDA, Washington, D.C, 2000.
9. Talapatra SK, Ray SC, Sen KC. The analysis of mineral constituents in biological materials. Part-I. Estimation of phosphorous, chlorine, calcium, magnesium, sodium and potassium in food stuffs. *Indian journal of Veterinary Science and Animal Husbandry*. 1940; 10:243.
10. Fiske CH, Subbarao Y. The colorimetric determination of

phosphorous. Journal of Biology and Chemistry. 1925; 66:375.

11. Van Soest PJ. Nutritional Ecology of the Ruminants. Edn 2, Ithaca, USA: Cornell University Press, 1994.
12. Poppi DP, MacRae J, Brewer A, Coop RL. Nitrogen transactions in the digestive tract of lambs exposed to the intestinal parasites *Trichostrongylus colubriformis*. British Journal of Nutrition. 1986; 55:593-602.
13. Roy A. Studies on nutrition-parasite interaction in goats with induced gastrointestinal nematodosis fed with medicated urea molasses mineral block. MVSc Thesis, Indira Gandhi Krishi Viswavidyalaya, Raipur, 2007.
14. Rowe JB, Nolan JV, Dechaneet G, Telani F. The effect of haemonchosis and blood loss into the abomasums on digestion in sheep. British Journal of Nutrition. 1988; 59:125-139.
15. Pathak AK, Tiwari SP. Effect of high plane of nutrition on the performance of *Haemonchus contortus* infected kids. Veterinary World. 2013; 6(1):22-26.
16. Abbott EM, Parkins JJ, Holmes PH. The effect of dietary protein on the pathophysiology of acute ovine haemonchosis. Veterinary Parasitology. 1986; 20:275-306.
17. Dutta FU, Nolan JV, Rowe JB, Gray GD. Protein supplementation improves the performance of parasitized sheep fed a straw-based diet. International Journal of Parasitology. 1998; 28:1269-1278.
18. Bunyeth H, Preston TR. Growth performance and parasite infestation of goats given cassava leaf silage, or sun-dried cassava leaves, as supplement to grazing in lowland and upland regions of Cambodia. Livestock Research for Rural Development. 2006; 18(2):1-7.
19. Mhomga LI, Nnadi PA, Chiejina SN, Idika IK, Ngongeh LA. Effect of protein supplementation on weight gain and dressing percentage of West African Dwarf goats experimentally infected with *Haemonchus contortus* and *Trichostrongylus colubriformis*. Global Advanced Research Journal of Agricultural Science. 2012; 1(9):279-287.
20. Preston TR, Leng RA. Matching ruminant production systems with available resources in the tropics and sub-tropics. Armidal: Penambul Books, New South Wales 2350, Australia, 1987, 181-190.
21. MacRae JC, Sharma GAM, Smith JS, Easton JF, Coop RL. Preliminary observations on the effect of *Trichostrongylus columbriformis* infestation on the energy and nitrogen metabolism of lambs. Animal Production. 1979; 28:456.
22. Poppi DP, MacRae JC, Corrigall W. Proceedings of the Nutrition Society. 1981; 40:116A.
23. Viswanathan TV, Fontenot JP. Effects of feeding different protein supplements on digestibility, nitrogen balance and calcium and phosphorus utilization in sheep. Asian-Australian Journal of Animal Science. 2009; 22(5):643-650.
24. Reveron AE, Topps JH, Gelman AL. Mineral metabolism and skeleton development of lambs affected by *Trichostrongylus columbriformis*. Research in Veterinary Science. 1974; 16:310-319.
25. Basabe J, Eiras DF, Romero JR. Nutrition and gastrointestinal parasitism in ruminant production. Archivos de zootecnia. 2009; 58:131-144.
26. Hume ID, Moir RJ, Somers M. Synthesis of microbial protein in the rumen – influence of the level of nitrogen intake. Australian Journal of Agricultural Research.

1970; 21:283-296.

27. Jain N, Tiwari SP, Singh P. Effect of urea molasses mineral granules on rumen fermentation pattern and blood biochemical constituents in goat kids fed sola (*Aeschonomea indica*) grass based diet. Veterinarski Arhiv. 2005; 75(6):521-530.